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2771 Hopyard Road Pleasanton, California 94612

FIRST QUARTER 2009 GROUNDWATER AND SVE MONITORING REPORT

HOPYARD CLEANERS

2771 Hopyard Road Pleasanton, California Self- Monitoring Program No. R2-2006-0059

Prepared by



engineers | scientists | innovators

475 14th Street, Suite 400 Oakland, California 94612

Project Number: WR0574

30 April 2009

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Geosyntec Consultants, Inc. 475 14th Street, Suite 400 Oakland, California 94612

Melissa Asher, P.E.

Engineer

Hanchih (Angela) Liang, Ph.D., P.E.

Senior Engineer

Project Number: WR0574

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1. INTRODUCTION

On behalf of the property owner, Ms. Clare Leung, Geosyntec Consultants (Geosyntec) prepared this first quarter 2009 groundwater and soil vapor extraction (SVE) monitoring report for Hopyard Cleaners located at 2771 Hopyard Road, in Pleasanton, California (the "Site"). A Site location map is provided in Figure 1. The work described in this report was performed in compliance with the California Regional Water Quality Control Board, San Francisco Bay Region (RWQCB) Order No. R2-2008-0032, issued on 29 May 2008.

1.1 Monitoring Well Network

The Site monitoring well network consists of seven wells (MW-1 through MW-7). For discussion purposes, the uppermost groundwater zone beneath the Site, which occurs from approximately 20 to 35 feet below ground surface (feet bgs), is referred to as the A Zone, and the deeper groundwater from approximately 40 to 60 feet bgs is referred to as the B Zone. Wells MW-1 through MW-4 are screened in the A Zone, and wells MW-5 through MW-7 are screened in the B Zone. B Zone wells MW-6 and MW-7 were installed in August 2008 and were monitored for the first time in the fourth quarter of 2008. Well completion details are summarized in Table 1. Well locations relative to the Site are shown on Figure 2.

1.2 SVE System

The SVE system was installed at the Site in August 2008 with five SVE wells (SVE-1 through SVE-5) located inside Hopyard Cleaners. Geosyntec conducted a pilot test of the SVE system on 19 and 21 August 2008. The SVE system installation, pilot test, and start-up were documented in the SVE System Installation and Pilot Test Report, which was submitted to the RWQCB on 29 September 2008. The full-scale SVE operations began on 21 August 2008. An Addendum to the SVE System Installation and Pilot Test Report, which included quarterly SVE influent volatile organic compounds (VOC) analysis and recommendations and conclusions, was submitted to the RWQCB on 1 December 2008. The SVE System Installation and Pilot Test Report and the Addendum to the SVE System Installation and Pilot Test Report was approved by the RWQCB on 9 December 2008. The SVE system layout is shown on Figure 2.



1.3 Work Performed This Quarter (First Quarter 2009)

The following work was performed in the first quarter of 2009:

- The first quarter groundwater monitoring event was performed on 26 January 2009 using passive diffusion bag (PDB) samplers. This work is discussed in detail in this report.
- SVE monitoring was conducted on 6 January, 21 January, 5 February, 19 February, 26 February, and 12 March 2009. This work is also discussed in detail in this report.
- On 26 January 2009, additional soil oxidant demand (SOD) soil samples were collected as detailed in the *Remedial Action Plan Addendum: Comprehensive Feasibility Study for ISCO & EISB* dated 24 November 2008. Results of the additional SOD sampling were submitted to the RWQCB on 16 March 2009 in a report titled *Results of Additional Soil Oxidant Demand Characterization*.
- On 26 January 2009, soil and groundwater samples were also collected for the enhanced in situ bioremediation (EISB) feasibility study. The EISB treatability study commenced on 9 February 2009 and is scheduled to be completed in by August 2009. Results of the groundwater Gene-Trac and anion analyses and preliminary treatability study results are presented in Section 5 of this report.

2. QUARTERLY GROUNDWATER MONITORING

Quarterly groundwater monitoring was performed at the Site on 26 January 2009. PDBs were used to collect samples from MW-1 through MW-7. A study to test the appropriateness of using PDBs was proposed in the *Results of Fourth Quarter 2007 Groundwater Monitoring* report submitted to the RWQCB on 31 January 2008¹ and was verbally approved by the RWQCB in a conference call on 12 March 2008. The PDB study was completed in the first and second quarters 2008. In general, the results of the study showed that cis-1,2-dichloroethene (cis-1,2-DCE), tetrachloroethene (PCE), and trichloroethene (TCE) concentrations were slightly higher in samples collected from PDBs compared to samples collected using a submersible pump. Sample results reported as non-detect using the conventional sampling method were also non-detect using the PDB sampling method. These results indicate that PDB samplers are an appropriate and reliable method of monitoring VOCs at this Site. Therefore, PDBs have replaced sampling via peristaltic pump starting in the third quarter 2008.

2.1 <u>Sampling and Analytical Procedures</u>

The groundwater sampling fieldwork was performed by Environmental Sampling Services, Inc. (ESS), of Martinez, California. ESS's report, including field procedures and sampling logs, is provided in Appendix A.

The PDBs were deployed on 10 December 2008, during the fourth quarter 2008 monitoring event, in monitoring wells MW-1 through MW-5. A stratification study was conducted on MW-6 and MW-7 during the fourth quarter 2008. Results of the stratification study indicated that stratification did not occur in either well, and PDBs were deployed in the middle of the well screens on 6 January 2009 in MW-6 and MW-7.

On 26 January 2009, the PDBs were removed from the wells and sampled. Samples were hand-delivered to Test America of Pleasanton, California, for analysis.

¹ Geosyntec Consultants, 2008. Results of the Fourth Quarter 2007 Groundwater Monitoring, Hopyard Cleaners, 2771 Hopyard Road, Pleasanton, California, Self-Monitoring Program No. R2-2006-0059, 31 January 2008.



Groundwater samples from the Site monitoring wells were analyzed for VOCs by EPA Method 8260B. New PDBs for the second quarter 2009 sampling event were deployed in wells MW-1 through MW-7 on 26 January 2009 after the first quarter 2009 monitoring event was completed.

2.2 **Groundwater Elevations and Flow Conditions**

Table 2 summarizes groundwater elevations measured during this and previous sampling events. During the first quarter 2009, groundwater in the A Zone (MW-1 through MW-4) beneath the Site was encountered between 16.17 and 17.86 feet bgs, which is between 308.41 and 309.52 feet above Mean Sea Level (MSL). Groundwater in the B Zone was encountered between 28.10 and 30.61 feet bgs, which corresponds to groundwater elevations ranging from 296.36 to 296.58 feet MSL.

Water levels measured during the first quarter 2009 event were used to construct groundwater elevation contours for the A Zone and B Zone, as shown in Figure 3 and 4, respectively. Table 3 summarizes groundwater gradients and flow directions for this and previous monitoring events. The first quarter 2009 A Zone groundwater contours indicate a general groundwater flow to the west-northwest with an average gradient of 0.0045 feet per feet (ft/ft) (23.8 feet per mile (ft/mi)). This gradient and flow direction is consistent with previous monitoring events, as shown on Table 3. The B Zone groundwater contours indicate general groundwater flow to the southwest under a gradient of approximately 0.0012 ft/ft (6.4 ft/mi), consistent with the fourth quarter 2008 monitoring event.

Groundwater elevations over time are shown on Figure 5. Groundwater in the A Zone monitoring wells has fluctuated from 308.8 to 314.6 feet MSL since monitoring began in November 2006. Groundwater elevations in the B Zone are lower than those measured in the A Zone, with elevations ranging from 293.3 to 307.4 feet MSL. Both the A Zone and B Zone groundwater elevations tend to fluctuate seasonally with higher elevations during the winter and spring and lower elevations in the summary and fall.

2.3 Data QA/QC

Geosyntec performed a quality assurance/quality control (QA/QC) review of the analytical data. Data were reviewed for completeness, accuracy, precision, sample



contamination, conformance with holding times, and detection limits within acceptable ranges. Based on this review, the data are acceptable.

2.4 Analytical Results

Laboratory analytical reports are provided in Appendix B. Table 4 summarizes analytical results for groundwater samples collected during the first quarter 2009 event together with historical results. Analytical results for the current sampling event are also shown on Figures 3 and 4 for the A Zone and B Zone, respectively. Isoconcentration contour maps for PCE and TCE are shown on Figures 6 through 8. The isoconcentration contours were drawn using current data from monitoring wells along with results from grab groundwater samples previously collected at the Site. Results are summarized by zone, as follows:

2.4.1 A Zone Wells: MW-1 through MW-4

Analytical results for samples taken from the four A Zone monitoring wells show the highest VOC concentrations at MW-2. During the first quarter 2009, the PCE concentration in both the original and duplicate samples collected from MW-2 was 12,000 micrograms per liter (μ g/L). The 12,000 μ g/L result is within the range of historical PCE concentrations at MW-2, which have ranged from 4,700 to 15,000 μ g/L. VOC concentrations observed during the first quarter 2009 in the other A Zone wells (MW-1, MW-3, and MW-4) were consistent with historical results.

2.4.2 B Zone Wells: MW-5 through MW-7

PCE is the only VOC detected in the B Zone groundwater. The highest detection of PCE was at the closest B Zone monitoring well to the Site, MW-5, and was at a concentration of 37 μ g/L. Farther downgradient of the Site, PCE was detected at MW-7 at 9.9 μ g/L and was not detected at MW-6.

2.5 Results Discussion

A time series of PCE and TCE concentration in monitoring wells that have been monitored for more than four quarters are shown on Figure 9. The highest concentrations of PCE and TCE have historically been detected in A Zone monitoring well MW-2, which shows a slight increasing trend over the past year. PCE and TCE



concentrations in the other A Zone wells MW-1, MW-3, and MW-4, and in B Zone well MW-5 are stable or declining.

As shown on Figure 10, concentrations in MW-2 tend to fluctuate inversely with groundwater elevations measured in this well. The highest PCE concentration of $15,000 \,\mu\text{g/L}$ corresponded to the lowest groundwater elevation observed at this well.

3. SVE SYSTEM MONITORING AND PERFORMANCE EVALUATION

The SVE system was installed at the Site in August 2008 with five SVE wells (SVE-1 through SVE-5) located inside Hopyard Cleaners. The full-scale SVE operations began on 21 August 2008. Startup monitoring of the SVE system was performed on day 1 through 5, day 7, and day 9 of system startup to evaluate system performance and air emissions for the Bay Area Air Quality Management District Permit to Operate (BAAQMD PTO). Monitoring was performed weekly for the first month and monthly, at a minimum, thereafter. During the first quarter 2009, Geosyntec conducted the system monitoring on 6 January, 21 January, 5 February, 19 February, 26 February, and 12 March 2009. The SVE system layout is shown on Figure 2. The SVE well locations and piping layout inside the dry cleaners is shown on Figure 11.

3.1 **SVE Monitoring Procedures**

SVE monitoring includes the following procedures:

- Perform photoionization detector (PID) screening via Tedlar® bags of:
 - o Samples collected from the system influent, mid-point between the two granular activated carbon (GAC) vessels, and the system effluent, and
 - o Samples collected at each SVE wellhead.
- Record vacuum response at each SVE wellhead;
- Record flow rate and vacuum response at the manifold;
- Record vacuum, temperature, and flow rate readings at system influent;
- Record hour meter;
- Inspect the moisture separator water level and drain into 55-gallon drums, if necessary; and
- Record the electrical meter reading.

As discussed in the SVE System Installation and Pilot Test Report and subsequent Addendum SVE System Installation and Pilot Test Report, influent SVE samples were collected in 1-liter Summa canisters for laboratory analysis by TO-15 during start-up testing and on a quarterly basis to correlate VOC concentrations with PID readings.

3.2 SVE Operation, Monitoring, and Maintenance

The system performance monitoring results are presented in Table 5. The laboratory analytical results for the SVE influent samples are summarized in Table 6 and the laboratory analytical reports are provided in Appendix B. The individual SVE well monitoring results are shown in Table 7.

The system was operated continuously 24 hours a day from the startup on 21 August 2008 through 2 September 2009, except for an approximately 2-hour time period on 29 August 2008 when the blower shut-off switch was tripped. Even though measures were taken to reduce the noise from the system blower, Geosyntec received complaints regarding the noise at night from residents in the vicinity of the dry cleaners (both across Hopyard Road and Valley Road). Therefore, on 3 September 2008, the SVE system was modified to run 14 hours a day from 8 am to 10 pm.

The PID readings measured from the wellheads after the startup and during the fourth quarter 2008 indicated that SVE-3 and SVE-5 were not as productive as SVE-1, SVE-2, and SVE-4, as shown in Table 7. In order to optimize the SVE system performance and efficiency, cycling of the SVE wells was started on 6 January 2009. The following operations and maintenance activities were performed during the first quarter 2009:

- 6 January 2009 the low-yield SVE wells (SVE-3 and SVE-5) were turned off.
- 21 January 2009 SVE system monitoring was conducted before SVE-3 and SVE-5 were turned back on and the other three wells (SVE-1, SVE-2, and SVE-4) were turned off. SVE system monitoring was conducted again approximately 30 minutes after wells were cycled, as shown on Table 5.
- **5 February 2009** SVE system monitoring was conducted before SVE-3 and SVE-5 were turned off and SVE-1, SVE-2, and SVE-4 were turned back on. SVE system monitoring was conducted again approximately 30 minutes after wells were cycled.
- 19 February 2009 SVE system monitoring was conducted before SVE-3 and SVE-5 were turned back on and the other three wells (SVE-1, SVE-2, and SVE-4) were turned off. SVE system monitoring was conducted again approximately 30 minutes after wells were cycled.
- **26 February 2009 to End of First Quarter 2009** SVE system monitoring was conducted on 26 February 2009. Based on influent SVE concentrations

 $(0 \mu g/m^3 \text{ as PCE})$ on 19 and 26 February 2009, all five SVE wells were then turned on for only 2 hours per day (8 am to 10 am). On 12 March 2009, system monitoring was conducted again, and the influent SVE concentration was measured at 1,097 $\mu g/m^3$.

Due to the anomalously high mid-point and effluent concentrations observed during the 5 February 2009 SVE system monitoring, the blower was moved in front of the carbon vessels in the treatment process by Mako on 19 February 2009. The blower increases the temperature of the extracted vapor and therefore decreases the amount of water vapor that condenses in the carbon vessels, resulting in increased efficiency of the carbon. Since the carbon must remain below 120 degrees Fahrenheit, the recirculation valve on the system was opened to control the temperature. Monitoring of the temperature immediately before the carbon vessels was added to the system monitoring program, as shown in Table 5.

3.3 **SVE Performance Evaluation**

Influent concentrations of VOCs into the SVE system in the first quarter 2009 have ranged from 0 to 4,830 micrograms per cubic meter ($\mu g/m^3$) as equivalent PCE (0.0 to 0.7 parts per million by volume [ppmv]) with the highest concentrations detected at the beginning of cycling the SVE wells (Table 5 and Figure 12). During the first quarter 2009, the average influent VOCs concentration was 1,345 $\mu g/m^3$ as equivalent PCE (0.240 ppmv). After seven months of operations, the SVE system has removed a total of approximately 9.4 pounds (lbs) (0.69 gallons) of VOCs as equivalent PCE (Table 5 and Figure 13).

The laboratory analytical results indicate that PCE is the primary COC being removed from the target remediation zone, as shown on Table 6 and in the laboratory analytical report provided in Appendix B. The PID reading of the sample collected on 21 August 2008 was significantly higher than the analytical result due to the time lapse between the PID measurement and sample collection. The laboratory samples collected on 2 September 2008, 5 December 2008, and 12 March 2009 were collected right after the PID measurements and the analytical results indicate similar VOC concentrations.

Figure 14 presents the time concentration plots of PCE and TCE influent concentrations at the SVE system. The PCE concentrations decreased from 24,000 μ g/m³ on 21 August 2008, to 8,500 μ g/m³ on 2 September 2008, to 2,300 μ g/m³ on 5 December 2008, and to 1,700 μ g/m³ on 12 March 2009, approaching the soil gas clean μ g standard of 1,400

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 $\mu g/m^3$. Similarly, TCE concentrations have remained below the soil gas cleanup standard of 4,100 $\mu g/m^3$ and have decreased from 280 $\mu g/m^3$ on 21 August 2008 to 160 $\mu g/m^3$ on 2 September 2008, to 64 $\mu g/m^3$ on 5 December 2008, and to 36 $\mu g/m^3$ on 12 March 2009. The decreasing trend of PCE and TCE concentrations shows that the SVE system has been effective at removing the COCs from the subsurface.

Table 7 presents the PID screening results of the SVE wells. Cycling of the wells demonstrates the potential rebound effects. However, after two months of cycling, the rebound effects have significantly decreased. As of 12 March 2009, all wellhead PID screening results are below the soil gas cleanup standards for PCE $(1,400 \, \mu g/m^3)$ except for SVE-2 $(1,780 \, \mu g/m^3)$.

The SVE system operation time was increased to 4 hours a day (8 am to 12 pm) on 10 April 2009 with all five SVE wells turned on. The system will be operated in this configuration during the second quarter 2009 and the system performance will be evaluated and reported in the second quarter 2009 monitoring report.

4. SOD RESULTS SUMMARY

In order to evaluate in situ chemical oxidation (ISCO) as a remedial option for the Site, the soil oxidant demand (SOD) at the Site was evaluated to estimate the amount of permanganate that would be required. The Soil Oxidant Demand Characterization Work Plan (Work Plan) was submitted to the RWQCB on 30 June 2008 and approved verbally by the RWQCB on 13 August 2008 during a teleconference and in a letter on 22 August 2008. Geosyntec collected soil samples for SOD analyses in August 2008, and the results were reported in the *Remedial Action Plan Addendum: Comprehensive Feasibility Study for ISCO & EISB* (RAP Addendum) dated 24 November 2009. The results indicated that the ISCO with MNA remedy may not be the most effective remedy to treat the groundwater due to the high SOD detected at the Site.

To confirm the SOD results, additional SOD characterization was proposed, along with an EISB feasibility study, in the RAP Addendum. Additional characterization included collecting samples from two additional locations at three depths. The samples were collected on 26 January 2009, and the results are discussed below.

4.1 SOD Results

Sampling procedures and analytical methods were detailed in the *Results of Additional Soil Oxidant Demand Characterization* submitted to the RWQCB on 16 March 2009. Results are shown on Table 8 and are summarized as follows:

- Soil properties for the samples collected near MW-2 (SOD-1) are consistent between the three sample depths with an average bulk density of 96.7 pounds per cubic feet (lb/ft3) and an average porosity of 44.3%.
- In the vicinity of MW-2, the average foc at SOD-2 is consistent with the average foc at SOD-1. The average focs were 1.07% and 0.99%, respectively.
- The average foc at SOD-3, near MW-1, was 0.66%, which is lower than the foc near MW-2.

In the vicinity of MW-2, the permanganate SOD ranged from 5.19 to 23.92 grams of permanganate per kilogram of soil (g/kg). The highest oxidant demand between was



observed from 21 to 22 ft bgs at 23.92 g/kg. This corresponds to a potassium permanganate demand of 31.78 g/kg.

In the vicinity of MW-1, the SOD at each depth was similar and overall lower than the SODs observed near MW-2. The permanganate SOD at SOD-3 ranged from 4.27 to 5.59 g/kg, which corresponds to a potassium permanganate SOD of 5.67 to 7.43 g/kg.

4.2 Permanganate Requirements for A Zone Groundwater Remediation

Table 9 presents the estimated amount of permanganate, along with estimated injection volumes and times, required to treat shallow groundwater, using the soil analytical results and PCE and TCE concentrations detected at the Site. Because of the high reactivity and short reaction time of the oxidants, an ample amount of oxidant is needed to, not only meet the soil oxidant demand, but to react with the COCs. The bold columns on Table 9 highlight a most conservative ISCO design needed to achieve oxidation of the contaminants at the Site.

Due to the estimated high permanganate requirements to treat A Zone groundwater at the Site, the SOD characterization shows that ISCO with monitored natural attenuation (MNA) may not be the most effective remedy. ISCO with MNA will be evaluated in comparison to EISB with MNA at that time and will be reported along with the EISB with MNA evaluation to the RWQCB by 30 September 2009.

