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10 JULY 1997

REPORT ON
MITOMYCIN RESIDUES
FOLLOWING DECONTAMINATION
OF SUPERGEN FACILITY

Submitted by:

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Director, Product Development

& Analytical Services

SUMMARY

Based on the measured limit of determination for Mitomycin in the wipes, all seventeentested 12cm wipe samples (floor) showed less than .2µg Mitomycin. The three 15cm wipe samples (2 ceiling, 1 floor) were found to have less than .3µg Mitomycin,

INTRODUCTION

This report describes the development of the analytical method and its application to test samples of wipes used to detect any residue of Mitomycin (hence referred to as MMC) in a facility occuppied by SuperGen. The wipes were applied to various surfaces as described in this report. The wiping process and decontamination were performed by DECON Environmental Services Services Inc. of Hayward, California.

The wipes were extracted with a known volume of deionized water and any extracted Mitomycin was measured by high pressure liquid chromatography (HPLC).

The analytical procedure developed and its limit of determination are described.

EXPERIMENTAL

Limit of Determination /Accuracy A.

An accurate volume of a known concentration of MMC was added to a dry blank wipe which was placed in a screw capped culture tube to which was added a sufficient volume of deionized water for a final volume of 5.0 mL. The extraction was performed by repeatedly inverting the tube by hand for 5 minutes or using a Fisher Roto-Rack for 5 minutes. The supernatant water in each tube was analyzed by HPLC for Mitomycin concentration at the following HPLC conditions:

Column:

Dipheny reverse phase, 5 micron, 4.0mm I.D.,

30cm length at 28-35°C

Mobile Phase:

250mL methanol, HPLC grade

5 mL 0.83N acetic acid

1.54 g.ammonium acetate, HPLC grade q.s. HPLC grade water to 1000mL

If HPLC grade materials are used, there is no need to filter through

a 0.45µ filter.

Flow Rate:

1.0 + - 0.1 mL/min.

Injection Volume:

5 microliters

Detector Wavelength: 365nm at 1.0AUFS

Temperature:

30-35⁰C

Peak Measurement: Peak height

The accuracy of the method as determined by the recovery of known amount of MMC is shown by the data in Table I.

<u>TABLE I</u>

SPIKING OF BLANK (DRY) WIPES WITH MITOMYCIN AND RECOVERY RATES

Wipe Diameter	μ g MMC Added ⁽¹⁾	Theoretical MMC in Extract µg/mL	μg/mL MMC Found in Extract	% Recovered
11cm	15µg	3.0µg/mL	3.57μg/mL	119
11cm	7.5µg	1.5µg/mL	1.37µg/mL	91
11cm	1,5µg	0.3μg/mL	0.3µg/mL	100
12cm	7.5µg	1.5μ g/mL	1.28µg/m L	85

Reference:

Lab book, pp. 45-49, 54

(1)

The MMC was added using an appropriate volume of a 15 µg/mL MMC aqueous solution.

B. <u>Limit of Determination</u>

The lowest concentration of MMC in the extract which can be reproducibly measured on separate days is used for calculating the limit of determination of MMC on a wipe. The limit differed for some wipes due to the larger volume of water needed to extact a larger size wipe. The lowest concentration was experimentally measured by diluting a known concentration of MMC in water and measuring the signal obtained on the chromatograph. This value was found to be 0.03µg/mL. The chromatographic response obtained using the HPLC conditions described above is shown in Figure 1. To calculate the limit of determination for each wipe, the volume of water used to extract each wipe is multiplied by the lowest concentration measurable.

C. Analysis of Wipes Used to Contact Decontaminated Surfaces

Upon receipt at Custom Testing & Development, the samples were placed in a freezer. The wipes are described as round laboratory filter papers of various sizes. Samples were removed from the freezer and each wipe weighed to determine the amount of water on the wetted test samples. The volume of water was calculated for each wipe by subracting the weight of a dry wipe of the same size as the wet wipe from the weight of the wet wipe. The difference, in grams, was taken as the volume in mL on each wipe. The wipes were placed in a screw cap culture tube. A sufficient volume of water was added to each test sample to bring the total volume in contact with each wipe as shown in Table II under "Extraction Volume." The tubes were shaken by hand for 5 minutes or placed on a Fisher Roto-Rack for 5 minutes. Each sample was chromatographed within one hour of addition of water used for extraction. Chromatograms of each sample reported in Table II are labelled as Figures 2 through 23.