5. EISB FEASIBILITY STUDY PRELIMINARY RESULTS

As detailed in the *Remedial Action Plan Addendum: Comprehensive Feasibility Study* for ISCO & EISB dated 24 November 2008, soil and groundwater samples were collected on 26 January 2009 for the enhanced in situ bioremediation (EISB) feasibility study in order to conduct the following:

- (1) A groundwater geochemistry and microbiology evaluation of the Site's ability to support reductive dechlorination; and
- (2) A treatability study to assess the potential for EISB treatment of VOCs in A Zone groundwater at the Site.

Sample collection, analytical methods, and preliminary results are summarized in the following sections.

5.1 Background on Reductive Dechlorination

The dominant PCE biodegradation mechanism in groundwater environments is reductive dechlorination, which involves the sequential replacement of chlorine atoms on the alkene molecule with hydrogen atoms. Under reducing conditions, PCE serves as an electron acceptor and is dechlorinated via TCE, to cis-1,2-DCE, to vinyl chloride (VC) and then to ethene. Hydrogen, typically produced during the bacterial metabolism of simple organic carbon compounds such as alcohols, organic acids, sugars or edible oils, serves as the electron donor in the dechlorination reactions.

5.2 **Groundwater Sample Collection and Analyses**

ESS collected groundwater samples using a peristaltic pump, and Enprob Environmental Probing of Oroville, California, collected soil samples using a direct push drilling rig. Groundwater samplescollected from MW-1, MW-2, and MW-3 were analyzed for the following:

- VOCs by EPA Method 8260B by Test America;
- Anions, including nitrate, nitrite, sulfate, and chloride by EPA Method 300.0 by Test America; and



• Gene-Trac Testing for *Dehalococcoides* and vinyl-chloride reductase by SiREM Laboratory of Guelph, Ontario, Canada (SiREM).

5.3 EISB Treatability Study Procedures

The EISB treatability study is being conducted by SiREM. The study includes anaerobic treatments constructed to estimate the rate and extent of reductive dechlorination achievable at the Site by indigenous microbial populations under existing Site conditions (intrinsic control) and through the addition of two electron donors (biostimulation). The impact of biostimulation and the additional dechlorination microorganisms (bioaugmentation) is assessed through the addition of a commercial bioaugmentation culture (KB-1[®]).

Using the soil samples collected near MW-2 and groundwater samples collected from MW-2, six treatment and control microcosms were constructed by filling 250-milliter (mL) glass bottles with approximately 150 to 200 mL of Site groundwater and 60 grams of Site soil, leaving nominal headspace for gas production (i.e. ethene, carbon dioxide, and methane). The microcosms were constructed in a disposable anaerobic glove bag and will be stored in an anaerobic chamber. One replicate of each treatment was amended with resazurin, which turns pink when exposed to oxygen, in order to monitor redox conditions. The six treatment/control microcosms are as follows:

- One anaerobic sterile control, which was amended with mercuric chloride and sodium azide;
- One anaerobic intrinsic control, which was unamended and used to measure intrinsic biodegradation;
- Two electron donor treatments, which were amended with two different electron donors (one amended with lactate and one with a commercial emulsified vegetable oil (EVO) called Newman Zone®); and
- Two electron donor and bioaugmented treatments, which were amended with the two electron donors (one with lactate and one with EVO) and KB-1[®] to assess the ability of these bacteria to promote or accelerate complete dechlorination.

The incubation of the treatability study microcosms started on 9 February 2009. Samples were collected from the control and treatment microcosms and were analyzed for VOCs and dissolved hydrocarbon gases (i.e. ethene, ethane, and methane). Samples from the electron donor amended microcosms were also analyzed for volatile fatty acids analyses (i.e. lactate, acetate, and propionate). Other analyses include the measurement of pH and anions.

5.4 **Preliminary Results**

Results of the Gene-Trac analyses and anion analysis are summarized in Table 10 and the laboratory analytical reports are included in Appendix C. Preliminary treatability study results are shown on Figure 15.

5.4.1 Current Site Conditions

The detections of TCE and cis-1,2-DCE in samples collected from groundwater monitoring wells at the Site are at relatively low concentrations compared to PCE concentrations, as shown on Table 4. This implies that some limited intrinsic reductive dechlorination is occurring at the Site. The lack of vinyl chloride detections at the Site indicate that reductive dechlorination may be stalling at cis-1,2-DCE.

As shown in Table 10, the high oxidation reduction potential (ORP) (63.5 to 150.6 millivolts), low dissolved oxygen (DO) concentrations (0.12 to 0.24 mg/L), and the presence of nitrate and sulfate in groundwater indicate that the Site A Zone groundwater is currently not under the strong reducing conditions.

Gene-Trac testing for microbial populations native to the Site show that *Dehalococcoides* species are detected at a moderate concentrations which may, or may not, be associated with observable dechlorination. Vinyl chloride reductase gene, which is capable of producing enzymes that break down vinyl chloride, was detected at extremely low concentrations at or below the sample specific quantitation limit. The results on the Gene-Trac testing explains the stall in reductive dechlorination at cis-1,2-DCE under the current Site conditions.

5.4.2 Treatability Study

The treatability study was started on 9 February 2009. Samples were taken on 9 February (Day 0), 23 February (Day 14), and 9 March 2009 (Day 28) from the anaerobic sterile control microcosm and the anaerobic intrinsic control microcosms. As

shown on Figure 15, on Day 28, the PCE and TCE concentrations in the anaerobic sterile control microcosm decreased by 9 percent and 16 percent, respectively. The PCE concentration in the anaerobic intrinsic control microcosm decreased 37 percent and the TCE concentrations increased 15 percent, indicating some intrinsic biodegradation.

In the two electron donor treatment studies, samples were taken on 9 February (Day 0), 23 February (Day 14), 9 March (Day 28), and 23 March 2009 (Day 42). As shown on Figure 14, dechlorination of PCE and TCE to cis-1,2,-DCE was rapid following amendment with both electron donors, but stalled at cis-1,2-DCE with little or no production of vinyl chloride or ethene.

In the bioaugmented treatability studies, samples were taken on 9 February (Day 0), 23 February (Day 14), 9 March (Day 28), 16 March (Day 35), 20 March (Day 39), 23 March (Day 42), 30 March 2009 (Day 49). As shown on Figure 15, complete dechlorination of PCE and TCE to ethene occurred in microcosms containing donor amendments (EVO and lactate) bioaugmented with KB-1[®]. Complete dechlorination occurred quickly in both bioaugmented microcosms: within 14 days in the Newman Zone® & KB-1® microcosms and within 21 days in the lactate & KB-1® microcosms. Low, transient concentrations of vinyl chloride were detected in both microcosms, with less vinyl chloride detected with EVO.

5.4.3 Discussion

The preliminary treatability results indicate that the Site is amenable to EISB with bioaugmentation. Concentrations of non-VOC electron acceptors (i.e. nitrate and sulfate) at the Site are low to moderate, and in the treatability study reducing conditions were quickly achieved as evident by the decreasing PCE concentrations in the beginning of the study. These results indicate that Site geochemistry is compatible with PCE reductive dechlorination, since other electron acceptors, such as nitrate and sulfate, are reduced quickly and large quantities of donor are not required to achieve reducing conditions optimal for PCE dechlorination.

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These results will be used along with the final results of the treatability study, which will be completed by mid-August 2009, to evaluate EISB as a remedial alternative for the Site. Results of this evaluation will be submitted to the RWQCB by 30 September 2009, as indicated in the "Approval of *Remedial Action Plan Addendum: Comprehensive Feasibility Study for ISCO & EISB*, and Requirement for Report for 2771 Hopyard Road, Pleasanton, Alameda County" letter from the RWQCB dated 16 December 2008.

6. FUTURE WORK

The following work will be completed during the second quarter 2009:

- The *Revised Remedial Action Plan*, including a human health risk assessment will be submitted to the RWQCB during the second quarter 2009.
- The next quarterly groundwater monitoring event will be performed in April 2009. Results of the second quarter 2009 monitoring report will be submitted to the RWQCB by 31 July 2009.
- SVE monitoring will continue on a monthly basis at a minimum with one sample being collected for TO-15 analysis during the second quarter 2009. Results of the monitoring will be presented in the second quarter 2009 monitoring report due to the RWQCB on 31 July 2009.
- Based on the SOD confirmation sampling results, a focused ISCO application at the Site is being evaluated and the evaluation will be presented in the second quarter 2009 monitoring report will be submitted to the RWQCB by 31 July 2009.
- The EISB treatability study will continue and the results will be reported in the second quarter 2009 monitoring report will be submitted to the RWQCB by 31 July 2009.

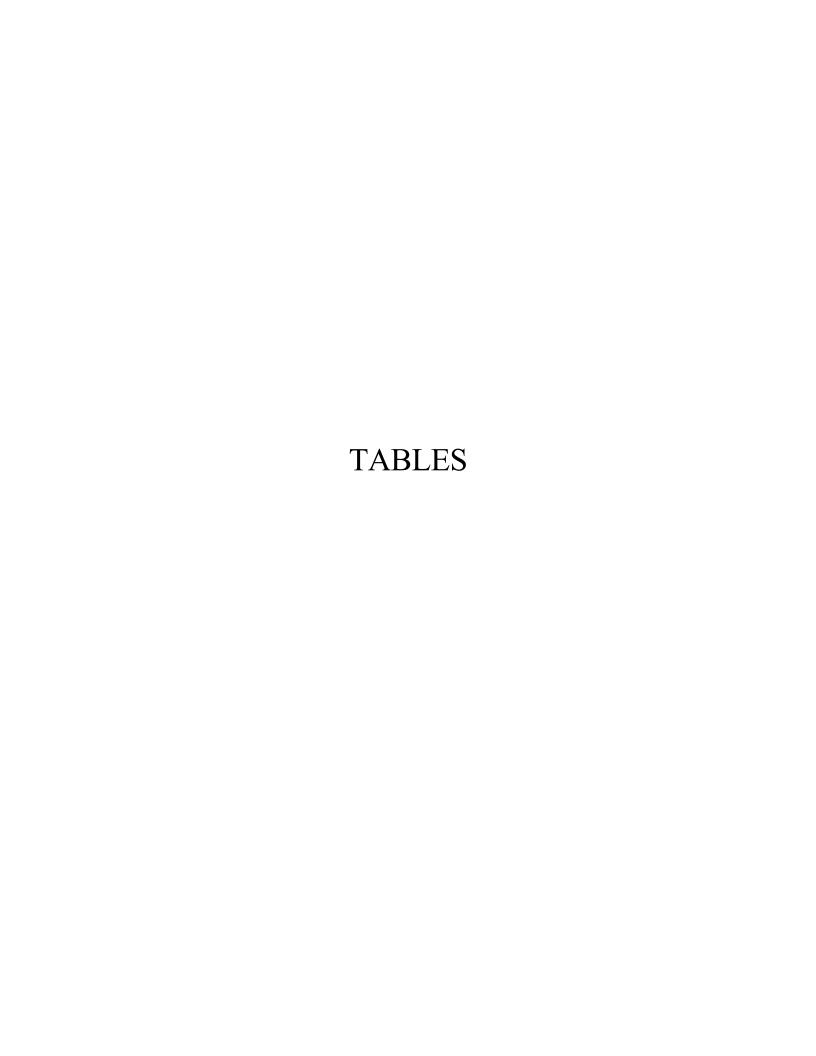


Table 1
Monitoring Well Construction Summary
Hopyard Cleaners
Pleasanton, California

Well I.D.	Date of Completion	Northing	Easting	TOC Elevation (MSL)	Total 1	-	Screen Inte	erval Depth ogs)	Well Casing Material	Well Diameter (inches)
					Borehole	Well	Top	Bottom		
A Zone Moi	nitoring Wells									
MW-1	9/29/2006	2071427.29	6157712.24	325.77	30	30	20.00	30.00	SCH 40 PVC	2
MW-2	9/26/2006	2071357.03	6157791.18	325.69	30	30	20.00	30.00	SCH 40 PVC	2
MW-3	9/27/2006	2071461.21	6157787.94	326.27	30	30	20.00 30.00		SCH 40 PVC	2
MW-4	7/20/2007	2071382.30	6157557.57	326.27	36.5	35	25.00 35.00		SCH 40 PVC	2
B Zone Mon	e Monitoring Wells									
MW-5*	7/19/2007	2071292.25	6157654.24	327.19	60	60	50.00	60.00	SCH 40 PVC	2
MW-6	8/19/2008	2071280.12	6157384.43	324.48	59	59	49.00 59.00		SCH 40 PVC	2
MW-7	8/20/2008	2071076.06	6157645.52	324.55	56	55	45.00	55.00	SCH 40 PVC	2

ft bgs = feet below ground surface

MSL = mean sea level

TOC = Top of Casing

Elevations are based on NAVD 88 Datum

^{*} Conductor casing was installed from 0 to 40 ft bgs.

Table 2
Groundwater Elevations
Hopyard Cleaners
Pleasanton, California

Well LD.	TOC		Depth to	Groundwater
(Screen Interval)	Elevation	Sample Date	Groundwater Below	Elevation
`	(ft MSL)		TOC (ft)	(ft MSL)
A Zone Monitoring		T		
MW-1	325.77	1/26/2009	16.71	309.06
(20-30 ft bgs)		12/10/2008	16.78	308.99
		7/14/2008	13.79	311.98
		5/16/2008	11.70	314.07
		2/15/2008	11.38	314.39
		1/3/2008	13.63	312.14
		8/3/2007	14.40	311.37
		5/11/2007	12.27	313.50
		2/9/2007	13.98	311.79
		11/20/2006	14.88	310.89
MW-2	325.69	1/26/2009	16.17	309.52
(20-30 ft bgs)		12/10/2008	16.24	309.45
		7/14/2008	13.23	312.46
		5/16/2008	11.30	314.39
		2/15/2008	10.87	314.82
		1/3/2008	13.21	312.48
		8/3/2007	13.72	311.97
		5/11/2007	11.87	313.82
		2/9/2007	13.55	312.14
		11/20/2006	14.36	311.33
MW-3	326.27	1/26/2009	17.10	309.17
		12/10/2008	17.17	309.10
(20-30 ft bgs)		7/14/2008	14.21	312.06
		5/16/2008	12.18	314.09
		2/15/2008	11.68	314.59
		1/3/2008	14.02	312.25
		8/3/2007	14.68	311.59
		5/11/2007	12.72	313.55
		2/9/2007	14.41	311.86
		11/20/2006	15.28	310.99
MW-4	326.27	1/26/2009	17.86	308.41
(25-35 ft bgs)		12/10/2008	18.41	307.86
		7/14/2008	13.81	312.46
		5/16/2008	12.12	314.15
		2/15/2008	12.05	314.22
		1/3/2008	14.73	311.54
		8/3/2007	15.85	310.42

Table 2
Groundwater Elevations
Hopyard Cleaners
Pleasanton, California

Well I.D. (Screen Interval)	TOC Elevation (ft MSL)	Sample Date	Depth to Groundwater Below TOC (ft)	Groundwater Elevation (ft MSL)
B Zone Monitoring				
MW-5	327.19	1/26/2009	30.61	296.58
(50-60 ft bgs)		12/10/2008	33.67	293.52
		7/14/2008	32.16	295.03
		5/16/2008	23.06	304.13
		2/15/2008	19.74	307.45
		1/3/2008	22.65	304.54
		8/3/2007	30.51	296.68
MW-6	324.48	1/26/2009	28.10	296.38
(49-59 ft bgs)		12/10/2009	31.14	293.34
MW-7	324.55	1/26/2009	28.19	296.36
(45-55 ft bgs)		12/10/2008	31.21	293.34

ft MSL = feet above mean sea level

TOC = Top of Casing

ft bgs = feet below ground surface

Elevations are based on NAVD 88 Datum

Table 3 Groundwater Gradient Summary Hopyard Cleaners Pleasanton, California

Data	Grad	lient	General
Date	ft/ft	ft/mi	Flow Direction
A Zone			
1/26/2009	0.0045	23.8	West-Northwest
12/10/2008	0.0068	36.1	West-Northwest
7/14/2008	0.0048	25.5	North
5/16/2008	0.0031	16.5	North-Northwest
2/15/2008	0.0038	20.5	Northwest
1/3/2008	0.0025	13.2	Northwest
8/3/2007	0.0070	37.0	West-Northwest
5/11/2007	0.0030	15.8	North-Northwest
2/9/2007	0.0010	5.3	North-Northwest
11/20/2006	0.0040	22.0	Northwest
B Zone			
1/26/2009	0.0012	6.4	Southwest
12/10/2008	0.0012	6.1	Southwest

ft/ft = feet per feet

ft/mi = feet per mile

Table 4
Groundwater Analytical Summary
Hopyard Cleaners
Pleasanton, California

Well I.D.	Sample Date	Sampling Method		le Organic Compo A Method 8260B (u					
(Screen Interval)			cis-1,2-DCE	PCE	TCE				
A Zone Monitoring Wel	lls								
MW-1	1/26/2009	PDB Sampler	240	1,700	320				
(20-30 ft bgs)	12/10/2008	PDB Sampler	250	1,900	350				
	7/14/2008	PDB Sampler	230	1,700	250				
	5/16/2008	Purge and Sample	250	1,600	280				
	5/16/2008	PDB Sampler*	260	1,900	310				
	2/29/2008	PDB Sampler*	330	2,000	330				
	2/15/2008	Purge and Sample	230	1,400	250				
	1/2/2008	Purge and Sample	230	1,600	270				
	8/3/2007	Purge and Sample	260	1,600	270				
	5/11/2007	Purge and Sample	310	2,500	310				
	2/9/2007	Purge and Sample	270 / 270	2,400 / 2,300	290 / 290				
	11/20/2006	Purge and Sample	370	3,100	370				
MW-2	1/26/2009	PDB Sampler	760 / 770	12,000 / 12,000	720 / 730				
(20-30 ft bgs)	12/10/2008	PDB Sampler	840 / 770	15,000/15,000	790 / 740				
	7/14/2008	PDB Sampler	820 / 830	9,500 / 8,100	8,100 530 / 500				
	5/16/2008	Purge and Sample	900 / 930	5,800 / 5,900	460 / 450				
	5/16/2008	PDB Sampler*	940	6,700	480				
	2/29/2008	PDB Sampler*	780	5,300	360				
	2/15/2008	Purge and Sample	690 / 690	4,100 / 4,000	320 / 300				
	1/2/2008	Purge and Sample	940 / 890	8,200 / 8,200	560 / 580				
	8/3/2007	Purge and Sample	1,200 / 1,100	8,000 / 8,100	590 / 570				
	5/11/2007	Purge and Sample	1,000 / 980	7,200 / 7,300	490 / 450				
	2/9/2007	Purge and Sample	760	4,700	350				
	11/20/2006	Purge and Sample	800 / 800	5,700 / 5,800	370 / 360				
MW-3	1/26/2009	PDB Sampler	4.6	42	4.7				
(20-30 ft bgs)	12/10/2008	PDB Sampler	5.6	60	5.5				
	7/14/2008	PDB Sampler	4.3	43	4.0				
	5/16/2008	Purge and Sample	5.0	39	4.3				
	5/16/2008	PDB Sampler*	5.4	46	4.4				
	2/29/2008	PDB Sampler*	6.9	58	5.9				
	2/15/2008	Purge and Sample	6.2	44	5.1				
	1/2/2008	Purge and Sample	5.2	46	4.6				
	8/3/2007	Purge and Sample	4.7	37	4.2				
	5/11/2007	Purge and Sample	5.5	43	4.4				
	2/9/2007	Purge and Sample	5.3	42	4.2				
	11/20/2006	Purge and Sample	9.5	93	7.2				

Table 4 Groundwater Analytical Summary Hopyard Cleaners Pleasanton, California

Well I.D. (Screen Interval)	Sample Date	Sampling Method		e Organic Compo Method 8260B (
(Sercen Interval)			cis-1,2-DCE	PCE	TCE
MW-4	1/26/2009	PDB Sampler	4.3	< 0.50	4.9
(25-35 ft bgs)	12/10/2008	PDB Sampler	4.0	< 0.50	3.7
	7/14/2008	PDB Sampler	4.7	< 0.50	4.0
	5/16/2008	Purge and Sample	3.7	< 0.50	2.6
	5/16/2008	PDB Sampler*	3.6	< 0.50	2.7
	2/29/2008	PDB Sampler*	3.4	< 0.50	3.0
	2/15/2008	Purge and Sample	4.2	< 0.50	4.0
	1/3/2008	Purge and Sample	4.2	< 0.50	3.5
	8/3/2007	Purge and Sample	4.6	< 0.50	3.5
B Zone Monitoring Wel	lls				
MW-5	1/26/2009	PDB Sampler	< 0.50	37	< 0.50
	12/10/2008	PDB Sampler	< 0.50	49	< 0.50
(50-60 ft bgs)	7/14/2008	PDB Sampler	< 0.50	31	< 0.50
	5/16/2008	Purge and Sample	< 0.50	24	< 0.50
	5/16/2008	PDB Sampler*	< 0.50	34	< 0.50
	2/29/2008	PDB Sampler (52.5 ft bgs)*	< 0.50	41	< 0.50
	2/29/2008	PDB Sampler (57.5 ft bgs)*	< 0.50	33	< 0.50
	2/15/2008	Purge and Sample	< 0.50	26	< 0.50
	1/3/2008	Purge and Sample	< 0.50	38	< 0.50
	8/3/2007	Purge and Sample	< 0.50	37	< 0.50
MW-6	1/26/2009	PDB Sampler	< 0.50	< 0.50	< 0.50
(49-59 ft bgs)	12/10/2008	PDB Sampler (51.5 ft bgs)*	< 0.50	< 0.50	< 0.50
	12/10/2008	PDB Sampler (56.5 ft bgs)*	< 0.50	< 0.50	< 0.50
MW-7	1/26/2009	PDB Sampler	< 0.50	9.9	< 0.50
(45-55 ft bgs)	12/10/2008	PDB Sampler (47.5 ft bgs)*	< 0.50	9.8	< 0.50
	12/10/2008	PDB Sampler (52.5 ft bgs)*	< 0.50	10	< 0.50

Notes:

Table shows only compounds detected above the laboratory reporting limit.

cis-1,2-DCE = cis-1,2-dichloroethene

PCE = tetrachloroethene

TCE = trichloroethene

"-- / --" = result on right represents duplicate sample

ft bgs = feet below ground surface

PDB = Passive Diffusion Bag Sampler

* Samples collected as part of the PDB comparison study. PDBs were deployed at two depths in the following wells to evaluate stratification: at 52.5 and 57.5 ft bgs in MW-5 for the 1st Quarter 2008 event, at 51.5 and 56.5 ft bgs in MW-6 for the 4th Quarter 2008 event, and at 47.5 and 52.5 ft bgs in MW-7 for the 4th Quarter 2008 event.

Table 5
SVE System Performance Monitoring Results
Hopyard Cleaners
2771 Hopyard Road, Pleasanton, California

				Q 7 1 Q PP					<u> </u>			35100 77		~ · · · ~ · · ·				
-		1	1	SYST	EM MEAS	SUREMEN					ı		MASS RE	MOVAL (CALCULA	ATIONS		
		Operation	Influent	System	Influent	Influent	Mid-	Effluent	Temp. Before				Total	PCE	Mass	Mass	Mass Removed	Cumulative
	Time	Time	Flow	Temp.	Vacuum	Conc.	Point	Conc.	GAC Vessels	¥7	El4-	El4-	Operation	Conc.	Removal Rate	Removal	Since Last	Mass
Sample		(Hour)	Rate	(°F)	(in Hg)	(ug/m^3)	Conc.	(ug/m^3)	(°F)*	Vacuum (in water)	Flowrate (cfm)	Flowrate (scfm)	Time (hr)	. 3	(lbs/hr)	Rate (lbs/day)	Sampling Event (lbs)	Removal (lbs)
Date			(ft/min)	` '			(ug/m ³)		` ′	(in water)	(CIIII)	(SCIII)	(111)	(mg/m³)	(IDS/III)	(IDS/day)	. ,	
21-Aug-08	9:15	7,569.2			10.0	89,700	1,380	690		136				89.7			0.00	0.00
22-Aug-08	9:25	7,593.3	4,590	83.5	10.0	37,950	2,070	0.0		136	210.54	136.22	24.17	38.0	0.0326	0.7817	0.79	0.79
23-Aug-08	10:00	7,618.0	4,690	78.3	9.5	4,830	1,380	690		129	215.13	144.06	48.75	4.8	0.0115	0.2770	0.28	1.07
24-Aug-08	14:02	7,646.0	4,550	79.5	10.0	6,210	2,070	0.0		136	208.71	136.04	76.78	6.2	0.0028	0.0675	0.08	1.15
25-Aug-08	16:22	7,672.4	4,450	87.2	10.0	7,590	2,070	690		136	204.12	131.17	103.12	7.6	0.0034	0.0814	0.09	1.24
27-Aug-08	8:14	7,712.1	4,520	74.0	10.0	45,540	690	0.0		136	207.33	136.53	142.98	45.5	0.0136	0.3261	0.54	1.78
29-Aug-08	8:02	7,757.7	4,380	77.9	9.5	12,420				129	200.91	134.64	190.78	12.4	0.0146	0.3508	0.70	2.48
2-Sep-08	9:14	7,853.3	4,250	77.5	10.0	12,420	690	0.0		136	194.95	127.54	287.98	12.4	0.0059	0.1424	0.58	3.06
8-Sep-08	8:40	7,996.2	4,290	76.8	8.5	14,490	690	0.0		116	196.78	138.60	379.14	14.5	0.0070	0.1677	0.64	3.69
18-Sep-08	10:40	8,238.2	4,300	79.0	8.0	4,830	0.0	0.0		109	197.24	141.59	520.31	4.8	0.0051	0.1230	0.72	4.42
8-Oct-08	10:00	8,715.1	4,300	83.8	8.0	5,520	0.0	0.0		109	197.24	140.34	799.92	5.5	0.0027	0.0653	0.76	5.18
17-Nov-08	9:30	9,675.1	4,300	66	8.0	6,210	0.0	0.0		109	197.24	145.09	1359.63	6.2	0.0032	0.0765	1.78	6.96
5-Dec-08	9:26	10,107.1	4,775	49.8	8.0	4,830	1,380	0.0		109	219.03	166.23	1611.59	4.8	0.0034	0.0825	0.87	7.83
6-Jan-09	9:10	10,847.7	4,610	53.5	7.5	1,380	0.0	0.0		102	211.46	162.96	2059.43	1.4	0.0019	0.0455	0.85	8.68
21-Jan-09	8:25	11,233.5	4,490	51.8	9.0	4,830	3,450	690		122	205.95	148.60	2268.99	4.8	0.0017	0.0415	0.36	9.04
21-Jan-09	15:30	11,240.5	3,445	67.8	10.5	3,450	2,070	2,070		143	158.02	102.64	2273.13	3.5	0.0016	0.0382	0.01	9.04
5-Feb-09	9:05	11,562.4	4,130	56.6	10.0	6,900**	5,520**	690**	-	136	189.44	128.95	2479.38	6.9**	0.0008	0.0200	0.17	9.22
5-Feb-09	10:30	11,563.8	4,470	59.1	10.0	154,600**	93,840**	104,880**	-	136	205.04	138.90	2480.21	154.56**	0.0009	0.0215	0.00	9.22
19-Feb-09	8:42	11,898.0	4,440	55.1	9.0	0.0	0.0	0.0	-	122	203.66	146.01	2675.16	0.0	0.0009	0.0226	0.18	9.40
19-Feb-09	12:00	11,899.7	3,110	63.8	10.0	0.0	0.0	0.0	102.3	136	142.65	95.77	2675.20	0.0	0.0000	0.0000	0.00	9.40
26-Feb-09	9:15	12,064.9	3,150	60.3	9.0	0.0	0.0	0.0	97.4	122	144.49	102.55	2771.60	0.0	0.0000	0.0000	0.00	9.40
26-Feb-09	10:07	12,068.8	3,500	60.9	8.0	0.0	0.0	0.0	94.8	109	160.54	119.25	2772.10	0.0	0.0000	0.0000	0.00	9.40
12-Mar-09	9:40	12,400.3	3,650	56.1	7.0	1,097	0.0	0.0	77.4	95	167.42	131.24	2800.06	1.1	0.0003	0.0065	0.01	9.41

Notes/Assumptions:

- 1. Inlet pipe diameter is 3".
- 2. SVE operations were reduced from 24 hours per day to 14 hours (8 am to 10 pm) per day on 3 September 2008. SVE operations were reduced to 2 hours (8 am to 10 am) per day on 26 February 2009.
- 3. Vapor density of PCE is estimated to be 6,900 g/m3 at 20C.
- 4. SCFM(at 14.7psia and 68° F) = CFM x([(Pg + Patm)/(Patm)] x [(68 + 460)/(Tact +460)])
- 5. Concentrations and mass removal calculated as mass of PCE.
- * On 19 February 2009, the blower was moved in front of the carbon vessels in the treatment process. Temperature measurement were collected before carbon vessels to confirm that vapor temperatures are below 12B prior to entering the carbon vessels.
- ** PID readings from 5 February 2009 were anomolously high, indicating possible instrumentation error. To be conservative, these influent concentrations were not included in mass removal calculations.

ft/min = feet per minute

ug/m³ = micrograms per cubic meter

°F = degrees fahrenheit

in Hg = inches mercury

in water = inches water

cfm = cubic feet per minute

scfm = standard cubic feet per minute

hr = hour

 $mg/m^3 = milligrams per cubic meter$

yr = year

lbs = pounds

Table 6 SVE Influent Sample Analytical Summary Hopyard Cleaners Pleasanton, California

				Sampl	le Date			
VOC	21-A	ug-08	2-Se	ep-08	5-De	ec-08	12-M	ar-09
units	ppmv	ug/m ³	ppmv	ppmv ug/m ³		ug/m ³	ppmv	ug/m ³
PCE	3.600	24,000	1.200	8,500	0.340	2,300	0.250	1,700
TCE	0.051	280	0.029	160	0.012	64	0.0068	36
Other ¹	0.022	66	0.0075	22	0.043	112.6	0.0134	35.1
Total VOCs	3.651	24,346	1.237	8,682	0.395	2,476.6	0.270	1,771.1
Influent PID Reading ²	13.8	95,220	1.8	12,420	0.7	4,830	0.159	1,097

Notes:

Table shows only compounds detected above the laboratory reporting limit

VOC - Volatile Organic Compound; analyzed by TO-15

ppmv - parts per million by volume

ug/m³ - micrograms per cubic meter

cis-1,2-DCE - cis-1,2-dichloroethene

PCE - tetrachloroethene

TCE - trichloroethene

PID - Photoionization Detector

- (1) Tetrahydrofuran was detected at a concentration of 0.022 ppmv on 21 August 2008; 2-butanone was detected at a concentration of 0.0075 ppmv on 2 September 2008; freon 12 was detected at a concentration of 0.0014 ppmv, ethanol was detected at 0.0082 ppmv, acetone was detected at 0.0099 ppmv, carbon disulfide was detected at 0.0025 ppmv, methylene chloride was detected at 0.0014 ppmv, 2-butanone was detected at 0.0025 ppmv, tetrahydrofuran was detected at 0.0014 ppmv, benzene was detected at 0.0045 ppmv, and toluene was detected at 0.0076 ppmv on 5 December 2008; and acetone was detected at 0.0079 ppmv, 2-butanono was detected at 0.0026 ppmv, and tetrahydrofuran was detected at 0.0029 ppmv on 12 March 2009.
- (2) PID screening results from the date sampling was conducted, as presented on Table 5. PID results are calculated from part per million by volume to ug/m³ as PCE.

Table 7 SVE Well Monitoring Results Hopyard Cleaners 2771 Hopyard Road, Pleasanton, California

			N	IANIFOL	LD		SV	E-1			SV	E-2			SV	E-3			SV	E-4			SV	/E-5	
Monitoring	Manitania a Evant	Extraction		Flow				PI	D ²			PII) ²			PI	D ²			PI	D ²			PII	D 2
Date	Monitoring Event	Duration 1 (hr/day)	Time	Rate (scfm)	Vacuum (in Hg)	Time	Vacuum (in Hg)	ug/m³ as PCE	ppmv	Time	Vacuum (in Hg)	ug/m³ as PCE	ppmv	Time	Vacuum (in Hg)	ug/m³ as PCE	ppmv	Time	Vacuum (in Hg)	ug/m³ as PCE	ppmv	Time	Vacuum (in Hg)	ug/m³ as PCE	ppmv
21-Aug-08	Start up Day 1	24	9:22	240		9:24	0	322,920	46.8	9:22	1.5	164,220	23.8	9:21	2.0	34,500	5.0	9:25	1.5	167,670	24.3	9:20	2.0	60,720	8.8
22-Aug-08	Start-up Day 2	24	9:41	240		9:42	0	141,450	20.5	9:40	1.75	82,800	12.0	9:38	2.0	14,490	2.1	9:44	1.5	57,960	8.4	9:37	2.0	28,980	4.2
23-Aug-08	Start-up Day 3	24	10:35	240		10:38	0	86,250	12.5	10:34	1.5	53,820	7.8	10:28	0	15,870	2.3	10:26	1.0	44,160	6.4	10:31	2.0	24,840	3.6
25-Aug-08	Start-up Day 5	24	16:52	235		16:50	0	64,170	9.3	16:58	0	33,810	4.9	16:55	1.0	11,040	1.6	4:46	1.0	33,120	4.8	16:53	2.0	17,940	2.6
27-Aug-08	Start-up Day 7	24	8:36	240		8:38	0	49,680	7.2	8:36	1.5	24,840	3.6	8:35	2.0	4,140	0.6	8:39	1.5	61,410	8.9	8:34	2.0	10,350	1.5
2-Sep-08	Start-up Day 13/Week 2	24	9:43	230	3.5	9:44	0	24,150	3.5	9:42	1.5	15,180	2.2	9:40	1.75	4,830	0.7	9:45	1.5	13,110	1.9	9:36	1.5	8,280	1.2
8-Sep-08	Start-up Week 3	14	8:58	230	3.75	9:01	0	17,940	2.6	8:59	1.25	19,320	2.8	8:58	1.5	16,560	2.4	9:02	1.25	8,280	1.2	8:57	1.5	14,490	2.1
18-Sep-08	1st Month	14	11:14	235	4	11:16	1.2	12,420	1.8	11:14	1.5	5,520	0.8	11:12	1.5	0	0.0	11:17	1.3	5,520	0.8	11:10	1.5	3,450	0.5
8-Oct-08	2nd Month	14	10:40	235	3.75	11:04	1.2	8,970	1.3	11:00	1.5	7,590	1.1	10:57	1.4	3,450	0.5	11:07	1.3	6,900	1.0	10:51	1.5	5,520	0.8
17-Nov-08	3rd Month	14	9:45	235	3.5	9:48	1.1	6,900	1.0	9:46	1.4	4,830	0.7	9:44	1.3	3,450	0.5	9:50	1.2	4,830	0.7	9:42	1.4	5,520	0.8
5-Dec-08	4th Month	14	11:20	240	3.5	11:21	1.1	4,830	0.7	11:19	1.3	3,450	0.5	11:18	1.3	2,070	0.3	11:22	1.1	3,450	0.5	11:17	1.4	3,450	0.5
6-Jan-09	5th Month	14	9:44	240	3.5	9:45	1.0	690	0.1	9:43	1.3	0	0	9:42	1.2	0	0.0	9:46	1.1	690	0.1	9:40	1.3	0	0.0
21-Jan-09	Cycle Wells ³	14	9:02	235	4.5	9:03	1.5	10,350	1.5	9:00	2.4	11,730	1.7	10:06	OFF	115,230	16.7	9:06	1.7	44,850	6.5	10:05	OFF	124,890	18.1
21-Jan-09	Cycle Wells ³	14	15:47	220	5.5	15:49	OFF	4,140	0.6	15:46	OFF	2,760	0.4	15:45	3.1	2,760	0.4	15:50	OFF	6,210	0.9	15:43	3.2	690	0.1
5-Feb-09	6th Month/Cycle Wells ³	14	9:27	230	5.0	9:28	OFF	84,180*	12.2*	9:26	OFF	73,830*	10.7*	9:24	2.9	74,520*	10.8*	9:29	OFF	178,710*	25.9*	9:40	2.9	252,540*	36.6*
5-Feb-09	Cycle Wells ³	14	10:41	230	4.5	10:43	1.5	189,750*	27.5*	10:42	1.2	158,700*	23.0*	10:40	OFF	107,640*	15.6*	10:45	1.5	230,460*	33.4*	10:39	OFF	142,830*	20.7*
19-Feb-09	Cycle Wells ³	14	9:02	235	4.5	9:03	1.5	0	0.0	9:02	2.3	0	0.0	9:00	OFF	40,710	5.9	9:04	1.5	0	0.0	8:59	OFF	15,180	2.2
19-Feb-09	Cycle Wells ³	14	12:10	165	3.0	12:10	OFF	0	0.0	12:09	OFF	0	0.0	12:07	2.0	0	0.0	12:12	OFF	0	0.0	12:06	2.1	0	0.0
26-Feb-09	Cycle Wells ³	2	9:29	165	3.0	9:31	OFF	21,390	3.1	9:28	OFF	17,940	2.6	9:27	2.0	0	0.0	9:32	OFF	65,550	9.5	9:26	2.1	0	0.0
26-Feb-09	Cycle Wells ³	2	10:19	230	2.0	10:19	0.7	690	0.1	10:18	0.7	0	0.0	10:17	0.7	0	0.0	10:20	0.8	690	0.1	10:16	0.9	0	0.0
12-Mar-09	7th Month	2	9:21	180	2.0	9:23	0.7	497	0.072	9:22	0.8	1,780	0.258	9:20	0.8	276	0.040	9:24	0.8	373	0.054	9:19	0.8	573	0.083

Notes:

- 1) A timer was installed on the system and was set to run from 8 am to 10 pm (14 hrs/day) on 3 September 2008. Operation was then reduced to 8 am to 10 am (2 hrs/day) on 26 February 2009 to optimize the system
- 2) PID screening was conducted using a MiniRae 2000 capable of detecting VOCs in the ppmv range. Beginning on 12 March 2009, a ppbRae was used to detect concentrations at lower levels, in the parts per billion
- 3) On 21 January, 5 February, 19 February, and 26 February 2009, monitoring was conducted twice: before cycling the SVE wells and approximately 30 minutes after cycling the SVE wells.
- *PID readings from 5 February 2009 were anomolously high, indicating possible instrumentation error.

ft = feet

min = minute

in Hg = inches of mercury

ug/m³ as PCE= micrograms per cubic meter as equivalent tetrachloroethene

ppmv = parts per million volume

scfm = standard cubic feet per minute

OFF = well turned off during well cycling

Table 8
Soil Oxidant Demand Laboratory Results Summary
Hopyard Cleaners
Pleasanton, California

Annaloute		TT 24		SOD-1			SOD-2		SOD-3				
Analys	S1S	Unit	21-22.5 ft bgs	27.5-29 ft bgs	30.5-32 ft bgs	21-22 ft bgs	25-26 ft bgs	29-30 ft bgs	21-22 ft bgs	25-26 ft bgs	29-30 ft bgs		
	Soil Type		Dark Grayish Brown Clay	Olive Gray Clay	Dark Gray Silt								
Cail Duam aution	Specific Gravity		2.77	2.78	2.79	-							
Soil Properties	Dry Bulk Density	lbs/ft ³	94.8	98.8	96.5								
	Porosity	%	45.2	43.1	44.6								
	Foc	%	0.78	1.32	1.12	1.23	1.02	0.73	0.64	0.71	0.64		
	Antimony	mg/kg	<2.0	<2.1	<2.0								
	Arsenic	mg/kg	5.1	7.7	1.6	-							
	Barium	mg/kg	130	260	170								
	Beryllium	mg/kg	< 0.51	0.60	0.50								
	Cadmium	mg/kg	< 0.51	< 0.52	< 0.50								
	Chromium	mg/kg	52	64	61								
	Cobalt	mg/kg	11	16	11								
Metals	Copper	mg/kg	25	33	37								
Wietais	Lead	mg/kg	5.9	7.3	7.7								
	Molybdenum	mg/kg	<1.0	<1.0	< 0.99								
	Nickel	mg/kg	71	81	79								
	Selenium	mg/kg	<2.0	<2.1	<2.0								
	Silver	mg/kg	<1.0	<1.0	< 0.99								
	Thallium	mg/kg	<1.0	<1.0	< 0.99								
	Vanadium	mg/kg	26	36	30								
	Zinc	mg/kg	41	52	58								
	MnO ₄	g/kg	9.48	22.37	12.09	23.92	5.19	10.31	4.27	5.12	5.59		
Soil Oxidant Demand	KMnO ₄ (1)	g/kg	12.60	29.73	16.07	31.78	6.90	13.70	5.67	6.80	7.43		
	NaMnO ₄ (1)	g/kg	11.32	26.70	14.43	28.54	6.19	12.30	5.10	6.11	6.67		

ft bgs = feet below ground surface

 MnO_4 = permanganate

 $KMnO_4$ = potassium permanganate

NaMnO_{4 =} sodium permanganate

Foc = fraction organic carbon

g/kg = grams per kilogram

mg/kg = milligram per kilogram

lbs/ft³ = pounds per cubic feet

SOD-1 samples were collected on 21 August 2008. SOD-2 and SOD-3 samples were collected on 26 January 2009.