D. <u>Discussion</u>:

The chromatographic conditions and extraction procedure demonsrates the ability of the method to accurately recover Mitomycin from the test wipes as shown in Table I.

The sensitivity of the method, 0.2 µg MMC for 12cm diameter wipes and 0.3µg MMC for 15cm diameter wipes has been calculated based on the ability to measure the concentration in the extraction volume shown in Figure I. These values are lower than those reported in the summary report dated 19 June 1997. Those values were based on a quick conservative evaluation whereas the values reported in this report can be confirmed by a more rigorous review of the experimental method and data analysis.

The data obtained on all test wipes reported in Table II indicate no detectable levels of MMC on any of the test wipes examined. The limit of determination, also referred to as the sensitivity of the method, is reported as 0.2µg MMC for 12cm diameter wipes and 0.3µg MMC for 15cm diameter wipes.

The significance of the levels found and the level of detection as it applies to equipment and facility cleaning in a manufacturing facility is discussed in an articles by K.M. Jenkins and A. J. Vanderwielen entitled "Cleaning Validation: An Overall Perspective," in *Pharmaceutical Technology* p 60-73, (1994).

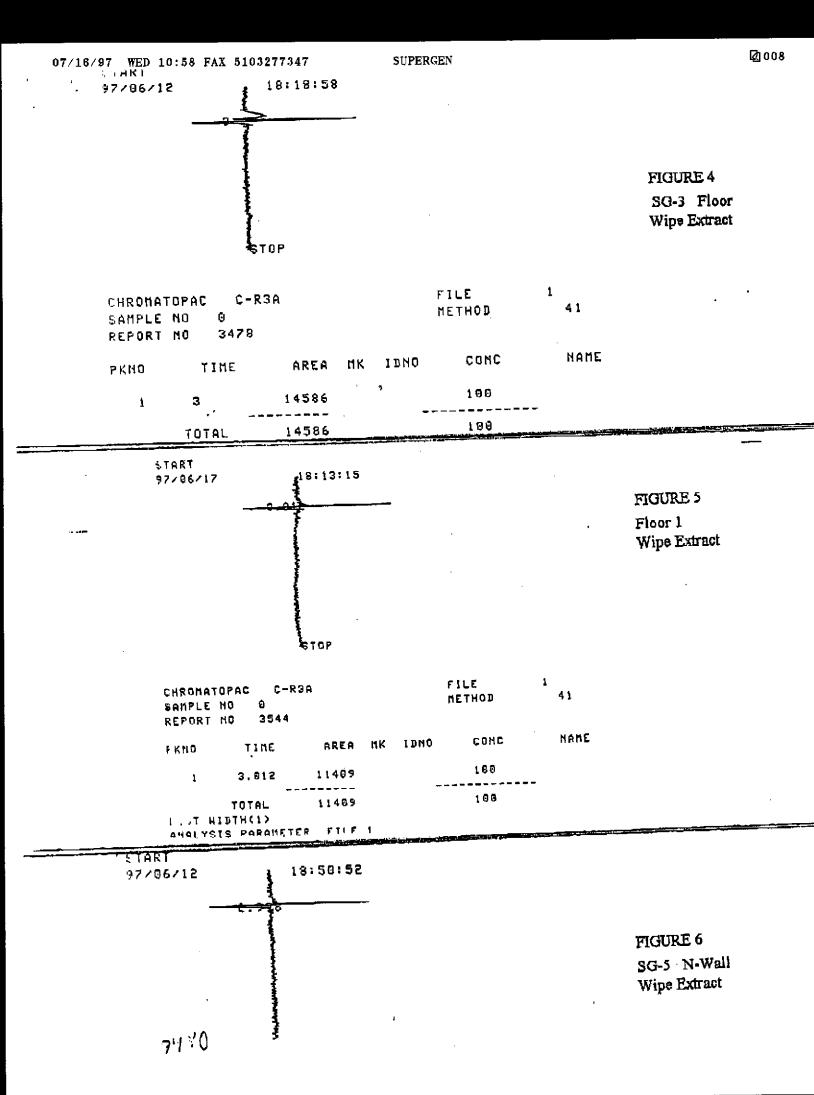
TABLE II

MITOMYCIN CONTENT OF WIPES USED ON VARIOUS SURFACES
AFTER DECONTAMINATION OF SUPERGEN FACILITY

Sample#	Wipe	Location	Extraction	µg/wipe
A control of the cont	diameter		volume	
Ceiling 1	15cm	Ceiling	9.0mL	<0,3μg
Ceiling 2	15cm	Ceiling	9,0mL	<0.3µg
SG-3	12cm	Floor	6.0mL	<0,2μg
Floor 1 .	15cm	Floor	9,0mL	<0.3μg
SG-5	12cm	N-Wall	6.0mL	<0.2µg
SG-6	12cm	N-Wall	6.0mL	<0.2µg
SG-7	12cm	S-Wall	6.0mL	<0.2µg
\$G-8	12cm	S-Wall	6.0mL	<0.2µg
SG-9	12cm	E-Wall	6.0 m L	<0.2µg
SG-10	12cm	E-Wall	6.0mL	<0.2μg