(1) NaMnO4 and KMnO4 values are calculated by the laboratory from the MnO4 SOD and relative molecular weights of the compounds.

Table 9 Permanganate Required for Treatment of Shallow Groundwater Hopyard Cleaners Pleasanton, California

SOD Sampling Location	Near MW-2											Near MW-1						
Amendment	KMnO ₄						NaMnO ₄						KMnO ₄			NaMnO ₄		
Sample Location	SOD-1	SOD-2	SOD-1	SOD-2	SOD-1	SOD-2	SOD-1	SOD-2	SOD-1	SOD-2	SOD-1	SOD-2	SOD-3	SOD-3	SOD-3	SOD-3	SOD-3	SOD-3
Sample Depth (ft bgs)*	21-22	21-22	27.5-28.5	25-26	30.5-31.5	29-30	21-22	21-22	27.5-28.5	25-26	30.5-31.5	29-30	21-22	25-26	29-30	21-22	25-26	29-30
Soil Oxidant Demand (mg/kg)	12.60	31.78	29.73	6.90	16.07	13.70	11.32	28.54	26.70	6.19	14.43	12.30	5.67	6.80	7.43	5.10	6.11	6.67
Estimated Amendment Requirement (lbs) (1)	58,400	147,000	137,500	32,100	74,400	63,500	130,000	327,100	305,900	71,400	165,600	141,300	26,400	31,700	34,500	58,800	70,400	76,900
Injection Volume (gal) (2)	343,200	863,700	807,800	188,500	437,300	373,100	58,200	146,400	136,900	31,900	74,100	63,200	155,400	186,000	203,000	26,300	31,500	34,400
Portion of Pore Volume (%) (3)	36%	93%	90%	20%	47%	40%	6%	16%	15%	3%	8%	7%	17%	20%	22%	3%	3%	4%
Estimated Injection Time (business days) ⁽⁴⁾	143	360	337	79	182	155	24	61	57	13	31	26	65	78	85	11	13	14

Notes:

Bold numbers indicate highest values.

--/-- = SOD-1 Result / SOD-2 Result

ft bgs = feet below ground surface

mg/kg = milligram per kilogram

lbs = pounds

gal = gallons

 $KMnO_4$ = potassium permanganate

 $NaMnO_4 = sodium permanganate$

 $lbs/yd^3 = pounds$ amendment per cubic yard of soil

*Samples Intervals for SOD-1 were 21-22.5, 27.5-29, and 30.5-32 ft bgs due to low sample recovery. The target sample intervals were achieved at SOD-2 and SOD-3.

- (1) Assumes dry crystals for KMnO₄ and 40% liquid solution for NaMnO₄.
- (2) Assumes 2% KMnO₄ solution and 10% NaMnO₄ solution, as recommended by Carus.
- (3) Desired percent of pore space is less than 10%.
- (4) Injection time only, assuming 5 gallons per minute injection rate at 1,500 pounds per square inch. Does not include time for solution preparation and time to move and setup on each injection point.
- (5) Permanganate requirements for SOD-2 and SOD-3 were calculated using the average porosity of the three samples collected at SOD-1.

Table 10 EISB Study - Gene-Trac and Anion Results Summary Hopyard Cleaners Pleasanton, California

		Field Par	Field Parameters Quantitative Gene-Trac Results							Anions Results (EPA Method 300.0)					
Well I.D.	Sample Date	ORP (mV)	DO (mg/L)	<i>Deha</i> % of Microbial Population	lococcoides Enumeration (cells/Liter)	Interpretation	Vinyl Chlorid % of Microbial Population	Enumeration (cells/L)	Chloride (mg/L)	Nitrate as NO3 (mg/L)	Nitrite as NO2 (mg/L)	Sulfate as SO4 (mg/L)			
MW-1	1/26/2009	63.5	0.15	0.008-0.03%	2E+05	Moderate Concentration ¹	NQ ²	Detected ³	83	<0.50	<0.50	130			
MW-2	1/26/2009	118.2	0.12	0.005-0.02% / 0.007-0.02%	2E+05 / 2E+05	Moderate Concentration ¹	NQ ² / 0.0009-0.003%	Inconclusive ⁴ / 3E+04	110 / 110	3.9 / 3.9	<0.50 / <0.50	140 / 140			
MW-3	1/26/2009	150.6	0.24	0.004-0.01%	1E+05	Moderate Concentration ¹	NQ ²	Detected ³	120	13	<0.50	140			

Notes:

- 1. Moderate Concentration indicates the presence of dehalococcoides at concentrations which may, or may not, be associated with observable dechlorination.
- 2. Not Quantifiable (NQ) indicates that vinyl chloride reductase was detected but not able to be quantified.
- 3. Detected but not quantifiable. The sample specific quantitation limit is 2E+04/liter.
- 4. The results indicate extremely low concentrations of vinyl chloride reductase DNA at or below the sample specific quantitation limit (1E+04/liter); however, test results were insufficient to assign a conclusive positive results for this sample.

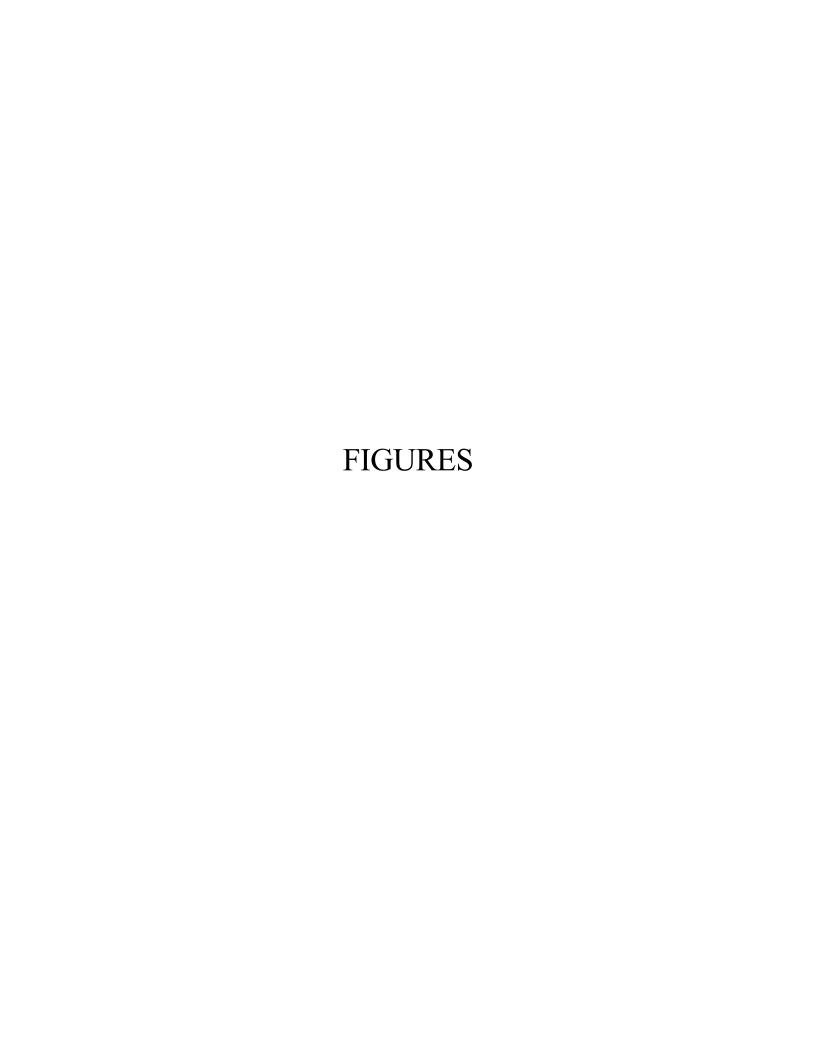
DO = dissolved oxygen

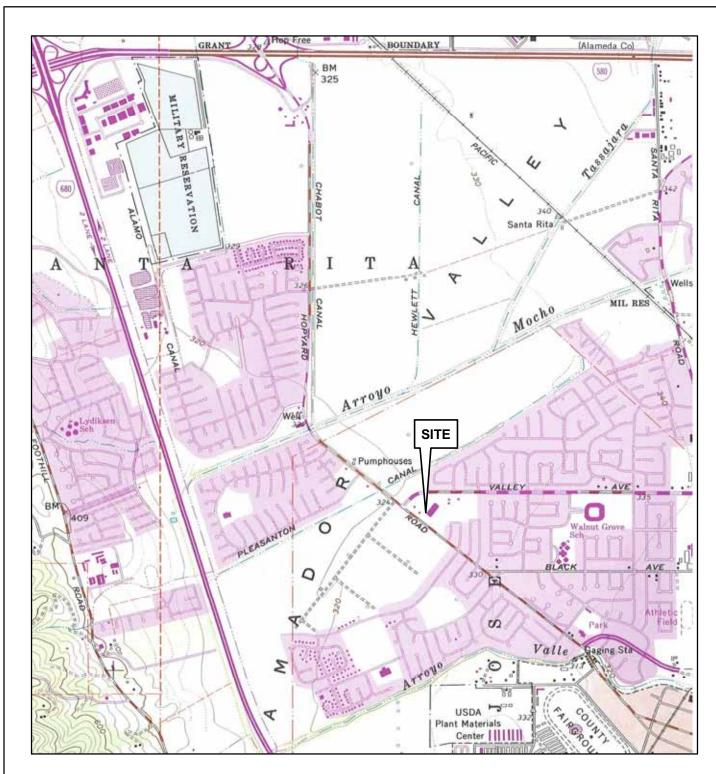
ORP = oxidation reduction potential

mg = milligrams

L = liter

mV = millivolts





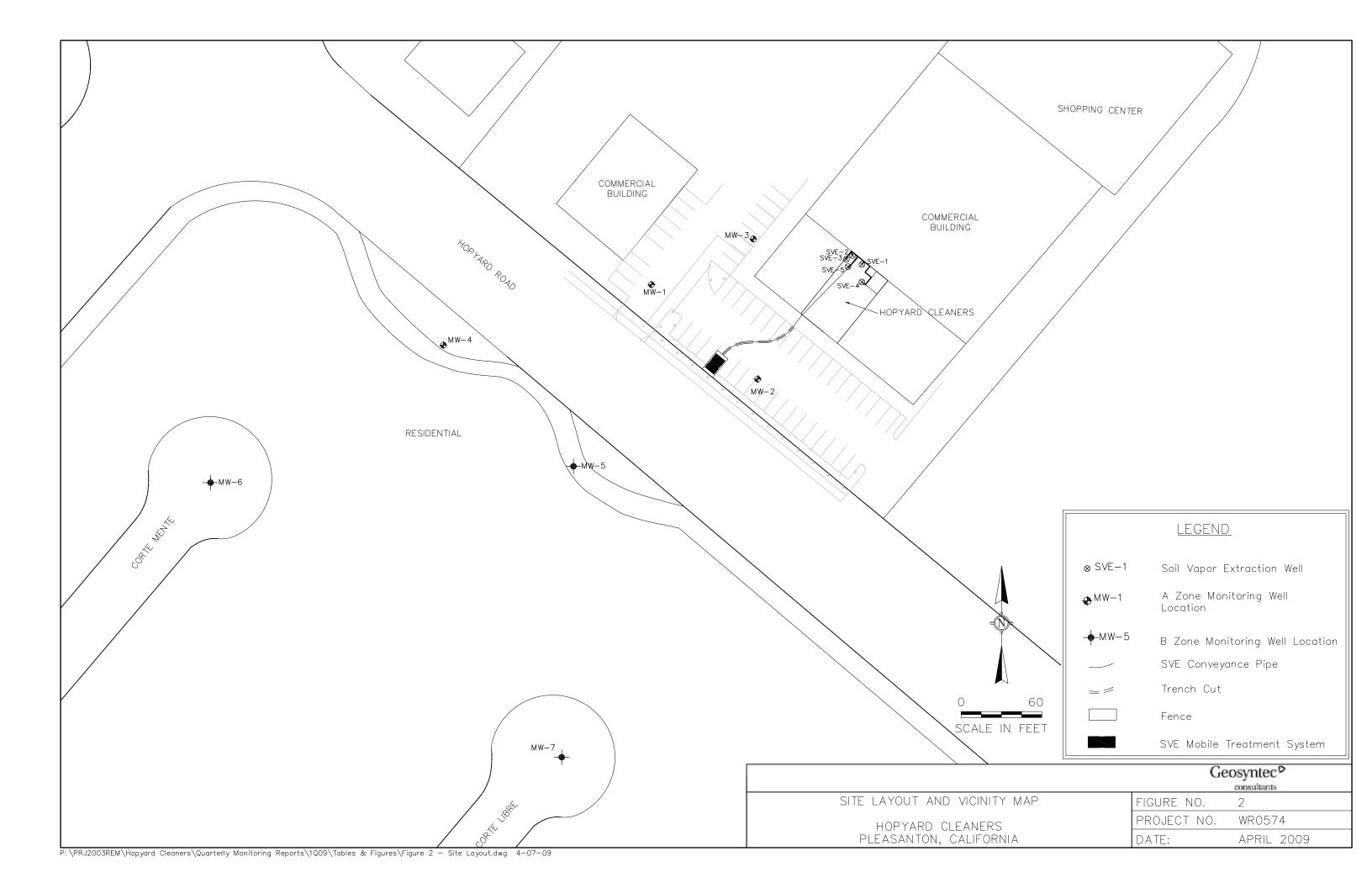
Topo Source: U.S.G.S 7.5 Minute Series, Dublin, CA Quadrangle (1980) Contour Interval = 40 Feet

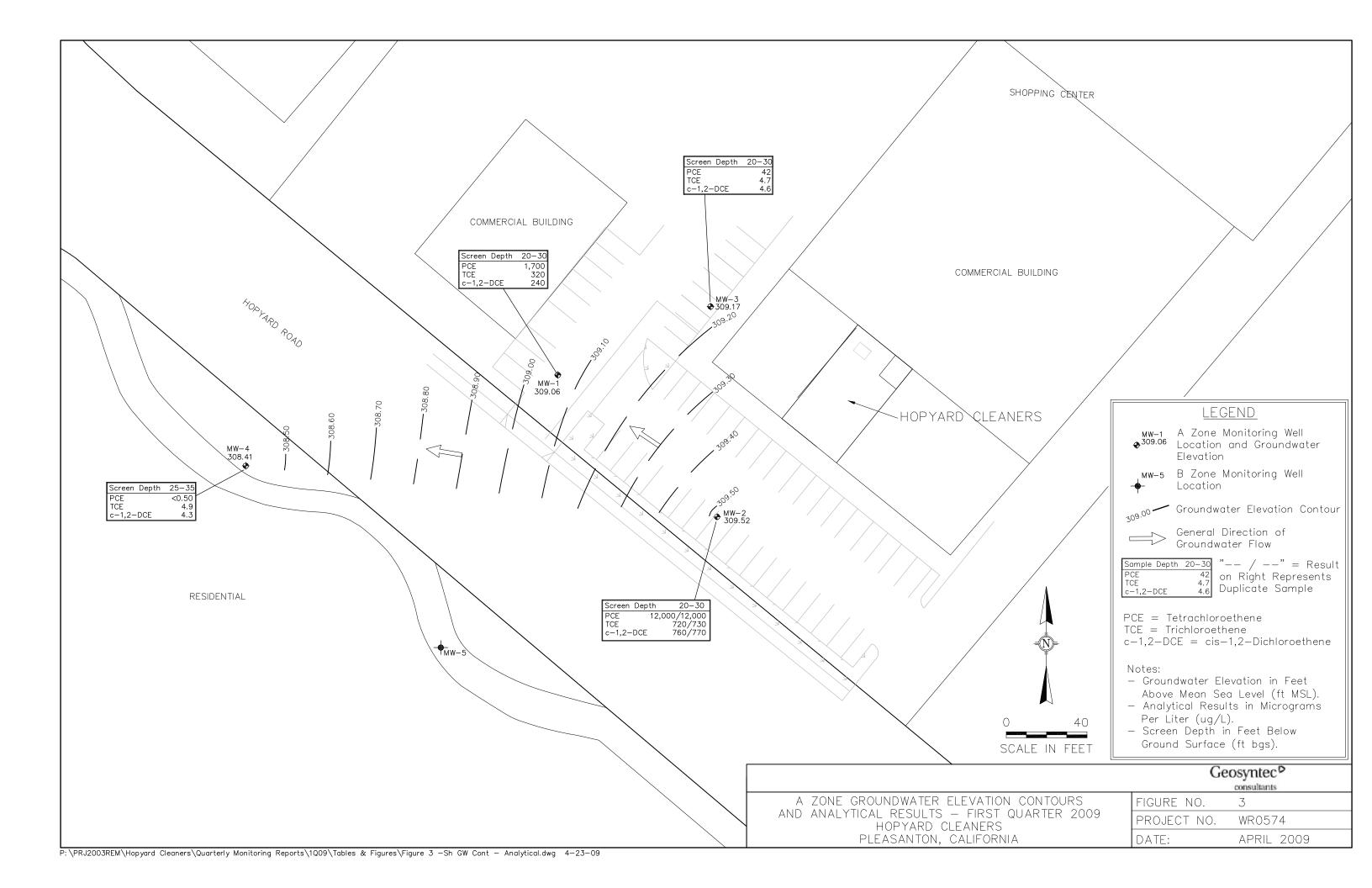
SITE LOCATION MAP HOPYARD CLEANERS 2771 HOPYARD ROAD PLEASANTON, CALIFORNIA

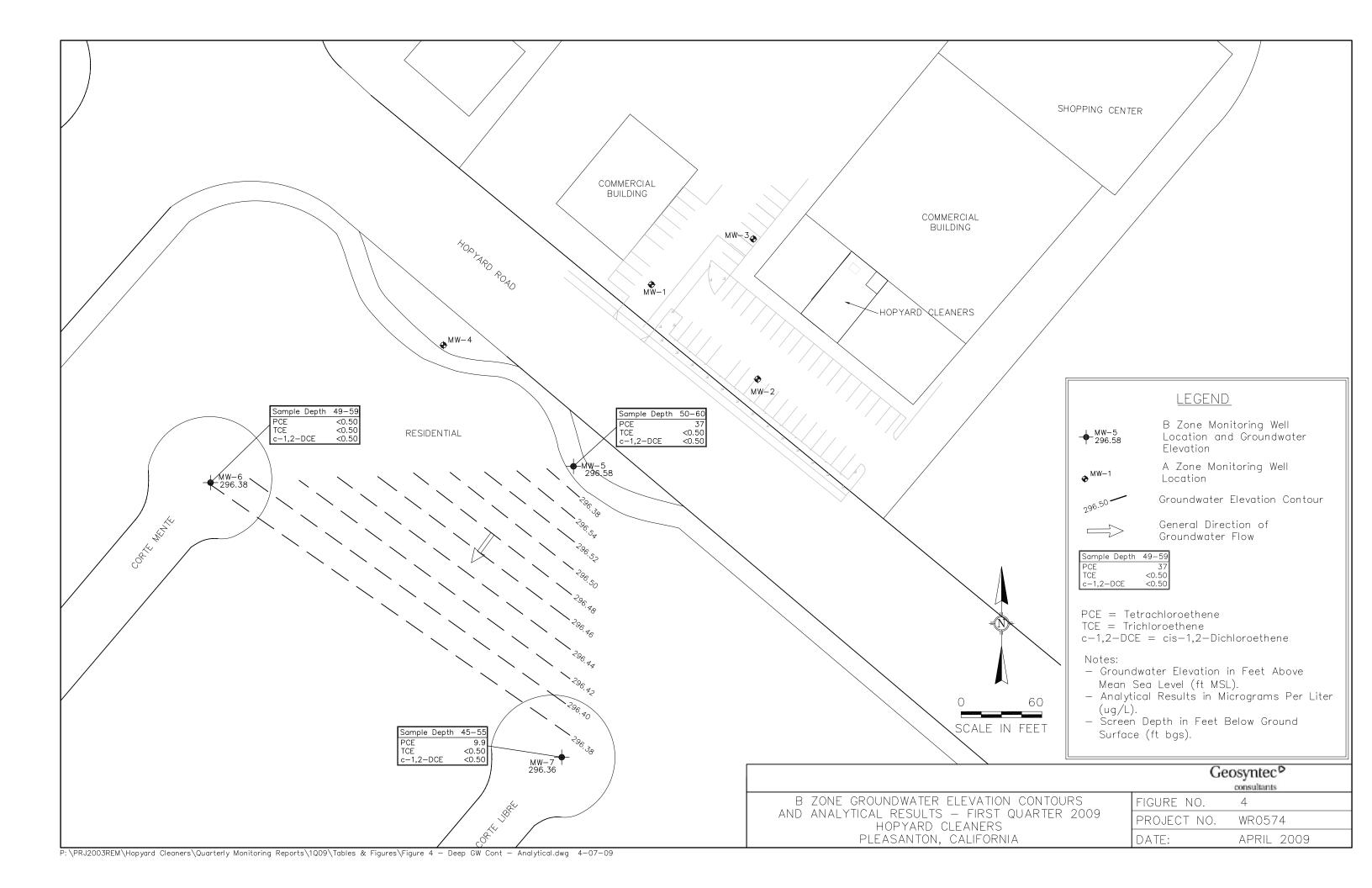


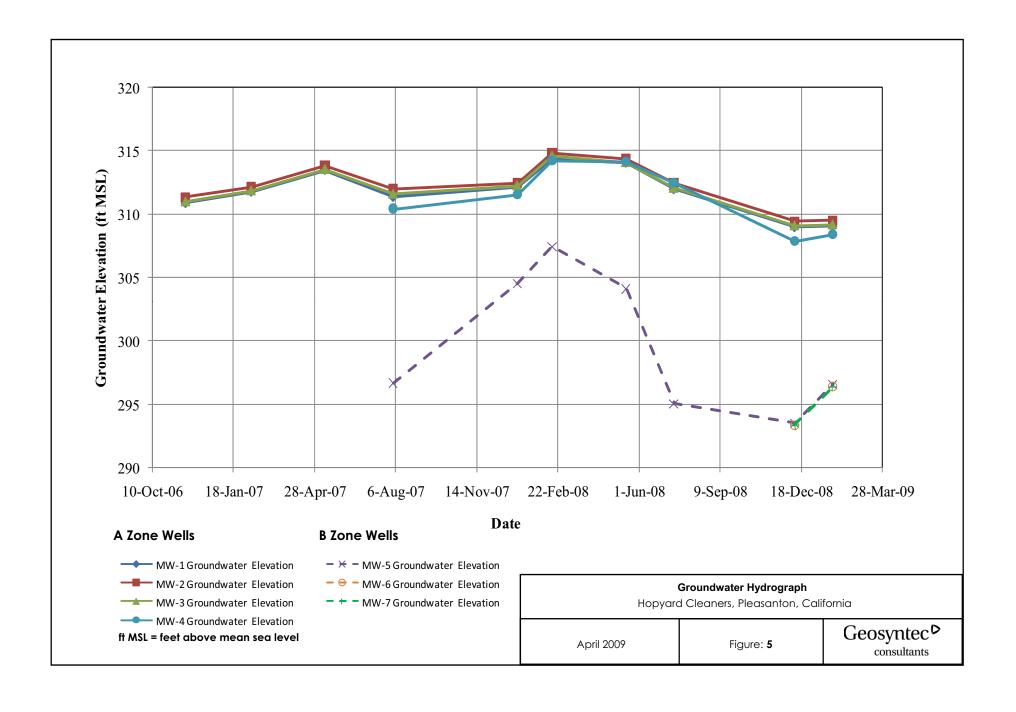


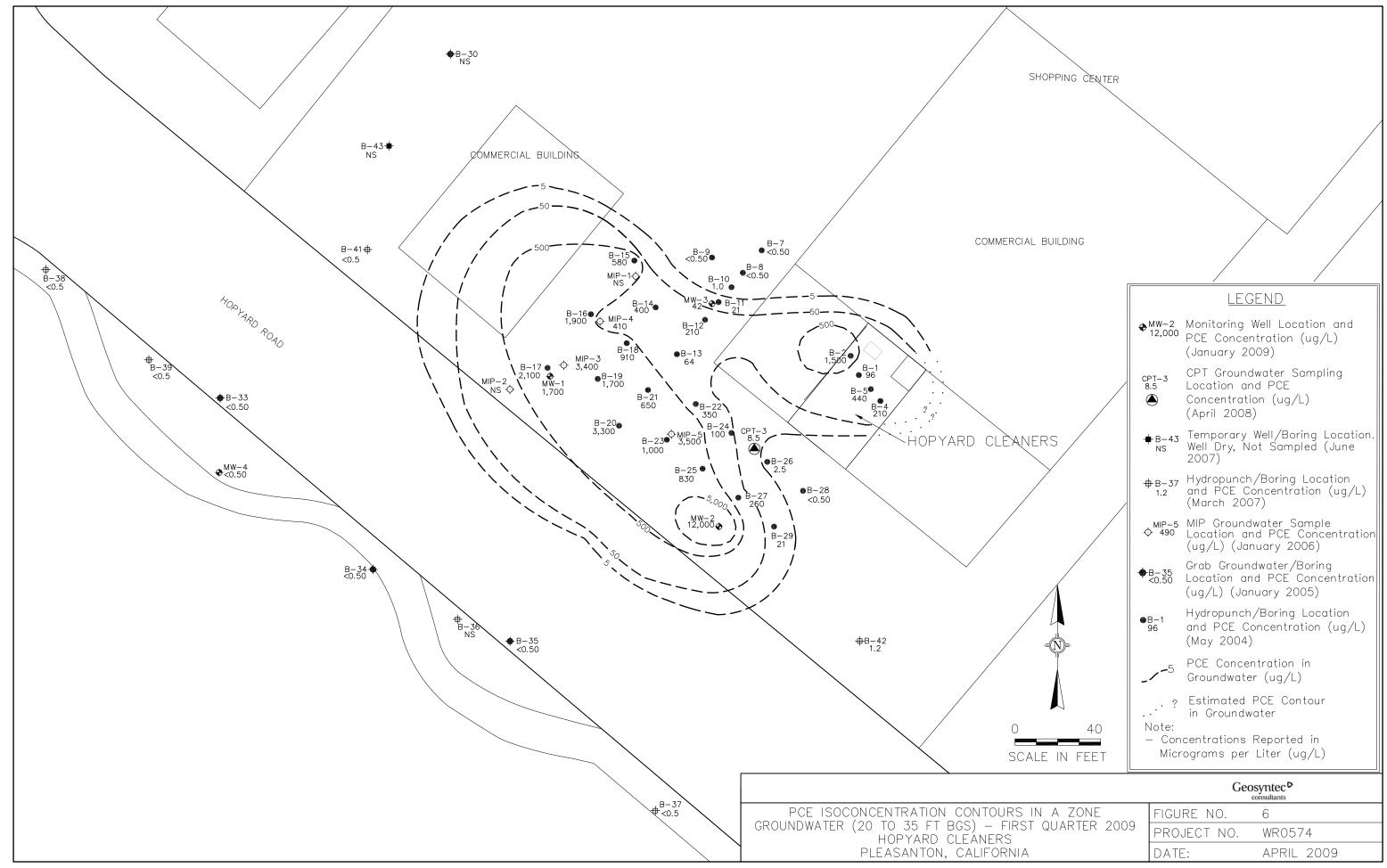
FIGURE NO.	1
PROJECT NO.	WR0574
DATE:	APRIL 2009

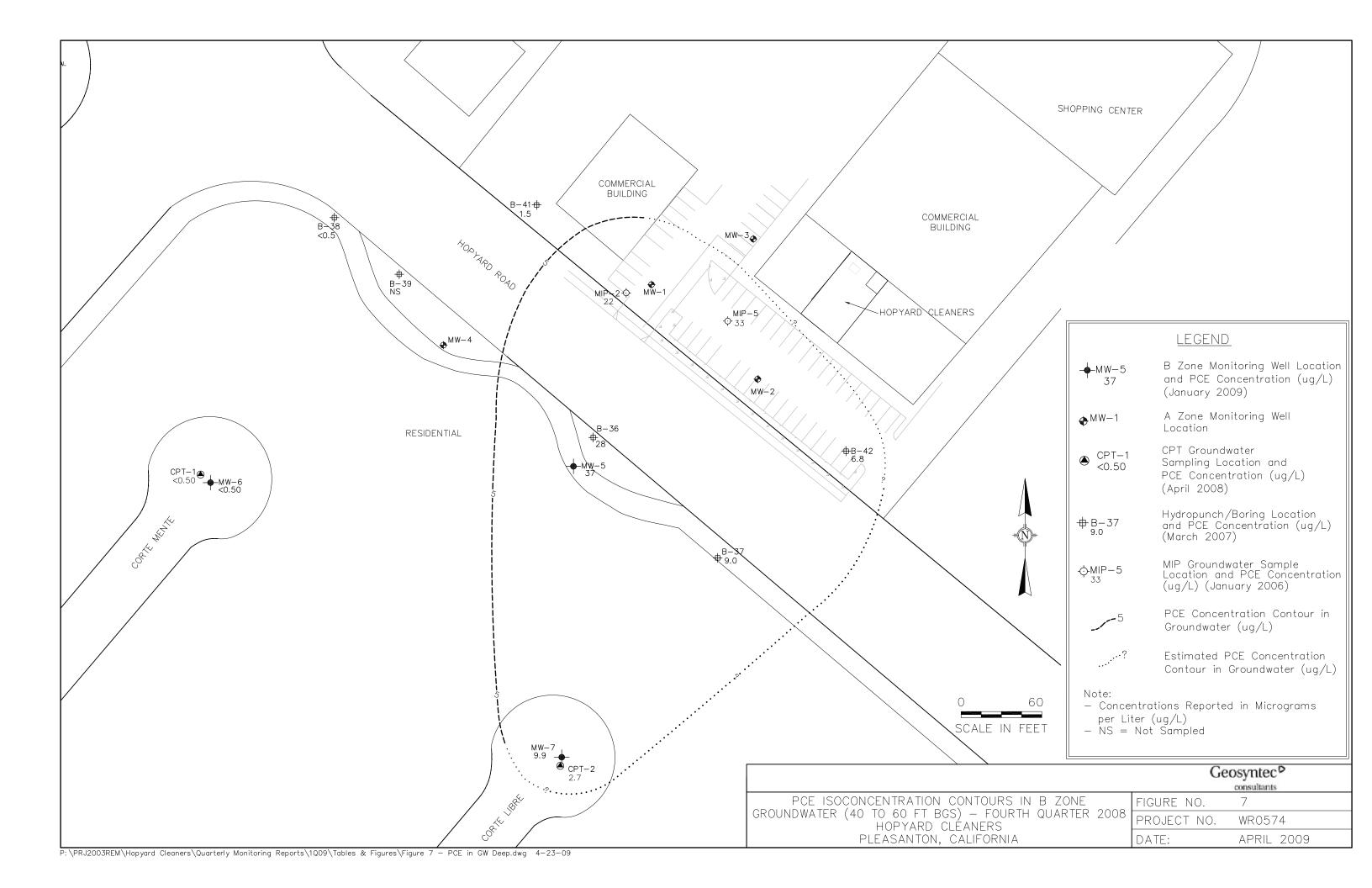


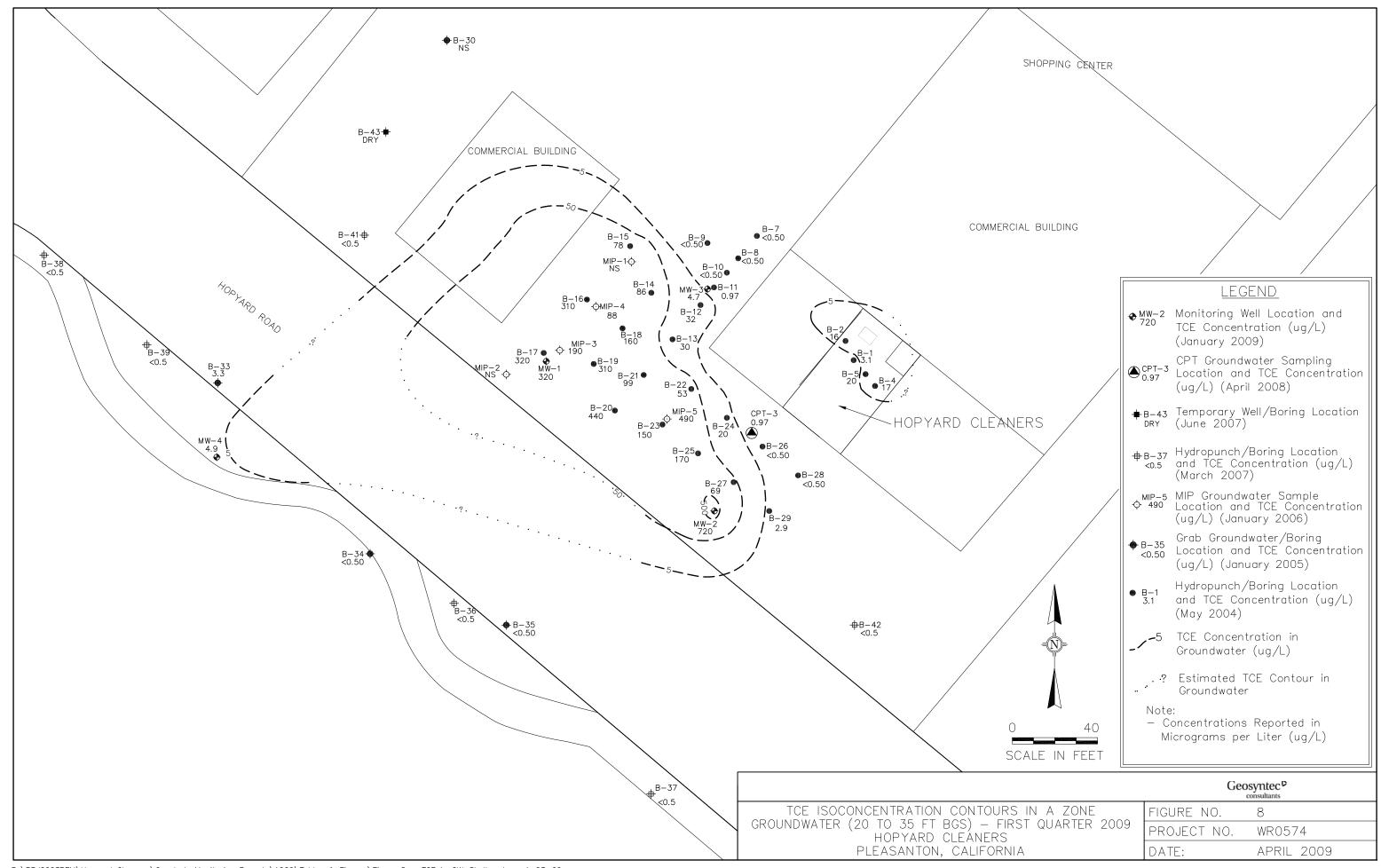


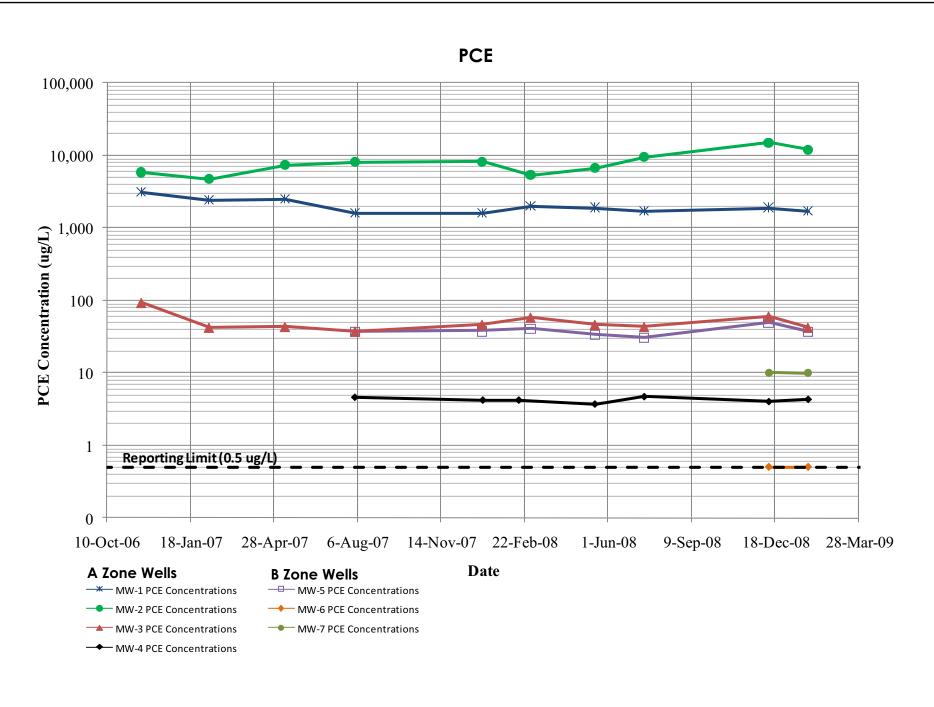


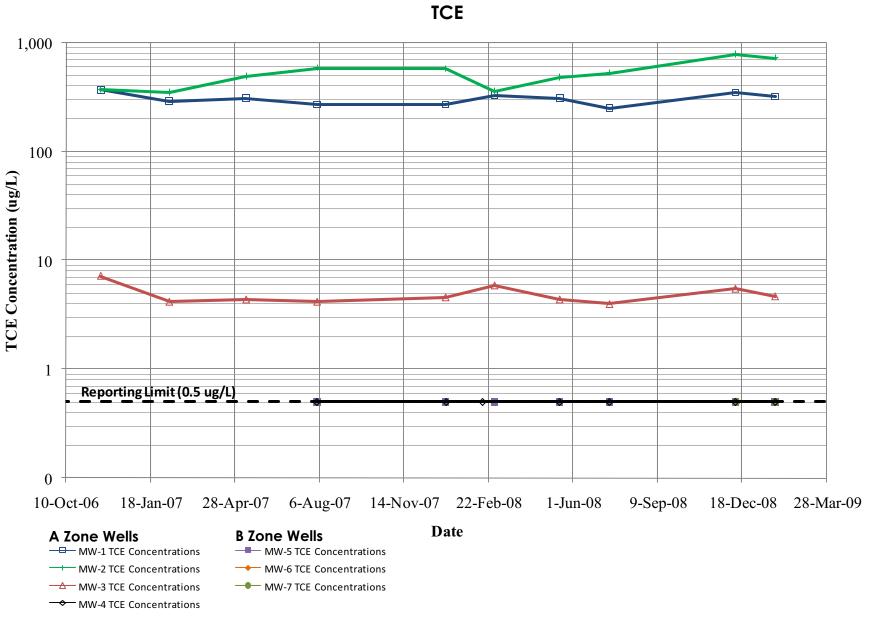












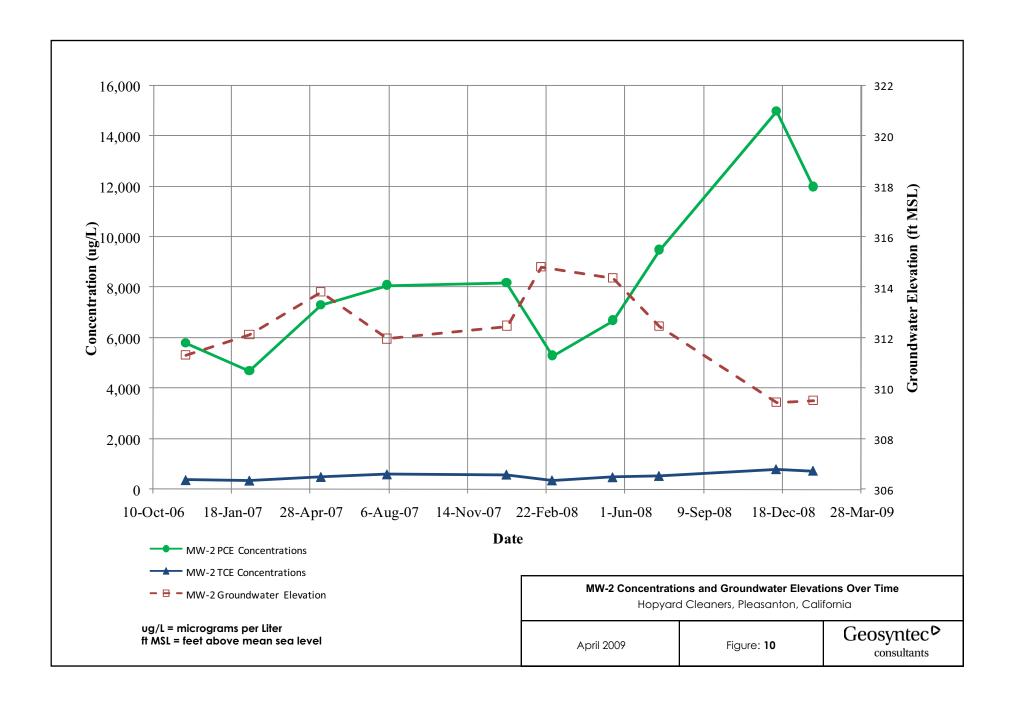
PCE and TCE Groundwater Concentrations Over Time
Hopyard Cleaners, Pleasanton, California