SG-11	12cm	W-Wall	6.0mL	<0.2µg
SG-12	12cm	W-Wali	6.0mL	<0.2µg
SG-13	12cm	Countertop	6.0mL	<0.2μg
SG-14	12cm	Countertop	6.0mL	<0.2μg
SG-15	12cm	Inside L-Cabinet	6.0mL	<0.2μg
SG-16	12cm	Wet Blsnk	6.0mL	<0.2μg
SG-17	12cm	Wet Blank	6.0mL	<0.2µg
SG-18	12cm	Wet Blank	6.0mL	<0.2μg
SG-19	12cm	Wet Blank	6.0mL	Sample lost
SG-20	12cm	Dry Blank	6.0mL	<0.2μg
SG-21	12cm	Dry Blank	6.0mL	<0.2µg
SG-22	12cm	Dry Blank	6.0mL	not tested
SG-23	12cm	Dry Blank	6.0mL	Used for recovery study

CHROMATOPAC C-R3A SAMPLE NO. REPORT NO 3542

FILE METHOD



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METHOD

3483

SAMPLE NO

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FILE 1

C-R3A 2486

CHROMATOPAC

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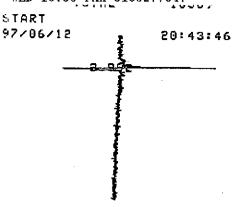


FIGURE 13 SG-12 W-Wall Wipe Extract

41

HAME

CHROMATOPAC C-R3A FILE SAMPLE NO METHOD REPORT NO 3487 PKHO TIME AREA IDNO CONC

2.992 10278 100

TOTAL 10278 100

TART

97/06/12

20:59:39

FIGURE 14 SG-13 Countertop Wipe Extract

CHROMATOPAC C-R3A FILE SAMPLE NO nETHOD 41 REPORT NO 3.488

PKHQ TIME AREA MK IDNO CONC NAME

2.995 10255 100

> TOTAL 10255 100

ETART

97/86/12 21:15:28

> FIGURE 15 SG-14 Countertop

Wipe Extract

CHROMATOPAC C-R3A SAMPLE NO

3489.

FILE METHAR

FILE CHROMATOPAC C-R3A KETHOD SAMPLE NO B REPORT NO 3476 NAME CONC AREA MK IDNO TIME PKNO 100 10291 100 10291 TOTAL

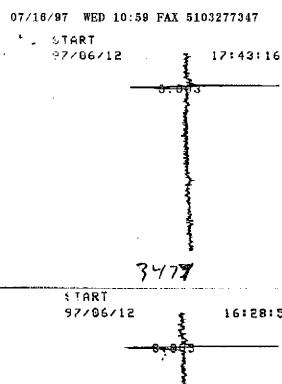


FIGURE 19 SG-18 Wet Blank Wipe Extract

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\$ TART 97/06/12 16:28:52

FIGURE 20 SG-20 Dry Blank Wipe Extract

CHROMATOPAC C-R3A FILE SAMPLE NO G METHOD 41 REPORT NO 3473 PKNO TIME AREA MK IDNO CONC NAME 1 - 3.003 10474 100 TOTAL 18474 100 START 97/06/12 16:48:07

FIGURE 21 SG-21 Dry Blank Wipe Extract

CHROMATOPAC C-R3A SAMPLE NO 0 REPORT NO 3474 FILE 1 METHOD 41