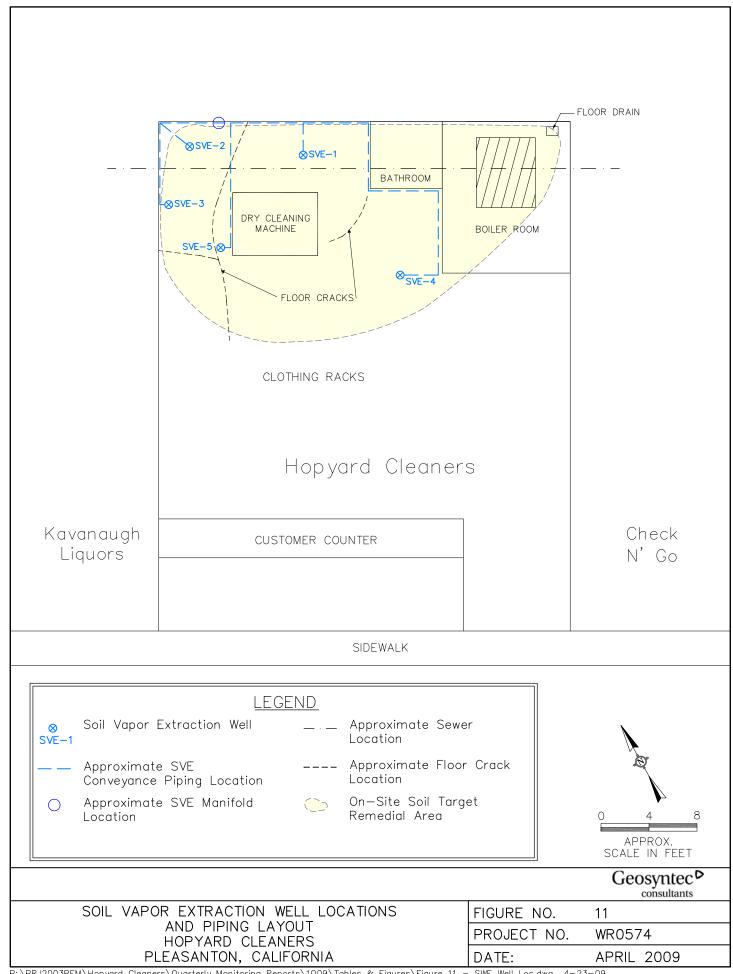
April 2009

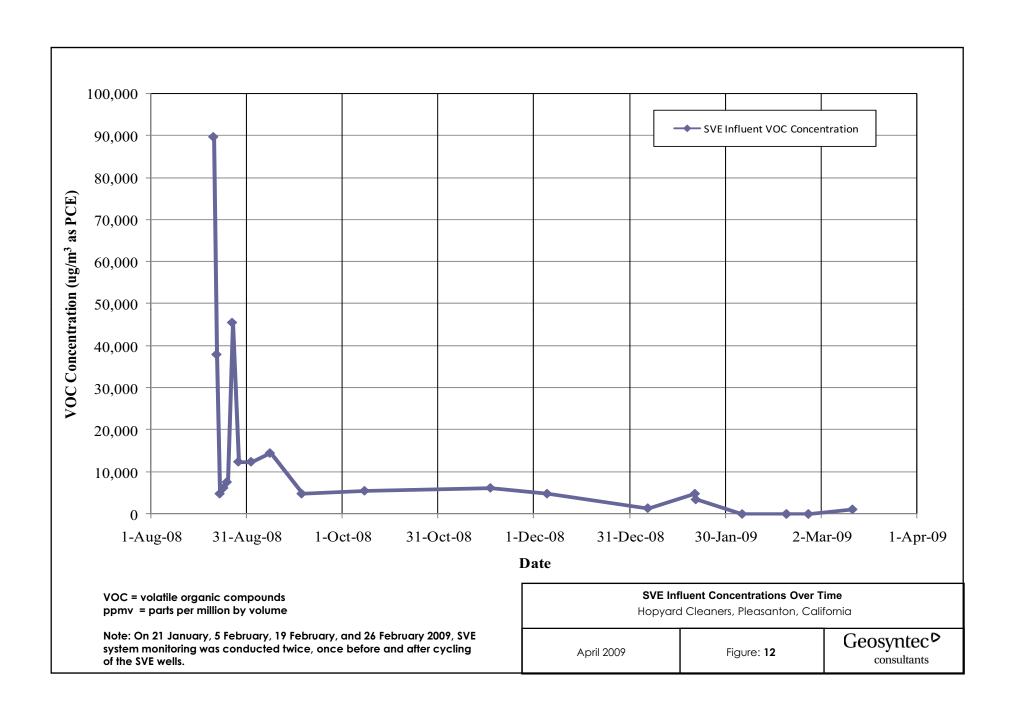
Figure: 9

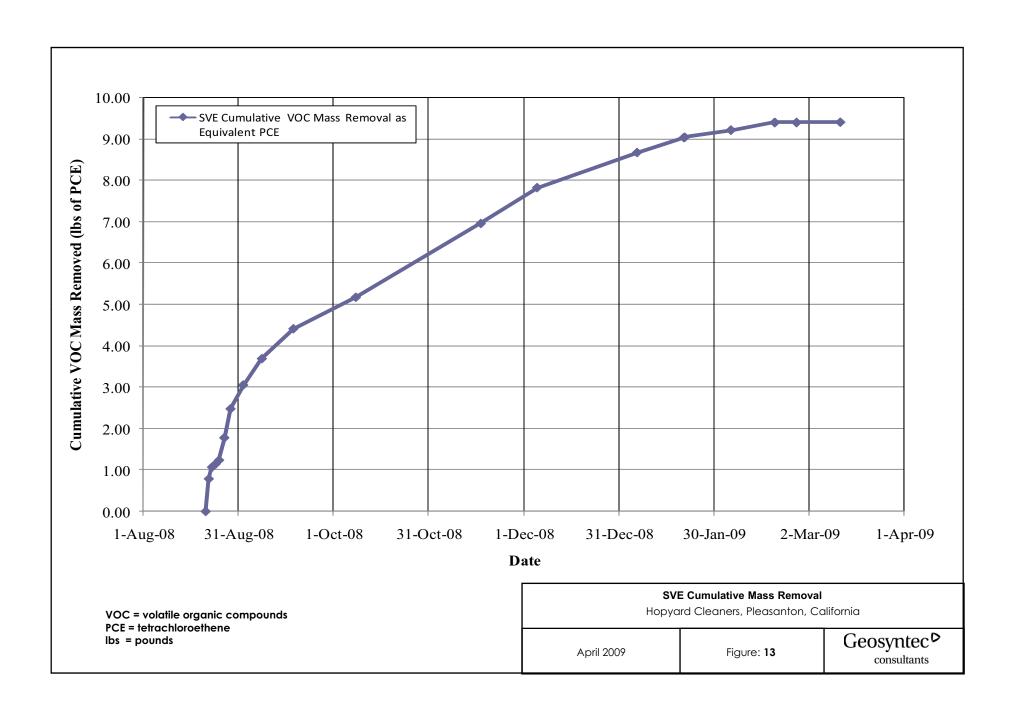
Geosyntec consultants

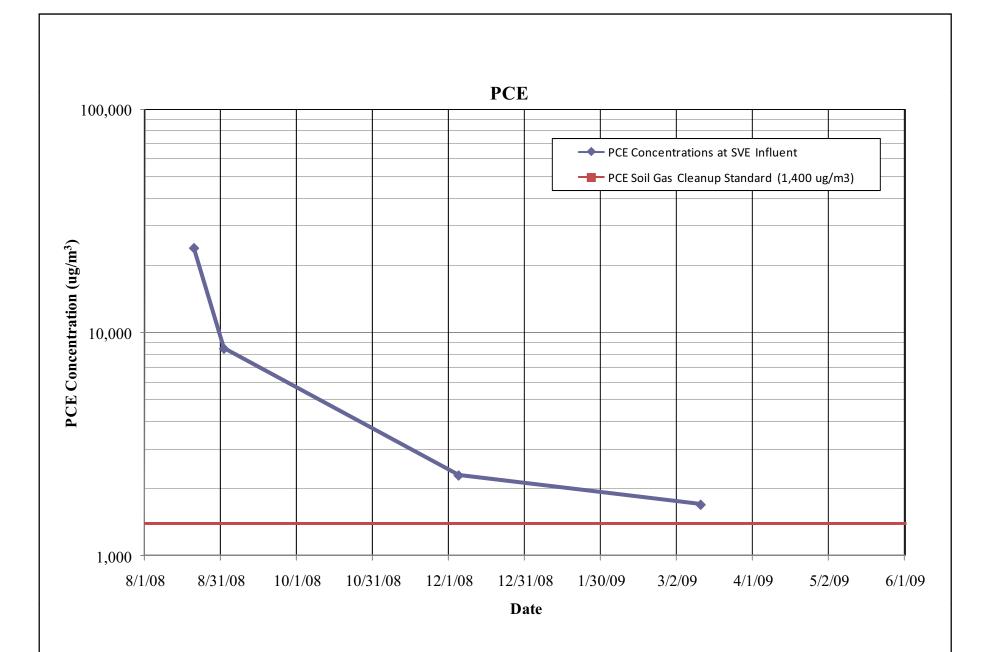
ug/L = micrograms per Liter

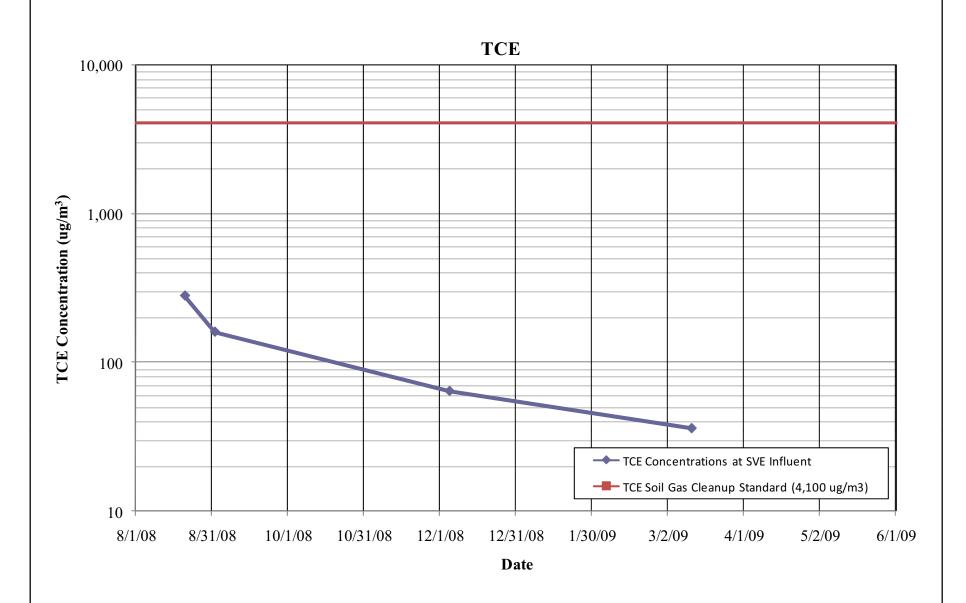












PCE = tetrachloroethene
TCE = trichloroethene
ug/m3 = micrograms per cubic meter
Soil Gas Cleanup Standards are 1,400 and 4,100 ug/m³ for PCE
and TCE, respectively, and are from the California Regional Water
Quality Control Board Order No. R2-2008-0032.

SVE Influent PCE and TCE Concentrations

Hopyard Cleaners, Pleasanton, California

April 2009

Figure: 14

Geosyntec consultants

CONTROLS Anaerobic Active Control Anaerobic Sterile Control 0.014 0.014 Concentration (mmoles/pottle) 0.012 0.008 0.006 0.006 0.012 Concentration (mmoles/bottle) 0.01 0.008 0.006 0.004 0.002 0 10 20 30 40 50 10 20 30 40 50 Time (Days) Time (Days) **DONOR AMENDMENT** EVO (Newman Zone®) Lactate 0.014 0.014 0.012 0.012 Concentration (mmoles/bottle) Concentration (mmoles/bottle) 0.01 0.01 0.008 0.008 0.006 0.006 0.004 0.004 0.002 0.002 0 20 30 40 50 20 Time (Days) 40 50 0 10 Time (Days) **BIOAUGMENTED DONOR AMENDMENT** EVO (Newman Zone®) & KB-1® Lactate & KB-1® 0.014 0.014 0.012 0.001 0.008 0.008 0.006 0.012 (mmoles/bottle) 0.01 Day 28: Day 28: Bioaugmented Bioaugmented with KB-1 0.008 Concentration (0.000 6.0004 0.004 0.002 0.002 0 0 10 20 30 40 40 50 10 30 Time (Days) Time (Days) →PCE -TCE -VC ---Ethene **Preliminary EISB Treatability Study Results** PCE = tetrachloroethene Hopyard Cleaners, Pleasanton, California TCE = trichloroethene cis-1,2-DCE = cis-1,2-dichloroethene VC = vinyl chloride Geosyntec^D Figure: 15 **April 2009** consultants

APPENDIX A

Environmental Sampling Services Field Report



February 3, 2009

Ms. Melissa Asher Senior Staff Engineer GeoSyntec Consultants 475-14th Street, Suite 450 Oakland, California 94612

SUBJECT: January 2009 Quarterly Groundwater Monitoring Event for Hopyard Cleaners, Pleasanton, California

Dear Ms. Asher,

Please find enclosed the Field Activity Report for the quarterly groundwater monitoring event at 2771 Hopyard Road that occurred January 26, 2009.

If you have any questions or concerns regarding this Field Activity Report, please do not hesitate to call me.

Sincerely,

Environmental Sampling Services, LLC

Jacqueline Lee Manager

Enclosure

FIELD ACTIVITY REPORT FOR

JANUARY 2009
QUARTERLY GROUNDWATER
MONITORING EVENT

HOPYARD CLEANERS 2771 HOPYARD ROAD PLEASANTON, CALIFORNIA

> Prepared for: GeoSyntec Consultants 475-14th Street, Suite 450 Oakland, California 94612

> > Date Prepared: February 2, 2009



FIELD ACTIVITY REPORT FOR

JANUARY 2009
QUARTERLY GROUNDWATER
MONITORING EVENT

HOPYARD CLEANERS 2771 HOPYARD ROAD PLEASANTON, CALIFORNIA

Task: Quarterly Groundwater Sampling Event

ESS Personnel: Jacqueline Lee and Stephen Penman

Geosyntec Personnel: Melissa Asher Date of Activities: January 26, 2009

Decontamination Procedures

All downhole equipment was cleaned with Liqui-Nox® laboratory-grade soap, potable water, and rinsed with distilled water prior to use and between each monitoring well.

Groundwater Level Measurements

Depth to groundwater for seven monitoring wells were measured and recorded following atmospheric equilibration of approximately thirty minutes. All readings were performed with a Solinst® Water Level Meter, Serial Number 49914, and referenced to the surveyor's mark or north rim at the top of PVC well casing (Table 1). Three successive readings that agreed to within one-hundredth of a foot determined depth to groundwater.

Field Equipment and Calibration

pH, Specific Conductance, Temperature, Dissolved Oxygen, and Oxidation Reduction Potential (ORP) were monitored with a YSI® Multi-parameter meter equipped with an in-line flow through cell. Turbidity readings were measured with a HF Scientific® Turbidity meter.

Equipment calibration was performed in accordance with the instrument's calibration and operating procedures. Calibration was performed prior to any monitoring activities (see Daily Equipment Calibration Sheet).

Solution standards: pH 4, 7, and 10, Specific Conductance @ 1,000 uS/cm, and Zobell for ORP were used for calibration purposes. Dissolved Oxygen was calibrated to air. Turbidity was checked against a 0.02 NTU standard. All equipment calibrated and functioned properly during monitoring activities.



Seven monitoring wells contained Passive Diffusion bags (PDBS), Samples for Volatile Organic Compounds were collected from each PDB.

During sample collection, all 40-ml VOA sample containers were filled while minimizing aeration or degassing. Each sample container was inverted and tapped lightly to check for air bubbles. The absence of air bubbles indicated a successful seal.

Low-Flow Well Purging & Sampling Procedures

A peristaltic pump and new pump tubing was used to purge and sample MW-1 through MW-3. Each monitoring well was purged at a rate no greater than 500-ml per minute until water quality parameters stabilized for three consecutive readings.

Each well was purged following procedures recommended by the United States Environmental Protection Agency Low Flow (Minimal Drawdown) Ground-Water Sampling Procedures, dated April 1996. Each well was purged at a rate no greater than 500-ml per minute until water quality parameters stabilized.

During low-flow purging activities, pH, Specific Conductance (uS), Temperature (Celsius), Dissolved Oxygen (mg/L), Oxidation/Reduction Potential (mV), and physical characteristics such as pumping water level, color, and odor (see Water Quality Sample Log Sheets) were monitored and recorded (see Water Quality Log Sheets).

EPA stabilization guidelines for low-flow sampling were used. Stabilization was achieved after the water quality parameters stabilized for three successive readings. The following criteria were used: \pm 0.1 for pH, \pm 3 % for Specific Conductivity, \pm 10% for Dissolved Oxygen, and \pm 10 mV for ORP.

Three criteria were used to determine Turbidity stabilization:

- 0-10 NTUs, no criterion
- 10-50 NTUs, ± 5 NTUs
- more than 50 NTUs, ± 10%

Groundwater samples were collected immediately following stabilization of water quality parameters by disconnecting the tubing from the flow through chamber.

Low flow sampling procedures was used to collect samples for Anions, Gene-Trac Analyses, and Feasibility Analysis.

Sample Handling

All sample labels were completed with waterproof ink and affixed to sample containers.

All sample containers were wiped dry, sealed in Ziploc® bags, and placed a chilled cooler for storage and shipment to the laboratory.

Following completion of groundwater sampling, seven new PDBs were installed. PDB was attached to the middle position for MW-5, MW-6, and MW-7.

Laboratories

TestAmerica of Pleasanton, California provided Trip Blank, sample containers with appropriate preservative, and conducted VOC and Anions (Nitrate, Nitrite, Chloride, and Sulfate) analyses.

SiREM Laboratory of Ontario, Canada provided sample containers with appropriate preservative for Initial VOCs and Feasibility analyses. Gene-Trac ample containers were provided by TestAmerica.



SiREM Laboratory of Ontario, Canada provided sample containers with appropriate preservative for Initial VOCs and Feasibility analyses. Gene-Trac ample containers were provided by TestAmerica.

Sample Containers

Each VOC sample set was contained in three, 40-ml VOA clear glass containers preserved with hydrochloric acid.

Each Anions sample was contained in a non-preserved 500-ml plastic container.

Each Gene-Trac sample was contained in a non-preserved, one-liter plastic container with minimum headspace.

The Feasibility Study sample was contained in two, non-preserved, 4-liter plastic containers with minimum headspace.

Quality Assurance /Quality Control Samples

All QA/QC samples were submitted to TestAmerica and SiREM for analysis.

One Trip Blank set was stored in the cooler with TestAmerica samples throughout the sampling event.

One blind PDB duplicate set was collected from MW-2 and labeled "MW-DUP @ 08:00". Due to the limited volume in the PDB, a total of four VOAs were filled for both the primary and duplicate sample set. The Anion and Gene-Trac Duplicate was also labeled "MW-DUP" with "14:30" as its fictitious sample time.

One equipment blank was requested but not collected by ESS.

No other QA/QC samples were requested.

Chain of Custody (COC) Form

All sample handling was conducted under standard chain of custody procedures. The COC included: sampler's name and signature, sample identification, sample date and time, and analysis request section.

Shipment of Samples

Samples for TestAmerica were relinquished January 26, 2009. Samples designated for SiREM was relinquished to Geosyntec for shipment.

Storage of Investigative Derived Wastewater (IDW)

Approximately 30 gallons of purged groundwater and decontamination water generated from this sampling event were stored an existing, labeled 55-gallon drum. The drum is stored inside the treatment system enclosure.

Comments

SiREM inadvertently omitted the one-liter plastic containers for Gene-Trac Method analyses. The Gene-Trac containers were obtained from TestAmerica.

ESS inadvertently did not collect the equipment blank.



With the exception of the above omission, the January 2009 monitoring event was performed as requested by Geosyntec.

Environmental Sampling Services, LLC

Jacqueline Lee Manager

Attachments:

Table 1: Summary of Groundwater Monitoring Event

Water Sample Log Sheets Equipment Calibration Sheet

Chain of Custodies



WATER	R QUALIT	Y SAMPLE	LOG SH	EET	WELL IDEN	ITIFICAT	ION: MV	V-1 DAT	E: 1/26	/09
Project I	Name: <u>Hop</u>	yard Cleane	rs Pleasan	ton, CA P	roject Task: Qu	arterly M	onitoring	Project/Tas	k No. WF	R0574
Project I	Manager: I	Melissa Ashe	er - Geosyr	itec Cons.	Lab: STL San	Francisco	Weath	er Condition	s: Clepr	£0001
Well De	scription:	3.5" 4"	5" 6" Of	her:	Well Type: 0	Sta	inless Ste	eel Other:		
					Type of lock					
					7-ft.(BTOC)					
					entrifugal Pun					
					Bailer Line:			/ Dedicated		
					Tap Water DI					
					ap Water DI I			tonyo		
					PE Bailer (Per					
					S - 05F1258A	9 / 600X	L 319340	R - 00C1522	2	
A		ion: See Da	The state of the s			V 05710	010			
					ial No.: 25083					70
					water) x "K" = 2					(BTOC)
					" = .653 (4" well)					
	0.100	(Z Holly K	resident to the	The Wall Street Walls	QUALITY PAR	AND THE SECTION	chesto	i) K - 1.4	o (o weii)	
					Specific			Dissolved	Water	
Date	Time	Discharge	pН	Temp.	Conductance	Turbidity	Redox	Oxygen	Level	Color
		(Liters)	DANSON I	(°C)	mS (LS)	(NTU's)	(mV)	(mg/L)	(BTOC)	
1/	4000000000		+/- 0.1		+/- 3%	+/- 10	+/- 10	+/- 10%		. b. L.L.
1/20/08	11:03	Initial	7.06	18.28	1177	16.4	99.3	1.75	16.83	Closely
1/26/09	11:00	0.5	7.00	18,95	1167	8.21	57.8	0.55	16.84	Clear
	11:09	1.0	7.00	19.42	1163	5.79	37.1	0,42	16.85	1.1
	11:11	1.5	6.99	19.92	1158	3.91	36,5	0.36	16.86	. 1
	11:14	2.0	6.99	20.16	1161	3.06	52.6	0.20	16.86	11
	11:16	2.5	6.98	20.26	1161	2.93	56.6	0.16	16.86	.1
	11:18	3.0	6.98	20,28	1163	2.38	61.1	0.15	16.86	И
	11:21	3.5	6.98	20.36	1164	2.19	63.5	0.15	16.86	11
	1	4.0								
Total Dis	charge:	39 Li	ters	2-27-15	Casing Volum	es Remov	ved: NA			
			0.00000	5 Gallon Dr	um(s) Poly T			vstem Oth	er.	
		1: 1/26/09			Analysis:					
					Trac analysis -			O/IS W/I (OI		
					S/MSD Equip			ld Blank 1	ah Salit	
					WHa) Whole					(ttagen)
Comme	no. Pops	concered	2 1.23	CO ADDIS	744) 1011 16	wer Piter	. 12 2 00	Aprily VOC	5: 10.71	C-777710
					1-4-6	C)		1//		
Recorder 668	o Alhambi	nen Penman	/ Cacki Le	Signati	553-6105 · (925] 372	-8108	Fax: [925]	Page 1 of 372-670	J ₅



WATER	R QUALITY	Y SAMPLE	LOG SH	EET	WELL IDEN	ITIFICAT	ION: MV	V-2 DAT	E: 1/26	109
Project I	Name: Hop	yard Cleane	rs Pleasan	ton, CA Pr	roject Task: Qu	uarterly M	onitoring			
					Lab: STL San					
Well De	scription: 2	3.5" 4"	5" 6" Ot	ther:	Well Type:	PVC) Sta	inless Ste	eel Other:	SC CHOSCONIA	
ls Well S	Secured?(Y	es/No Bo	It Size: 9/1	16"	Type of lock	/ Lock nu	mber: Ma	ster		
Observa	ations / Con	nments: set	pump inta	ke @ 25,3	ft.(BTOC)	Screen I	nterval: 2	20' to 30'		
					entrifugal Pun	p Perist	altic Pum	Other:		
		lew) Cleane			Bailer Line:			Dedicated		
					ap Water DI					
					ap Water DI I					
					PE Bailer (Pe					
					S - 05F1258A	D/ 600X	L 319340F	R - 00C1522	2	
		on: See Da								
					ial No.: (25083					10 m
) Water Leve					(BTOC)
10 -	("K"= 0.163 /	"K" ((low "C	DIW) = 14	(II.of)	ص = "Water) x "K"	2.3 (G	als./CV) x [NA (No. of C	V) = NA (C	Bals.)
	(1 - 0.103)	z weil) K		Company of the Compan	" = .653 (4" well	destruction of the contract) "K" = 1.4	6 (6" well)	
			FIEL	D WATER	QUALITY PAR	RAMETER	RS			
Date	Time	Discharge	pН	Tomo	Specific	Totalida	Deden	Dissolved	200	
Date	Time	(Liters)	рп	Temp. (°C)	Conductance mS (uS)	(NTU's)	Redox (mV)	Oxygen (mg/L)	Level (BTOC)	Color
		(=10.0)	+/- 0.1	(0)	+/- 3%	+/- 10	+/- 10	+/- 10%	(6100)	1
26/09	13:32	Initial	7,04	18.90	1495	42,3	110.7	2,45	16,33	Sight-
1	13:35	0.5	6.95	19.11	1495	38.0	119.2	0,53	16.34	F1 '
	13:37	1.0	6.92	19.20	1496	31.8	122.1	0.41	16.34	+1
	13:39	1.5	6.91	19.29	1496	27.7	121.0	0.31	16.34	+1
	13:41	2.0	19.0	19.37	1495	19.6	120.6	0.34	16.34	н
	13:43	2.5	6.91	19.47	1494	9.92	119.2	Transport N	16.34	Clear
	13:45	3.0	6.91	19.50	1493	7.64	117.9	0.20	16.34	11
	13:47	3.5	6.91	19.56	1490	6.96	116.7	0.18	16.34	II.
\checkmark	13:49	4.0	6.91	19.54	1492	4.95	Sec. 155 7	121	16.34	ч
otal Dis	charge:	6.4 Li	ters		Casing Volum	es Remov				
ethod o	of disposal of	of discharge	d water: (5		um() Poly T			stem Oth	er:	
ate/Tim	e Sampled	1/25/09	@ 14:0	0	Analysis: \					
nions (3	300.0) - 1 50	00ml Poly n/	p; Dhc/VC	r by Gene-T	rac analysis -	11:4.50	- 1 - 1 A .	F-61.72-6	00	heal 42
A/QC:	MW-DUP	@ 14:3	o Du	olicate) MS	/MSD Equipr	nent Ring	eate Fiel	d Blank L	b Solit	Per Peri LY
ommen	ts: PDB<	Collected	@ 9'Sh -	ה"מומי- מומי	(10:50)" A	UDAC W/ L	eate Fiel	o Didlik La	ibo lasal	- 11.13
le wa	£ 49914.	TO SECIEN	1130	1-100-001	(10,00) W	TOPA 1 P	per s	ample .V	ISPSI TIV	- 1947
/ Cal/act	111111				-			200		
	-	en Penman		_)	-	> lif	_	



WATER	QUALITY	SAMPLE	LOG SHE	ET	WELL IDEN	TIFICATI	ON: Mu	1-2 Cont	inued Pa	ge 2 of 2
Project N	lame: Hop	yard Cleane	rs Pleason	nton, ca	Project No.:	NR057	4			
			FIELD WA	TER QUA	LITY PARAME	TERS CO	NTINUE	D		
Date	Time	Discharge (Liters)	pH ± 0.1	Temp. (°C)	Specific Conductance mS uS ± 3%	Turbidity (NTUs) ±10 NTU's	Redox (mV) ± 10mv	Dissolved Oxygen (mg/L) ± 10%	Water Level (BTOC)	Color
1/26/09	13:53	4.5	6.91	19,46	1488	4,86	116.7	0.14	16.34	clear
	13:55	5.0	6.91	19,48	1489	4.73	117.0	0.12	16.34	u
	13:57	5.5	6.91	19.40	1490	4.61	117.6	0.12	16.34	
1	13:59	6.0	6.91	19.35	1490	3.69	118.2	0.12	16.34	41
		6.5								
		7.0								
		7.5								
		8.0								
		8.5								
		9.0								
		9.5								
		10.0								
		10.5								
		11.0								
		11.5								
		12.0								
		12.5								
		13.0								
		13.5								
		14.0								
Total Dis Commer		6.4 Li	ters		Casing Volum	es Remov	ed: <u>NA</u>			
Recorde	d by: Steph	en Penman	Jacki Lee) Signatur	re: Atyl F	2		+M		



WATER	R QUALITY	SAMPLE	LOG SHE	ET	WELL IDEN	TIFICAT	ON: MV	I-3 DAT	E: 1/26/	109
Project Project Project Well De Is Well : Observa Purge N Pump L Method Method Samplin YSI Mut Equipmed Method Water L TD =	Name: Hope Manager: Manager: M	yard Cleaner Melissa Ashe 3.5" 4" es/ No Bol nments: set Teflon / PE lew/ Cleaner Pump: NA Bailer: NA PDBS Disp Meter/Prob ion: See Da Water Leve t (DTW):	rs Pleasant er - Geosyn 5" 6" Otl It Size: 9/1 pump intak Disposable Alconox It Alconox It Teflon Ba er Serial No ally Equipment It Slope In It It It	on, CA Protec Cons. Inter: 6" 6 Bailer Cons. Inter: 7 Bailer Cons. Inter: 7 Bailer Cons. Inter: 8 Bailer Cons. Inter: 8 Bailer Cons.	oject Task: Quab: STL San Well Type (F Type of lock Int.(BTOC) entrifugal Pum Bailer Line: (N Tap Water DIF Tap Water Leve Tap Water Leve Tap Water Leve Tap Water Leve	Francisco Franci	onitoring Weather Inless Stember: Ma Interval: 2 Inter	Project/Tasker Condition rel Other: ster 20' to 30' Other: Dedicated rer: R - 00C1522 Reading: NA (No. of CV	k No. WR s: Clear 1	(BTOC)
	K = 0.163	2 welly K			" = .653 (4" well) QUALITY PAR) "K" = 1.4	6 (6" well)	
Date	Time	Discharge (Liters)	pH +/- 0.1	Temp.	Specific Conductance mS us +/- 3%			Dissolved Oxygen (mg/L) +/- 10%	Water Level (BTOC)	Color
1/26/09	09:41	Initial	6.73	15,80	1627	4.38	180.7	1.76	17.33	Clear
	09:45	0.5	6.9.3	16.37	1638	4.01	169.8	0.63	17.34	ri .
	09:48	1.0	6.88	16.89	1635	2,30	162.6	0,50	17.36	11
	09:51	1.5	6.88	17,32	1636	194	157.7	0.27	17.36	41
	09:54	2.0	6.87	17.56	1634	1.85	155.2	0.24	17.37	*1
	09:57	2.5	6.83	17.77	1633	1.81	153.0	0.24	17.37	11
	10:00	3.0	6.83	17.93	1633				17.37	.1
Method Date/Tir Anions (ne Sampleo 300.0) - 1 5	of discharge ाः <mark>। /2<i>६</i>/<i>व</i>9 00ml Poly n</mark>	@(0 :_/p; Dhc/VC	o > r by Gene-	Casing Voluments) Poly 1 Analysis: Trac analysis - S/MSD Equip	Tank Tre VOCs (82	eatment S 60B) - 3 \ Poly N/p	OAs w/HCI		
Comme	nts: PDB:		le q:is (3 VOAs 47	HCI). Wir les					705



WATER	QUALITY	Y SAMPLE	LOG SHE	ET	WELL IDEN	TIFICATI	ON: MV	I-4 DAT	E: 1/26	109
Project N	ame: Hop	yard Cleaner	s Pleasant	on, CA P	roject Task: Qu	arterly Mo	nitoring	Project/Tas	k No. WR	0574
THE RESERVE OF THE PARTY OF THE					Lab: STL San					
					Well Type:					
					Type of lock					
Observat	ions / Con	nments: set	pump intak	e@ NA	ft.(BTOC)	Screen In	iterval: 2	20' to 30'		
Purge Me	ethod: (NA	Teflon / PE	Disposable	e Bailer (Centrifugal Pum	p Perista	altic Pump	Other:		
Laborate and School of Street		lew / Cleane			Bailer Line:			Dedicated		
	-			100	Tap Water DI F					
Contract of the same				A STATE OF THE PARTY OF THE PAR	Tap Water DIF					
A strictly of the con-		-			PE Bailer Per					
					PS - 05F1258AH	1 / 600XI	_ 319340F	R - 00C1522	2	
		ion: See Da	Salind 1			(49914))	1.1. s 1507 17 17 60 0 61		
			A STATE OF THE PARTY OF THE PAR		rial No.: 25083					
Water Le	vel at Star	t (DTW): _1	7.860 8:	43 (BTO	c) Water Level	Prior To	Sampling	17.86 e	10:50	(BTOC)
					water) x "K" =					
-	K"= 0.163	(2" well) "K"			(" = .653 (4" well)) "k" = 1.4	6 (6" well)	
			FIEL	D WATER	QUALITY PAR	RAMETER	RS			
-0.00 m					Specific		-	Dissolved	Water	
Date	Time	Discharge (Liters)	рН	Temp. (°C)	Conductance mS uS	(NTU's)	Redox (mV)	Oxygen (mg/L)	Level (BTOC)	Color
		(Liters)	+/- 0.1	(0)	+/- 3%	+/- 10	+/- 10	+/- 10%	(6100)	
1/26/09	NA	Initial	NA	AU	NA	NA	NA	NA	NA	NA
1	1	0.5	1	T T	1	1	1	1	1	ľ
		1.0								
		1.5						-		
-		2.0								
	-	2.5	-							
									\vdash	
		3.0					-			
		3.5	1	1		1	4	1	1	1
•	- V	4.0								
The State of the		0 Li	2012 Table 1		Casing Volum					
					rumos) Poly T					
Date/Tim	e Sampleo	1: 126/09	_@_10:	52	Analysis:	VOCs (82	60B) - 3 \	OAs w/HCI		
99443355C4840563	None	_ ~			S/MSD Equip				ab Split	
Commen	ts: Had	to pump	out su	rface w	ater from 1	well a	nnulus.			
		A 150								
Recorded	by: Step	hen Penman	/Qacki Le	e) Signat	ture:				Page 1 of	l
668	0 Alhamb	ra Ave., #10	12 · Marti	nez, CA 9	4553-6105 •	(925) 37	2-8108	Fax: [925		



WATER	QUALIT	Y SAMPLE	LOG SHE	EET	WELL IDEN	TIFICATI	ON: MV	/-5 DAT	E: 1/26/0	9
Project N	ame: Hop	yard Cleaner	s Pleasant	on, CA Pr	oject Task: Qu	arterly Mo	nitoring	Project/Tasl	k No. WR	0574
Committee of the Commit			The state of the s	The state of the s	Lab: STL San	A STATE OF THE PARTY OF THE PAR		B. 시작 아래 중요하다 네 모든 1.0 Head		100 mg// 100
Well Des	cription:	3.5"	5" 6" Ot	her:	Well Type:(F	VC) Sta	inless Ste	el Other:		
Is Well S	ecured?	es No Bolt	Size: <u>9/4</u>	Ge 15/16"	Type of lock	Lock nur	nber: Ma	ster		
Observat	ions / Con	nments: set	pump intal	e@ NA	ft.(BTOC)	Screen In	iterval: 2	20' to 30'		
Purge Me	thod: NA	Teflon / PE	Disposabl	e Bailer C	entrifugal Pum	p Perista	altic Pump	Other:		
Pump Lin	es: NA N	New / Cleaned	d / Dedicat	ted	Bailer Line: (A New /	Cleaned /	Dedicated		
		700			ap Water DIF		Account the second			
				77.	ap Water DIF					
Contract of the Contract of th					PE Bailer Per			trief of the Sources of the original		
					S - 05F1258AF	4 / 600XL	_ 319340F	R - 00C1522	2	
Equipme	nt Calibrat	ion: See Dai	ly Equipm	ent Calibra	tion Sheet	49914	av.			
Talanta and the property of the					ial No.: 25083			THE RESERVE OF THE PARTY OF THE	and the same of the same	
) Water Level					
					water) x "K" = <u></u>					als.)
	K"= 0.163	(2" wellD "K"	= 0.50 (3.5	"well) "K	" = .653 (4" well)	"K" = 1.	02 (5" well) "k" = 1.4	6 (6" well)	
			FIEL	D WATER	QUALITY PAR	RAMETER	RS			
10/21/17	Time Discharge pH Temp. Conductance Turbidity Redox Oxygen L									re coercine
Date	Time	0.000	pН	100000000000000000000000000000000000000			120 120 120	1, 177 VICE (400) 110	Level	Color
		(Liters)	+/- 0.1	(°C)	mS uS +/- 3%	(NTU's) +/- 10	(mV) +/- 10	(mg/L) +/- 10%	(BTOC)	
1/26/09	NA	Initial	NA	NA	MA	NA	NA	NA	NA	AA
120101	1	0.5	1	1	1	1	1	1	1	/
			-						\Box	_
		1.0	_						\vdash	-
		1.5				_			\vdash	-
		2.0	_						-	
\Box		2.5								
		3.0								
		3.5								
1	1	4.0	1	1	4	7	4	1	1	1
Total Disc	charge:	O Li	ters		Casing Volum	es Remov	ed: NA			
			d water: 5	5 Gallon Dr	rum(s) Poly T			vstem Oti	ner:	
		1: 126/09						OAs w/HCl		
							, , ,			
OA/OC:	None	@ -	Du	nlicate MS	S/MSD Equipr	nent Rine	eate Fie	ld Blank I	ah Solit	
				phodic: WK	JAMOD Equipi	Herit Falls	cate i ic	id Dialik L	ab Opin	X
Commen										
A. Carrier and Control				_	\ ./ -					
		hen Penman			1553/6105 ·	925137	- 2-81118 1	Fax: 1925	Page 1 of	75



WATER	QUALITY	SAMPLE	LOG SHE	ET	WELL IDEN	TIFICATI	ON: MW	-6 DAT	E: 1/26,	/09
Project N	lame: Hop	yard Cleaners	s Pleasant	on, CA Pr	oject Task: Qu	arterly Mo	nitoring F	Project/Task	No. WR	0574
A 17 10 10 10 10 10 10 10 10 10 10 10 10 10					Lab: STL San					
					Well Type: €					
					Type of lock					
	~				ft.(BTOC)					
Purge Me	ethod: NA	Teflon / PE	Disposable	e Bailer C	entrifugal Pum	p Perista	Itic Pump	Other:		
Pump Lir	nes: NAN	lew / Cleaned	d / Dedicat	ed	Bailer Line:	A New /	Cleaned /	Dedicated		
Method o	of Cleaning	Pump: (A)	Alconox I	_iqui-nox 1	Tap Water DIF	Rinse Oth	er:			
Method o	of Cleaning	Bailer: NA	Alconox I	iqui-nox 1	ap Water DIF	Rinse Oth	er:			
Sampling	g Method:(EDBS) Disp.	Teflon Ba	iler Disp.	PE Bailer Per	istaltic Pu	ımp Oth	er:		
YSI Muti	-Paramete	r Meter/Probe	e Serial No	.: 556 MP	S - 05F1258AH	4 / 600XL	. 319340F	R - 00C1522		
Equipme	nt Calibrat	ion: See Dai	ly Equipm	ent Calibra	tion Sheet	49914				
					rial No.: 25083					
	D. 1 (0/1)				c) Water Leve					
					water) x "K" = _					als.)
	"K"= 0.163	(2" well) "K"	= 0.50 (3.5	" well) "K	" = .653 (4" well)	"K" = 1.	02 (5" well) "k" = 1.4	6 (6" well)	
			FIEL	D WATER	QUALITY PAR	RAMETER	s			
					Specific			Dissolved	Water	
Date	Time	Discharge	pН	Temp.	Conductance	11 3 2 1 5 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	04/40/79/300000	Oxygen	Level	Color
		(Liters)	+/- 0.1	(°C)	mS uS +/- 3%	(NTU's) +/- 10	(mV) +/- 10	(mg/L) +/- 10%	(BTOC)	
1/26/09	NA	Initial	MA	41.0	NA	,	NA	NA	NA	NA
126101	NA.		101	NA	NA	NA	1	1	,	1
-		0.5					_			
		1.0								
		1.5		3						
		2.0								
		2.5								
		3.0								
		3.5								
1		4.0	1	4	1	1	1	4	V	1
Total Dia	nharan:	0 Li	loro		Casing Volum	oc Pomo	rod: NA			
			_	E Callan D	rum(s) Poly			untom Ott	200	
PROPERTY OF STREET		d: <u>126/09</u>	Charles and the control of the contr					/OAs w/HCl		
Date/Tin	ie Sampie	120101	_ @ _11.	20	Allalysis.	VOCS (62	00b) - 3 V	OAS WITCH		
				en variable de la Trava		NOT SPACE THAT WE COL	orestes de la labora	ananaan manning an		
	None	_ ~ ~			S/MSD Equip	ment Rins	eate Fie	ld Blank L	ab Split	
Commer	its: NM	: was not	measu	red.						
ļ										
Recorde	d by: Step	hen Penman	Vaçki Le	e) Signat	ure: per	• :-			Page 1 of	1
- 60	80 Alhan	ibra Ave., #	UZ · Mar	tinez, CA	94958-6105	1925 3	72-8108	• Fax: 192	51 372-6	705



WATER	QUALIT	Y SAMPLE	LOG SHE	ET	MELL IDEN.	TIFICATI	ON: MW	-7 DATE	=: 1/26	/09
Project N	lame: Hop	yard Cleaner:	s Pleasant	on, CA Pr	roject Task: Qu	arterly Mo	nitoring f	Project/Task	No. WR	0574
					Lab: STL San					
					Well Type: (F					
Is Well S	Secured? Y	es / No Bolt	Size: 9/4	23/4"	Type of lock	Lock nur	nber: Ma	ster		
Observa	tions / Con	nments: set p	oump intak	e@ NA	ft.(BTOC)	Screen In	terval: 2	0' to 30'		
Purge M	ethod: NA	Teflon / PE	Disposable	e Bailer C	entrifugal Pum	p Perista	Itic Pump	Other: _		
					Bailer Line: (
					Γap Water DI F					
					Tap Water DIF					
					PE Bailer Per					
					S - 05F1258AF	1 / 600XL	. 319340F	R - 00C1522		
		ion: See Dai				(49914)			
					rial No.: 25083					
	NM av				c) Water Level					
TD =	-	-			water) x "K" =					als.)
	"K"= 0.163	(2" well) "K"			" = .653 (4" well)) "k" = 1,4	6 (6" well)	_
			FIEL	D WATER	QUALITY PAR	RAMETER	S			
NESSON E	missions	12400499877			Specific			Dissolved	Water	
Date	Time	Discharge (Litera)	pН	Temp. (°C)	Conductance mS uS	(NTU's)	Redox (mV)	Oxygen (mg/L)	Level (BTOC)	Color
		(Liters)	+/- 0.1	('0)	+/- 3%	+/- 10	+/- 10	+/- 10%	(6100)	
1/26/09	NA	Initial	NA	NA	NA	NA	NA	NA	NA	NA
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Total Dis	charge:	Ø Li	tore		Casing Volum	es Remo	red: NA			
	_			5 Gallon D	rum(s) Poly			vstem Oti	nor:	
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Comme	nts: NM:	Was not	measone	d						
					10 22					
Recorde	d by: Step	hen Penman	Jacki Le	e) Signal	ure:				Page 1 of	1



Table 1: January 2009 Quarterly Groundwater Monitoring Event

Project Name: Hopyard Cleaners

Project Location: 2771 Hopyard Road, Pleasanton, California

Well/Sample Identification	Date of Measurement	Time of Measurement	Depth to Groundwater (Ft., TOC)	Well Depth (Ft., TOC)	Sample Date	PDB Sample Time	Anion, Gene- Trac & Feasibilty Sample Time	QA/QC Type	QA/QC Sample Identification
MW-1	1/26/2009	8:31	16.71	30.27	1/26/2009	9:33	11:23	None	NA
MW-2	1/26/2009	8:33	16.17	30.31	1/26/2009	9:50	14:00	Duplicate	MW-DUP
MW-3	1/26/2009	8:27	17.10	30.29	1/26/2009	9:15	10:02	None	NA
MW-4	1/26/2009	8:43	17.86	34.56	1/26/2009	10:52	NA	None	NA
MW-5	1/26/2009	8:38	30.61	59.96	1/26/2009	10:15	NA	None	NA
MW-6	1/26/2009	8:55	28.10	NM	1/26/2009	11:20	NA	None	NA
MW-7	1/26/2009	9:00	28.19	NM	1/26/2009	11:45	NA NA	None	NA

Legend:

TOC = Top of Well Casing

NA = Not Applicable

NM = Not Measured



SITE NAME: Hopyard Cleaners SITE LOCATION: Pleasanton, CA

TASK: January 2009 Quarterly Groundwater Monitoring Event

DAILY EQUIPMENT CALIBRATION SHEET

		-		Dissolved		pH		Specific Conductance	ORP	TURBIDITY
DATE	TIME	Instrument Serial Number	Probe Serial Number	Oxygen (%)	4	7	10	1,000 uS/cm	mV	NTU's
26/08	9:23	556 MPS	05F1258AH	199	4.00	7.00	10.01	1,000	234.9	0.02

Report To: Angela Lie Company: Geosyntec	Consultants street, Suite 450 CA 94612 eosyntec.com	sher Tel Fax Pro	Martin S Tel: (9 Log C	nez, Ca 925) 37 ode: E (510) 2 836-30 me:	<u> 36</u>	a 945 3 Fa www	553- x: (9	925)	372-		LA				DT	IAIN 'IME	O I	F C	US				COR	æ	Ø	(Page <u>l</u> of 1 Other:
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APPENDIX B

Groundwater and SVE Monitoring Laboratory Analytical Reports



ANALYTICAL REPORT

Job Number: 720-17838-1

Job Description: Hopyard Cleaners

For:

Geosyntec Consultants, Inc.. 475 14th Street, Suite 450 Oakland, CA 94612

Attention: Ms. Melissa Asher

Approved for release Dimple Sharma Project Manager I 1/30/2009 4:57 PM

Designee for
Melissa Brewer
Project Manager I
melissa.brewer@testamericainc.com
01/30/2009

cc: Ms. Angela Liang

Job Narrative 720-J17838-1

Comments

No additional comments.

ReceiptAll samples were received in good condition within temperature requirements.

GC/MS VOA

No analytical or quality issues were noted.

EXECUTIVE SUMMARY - Detections

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Lab Sample ID Analyte	Client Sample ID	Result / Qualifier	Reporting Limit	Units	Method
720-17838-2	MW-3				
cis-1,2-Dichloroethe	ne	4.6	0.50	ug/L	8260B
Tetrachloroethene		42	0.50	ug/L	8260B
Trichloroethene		4.7	0.50	ug/L	8260B
720-17838-3	MW-1				
cis-1,2-Dichloroethe	ne	240	20	ug/L	8260B
Tetrachloroethene		1700	20	ug/L	8260B
Trichloroethene		320	20	ug/L	8260B
720-17838-4	MW-2				
cis-1,2-Dichloroethe	ne	760	100	ug/L	8260B
Tetrachloroethene		12000	100	ug/L	8260B
Trichloroethene		720	100	ug/L	8260B
720-17838-6	MW-5				
Tetrachloroethene		37	0.50	ug/L	8260B
720-17838-7	MW-DUP				
cis-1,2-Dichloroethe		770	100	ug/L	8260B
Tetrachloroethene	ile.	12000	100	ug/L ug/L	8260B
Trichloroethene		730	100	ug/L	8260B
720-17838-8	MW-4				
cis-1,2-Dichloroethe		4.3	0.50	ug/L	8260B
Trichloroethene	ii.	4.9	0.50	ug/L ug/L	8260B
720-17838-11	MW-7				
Tetrachloroethene		9.9	0.50	ug/L	8260B

METHOD SUMMARY

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Description	Lab Location	Method	Preparation Method
Matrix: Water			
Volatile Organic Compounds (GC/MS) Purge and Trap	TAL SF TAL SF	SW846 8260B	SW846 5030B
EPA 300.0		EPA 300.0	

Lab References:

= TestAmerica Morgan Hill

TAL SF = TestAmerica San Francisco

Method References:

EPA = US Environmental Protection Agency

SW846 = "Test Methods For Evaluating Solid Waste, Physical/Chemical Methods", Third Edition, November 1986 And Its Updates.

SAMPLE SUMMARY

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Lab Sample ID	Client Sample ID	Client Matrix	Date/Time Sampled	Date/Time Received
720-17838-1	TRIP BLANK	Water	01/26/2009 0730	01/26/2009 1545
720-17838-2	MW-3	Water	01/26/2009 0915	01/26/2009 1545
720-17838-3	MVV-1	Water	01/26/2009 0933	01/26/2009 1545
720-17838-4	MW-2	Water	01/26/2009 0950	01/26/2009 1545
720-17838-5	MW-3	Water	01/26/2009 1002	01/26/2009 1545
720-17838-6	MW-5	Water	01/26/2009 1015	01/26/2009 1545
720-17838-7	MW-DUP	Water	01/26/2009 1050	01/26/2009 1545
720-17838-8	MW-4	Water	01/26/2009 1052	01/26/2009 1545
720-17838-9	MW-6	Water	01/26/2009 1120	01/26/2009 1545
720-17838-10	MW-1	Water	01/26/2009 1123	01/26/2009 1545
720-17838-11	MW-7	Water	01/26/2009 1145	01/26/2009 1545
720-17838-12	MW-2	Water	01/26/2009 1400	01/26/2009 1545
720-17838-13	MW-DUP	Water	01/26/2009 1430	01/26/2009 1545

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: TRIP BLANK

 Lab Sample ID:
 720-17838-1
 Date Sampled:
 01/26/2009 0730

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1324 Final Weight/Volume: 40 mL

Acetone ND 50 Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromomethane ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 Carbon disulfide ND 1.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroform ND 1.0 Chloroform ND 1.0 Chloroform ND 1.0 Chloroformethane ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,2-Dichlorobenzene ND 0.50	Analyte	Result (ug/L)	Qualifier	RL
Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 Sender (MEK) ND 1.0 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon tetrachoride ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 5.0 Chlorodethane ND 0.50 Chlorodethane ND 1.0 Chlorodethane ND 1.0 Chlorodouluene ND 1.0 Chlorodouluene ND 0.50 Chlorodouluene ND 0.50 Chlorodouluene ND 0.50 Chlorodouluene ND 0.50 L'A-Chlorodouluene ND 0.50	Methyl tert-butyl ether			
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4-Isopropyltoluene ND 1.0				
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Mentale Annua :111	Methylene Chloride	ND		5.0

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: TRIP BLANK

 Lab Sample ID:
 720-17838-1
 Date Sampled:
 01/26/2009 0730

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1324 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	ND		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	ND		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	%Rec	Acceptance	Limits
4-Bromofluorobenzene	99	74 - 131	
1,2-Dichloroethane-d4 (Surr)	100	76 - 132	
Toluene-d8 (Surr)	98	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-3

 Lab Sample ID:
 720-17838-2
 Date Sampled:
 01/26/2009 0915

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1538 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
Methyl tert-butyl ether	ND		5.0
Acetone	ND		50
Benzene	ND		0.50
Dichlorobromomethane	ND		0.50
Bromobenzene	ND		1.0
Chlorobromomethane	ND		1.0
Bromoform	ND		1.0
Bromomethane	ND		1.0
2-Butanone (MEK)	ND		50
n-Butylbenzene	ND		1.0
sec-Butylbenzene	ND		1.0
tert-Butylbenzene	ND		1.0
Carbon disulfide	ND		5.0
Carbon tetrachloride	ND		0.50
Chlorobenzene	ND		0.50
Chloroethane	ND		1.0
Chloroform	ND		1.0
Chloromethane	ND		1.0
2-Chlorotoluene	ND		0.50
4-Chlorotoluene	ND		0.50
Chlorodibromomethane	ND		0.50
1,2-Dichlorobenzene	ND		0.50
1,3-Dichlorobenzene	ND		0.50
1,4-Dichlorobenzene	ND		0.50
1,3-Dichloropropane	ND		1.0
1,1-Dichloropropene	ND		0.50
1,2-Dibromo-3-Chloropropane	ND		1.0
Ethylene Dibromide	ND		0.50
Dibromomethane	ND		0.50
Dichlorodifluoromethane	ND		0.50
1,1-Dichloroethane	ND		0.50
1,2-Dichloroethane	ND		0.50
1,1-Dichloroethene	ND		0.50
cis-1,2-Dichloroethene	4.6		0.50
trans-1,2-Dichloroethene	ND		0.50
1,2-Dichloropropane	ND		0.50
cis-1,3-Dichloropropene	ND		0.50
trans-1,3-Dichloropropene	ND		0.50
Ethylbenzene	ND		0.50
Hexachlorobutadiene	ND		1.0
2-Hexanone	ND		50
Isopropylbenzene	ND		0.50
4-Isopropyltoluene	ND		1.0
Methylene Chloride	ND		5.0

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-3

 Lab Sample ID:
 720-17838-2
 Date Sampled:
 01/26/2009 0915

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1538 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	42		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	4.7		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	%Rec	Acceptance	Limits
4-Bromofluorobenzene	103	74 - 131	
1,2-Dichloroethane-d4 (Surr)	96	76 - 132	
Toluene-d8 (Surr)	97	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-1

 Lab Sample ID:
 720-17838-3
 Date Sampled:
 01/26/2009 0933

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 40 Initial Weight/Volume: 40 mL

Date Analyzed: 01/29/2009 1721 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
Methyl tert-butyl ether	ND		200
Acetone	ND		2000
Benzene	ND		20
Dichlorobromomethane	ND		20
Bromobenzene	ND		40
Chlorobromomethane	ND		40
Bromoform	ND		40
Bromomethane	ND		40
2-Butanone (MEK)	ND		2000
n-Butylbenzene	ND		40
sec-Butylbenzene	ND		40
tert-Butylbenzene	ND		40
Carbon disulfide	ND		200
Carbon tetrachloride	ND		20
Chlorobenzene	ND		20
Chloroethane	ND		40
Chloroform	ND		40
Chloromethane	ND		40
2-Chlorotoluene	ND		20
4-Chlorotoluene	ND		20
Chlorodibromomethane	ND		20
1,2-Dichlorobenzene	ND		20
1,3-Dichlorobenzene	ND		20
1,4-Dichlorobenzene	ND		20
1,3-Dichloropropane	ND		40
1,1-Dichloropropene	ND		20
1,2-Dibromo-3-Chloropropane	ND		40
Ethylene Dibromide	ND		20
Dibromomethane	ND		20
Dichlorodifluoromethane	ND		20
1,1-Dichloroethane	ND		20
1,2-Dichloroethane	ND		20
1,1-Dichloroethene	ND		20
cis-1,2-Dichloroethene	240		20
trans-1,2-Dichloroethene	ND		20
1,2-Dichloropropane	ND		20
cis-1,3-Dichloropropene	ND		20
trans-1,3-Dichloropropene	ND		20
Ethylbenzene	ND		20
Hexachlorobutadiene	ND		40
2-Hexanone	ND		2000
Isopropylbenzene	ND		20
4-Isopropyltoluene	ND		40
Methylene Chloride	ND		200

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-1

 Lab Sample ID:
 720-17838-3
 Date Sampled:
 01/26/2009 0933

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 40 Initial Weight/Volume: 40 mL
Date Analyzed: 01/29/2009 1721 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		2000
Naphthalene	ND		40
N-Propylbenzene	ND		40
Styrene	ND		20
1,1,1,2-Tetrachloroethane	ND		20
1,1,2,2-Tetrachloroethane	ND		20
Tetrachloroethene	1700		20
Toluene	ND		20
1,2,3-Trichlorobenzene	ND		40
1,2,4-Trichlorobenzene	ND		40
1,1,1-Trichloroethane	ND		20
1,1,2-Trichloroethane	ND		20
Trichloroethene	320		20
Trichlorofluoromethane	ND		40
1,2,3-Trichloropropane	ND		20
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		20
1,2,4-Trimethylbenzene	ND		20
1,3,5-Trimethylbenzene	ND		20
Vinyl acetate	ND		2000
Vinyl chloride	ND		20
Xylenes, Total	ND		40
2,2-Dichloropropane	ND		20
Surrogate	%Rec	Accept	ance Limits
4-Bromofluorobenzene	102	74 - 1	
1,2-Dichloroethane-d4 (Surr)	108	76 - 1	32
Toluene-d8 (Surr)	98	82 - 1	20

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-2

 Lab Sample ID:
 720-17838-4
 Date Sampled:
 01/26/2009 0950

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 200 Initial Weight/Volume: 40 mL

Date Analyzed: 01/29/2009 1754 Final Weight/Volume: 40 mL

Methyl tert-butyl ether Acetone	ND ND ND	1000
Acetone		40000
	ND	10000
Benzene	110	100
Dichlorobromomethane	ND	100
Bromobenzene	ND	200
Chlorobromomethane	ND	200
Bromoform	ND	200
Bromomethane	ND	200
2-Butanone (MEK)	ND	10000
n-Butylbenzene	ND	200
sec-Butylbenzene	ND	200
tert-Butylbenzene	ND	200
Carbon disulfide	ND	1000
Carbon tetrachloride	ND	100
Chlorobenzene	ND	100
Chloroethane	ND	200
Chloroform	ND	200
Chloromethane	ND	200
2-Chlorotoluene	ND	100
4-Chlorotoluene	ND	100
Chlorodibromomethane	ND	100
1,2-Dichlorobenzene	ND	100
1,3-Dichlorobenzene	ND	100
1,4-Dichlorobenzene	ND	100
1,3-Dichloropropane	ND	200
1,1-Dichloropropene	ND	100
1,2-Dibromo-3-Chloropropane	ND	200
Ethylene Dibromide	ND	100
Dibromomethane	ND	100
Dichlorodifluoromethane	ND	100
1.1-Dichloroethane	ND	100
1,2-Dichloroethane	ND	100
1,1-Dichloroethene	ND	100
cis-1,2-Dichloroethene	760	100
trans-1,2-Dichloroethene	ND	100
1,2-Dichloropropane	ND	100
cis-1,3-Dichloropropene	ND	100
trans-1,3-Dichloropropene	ND	100
Ethylbenzene	ND	100
Hexachlorobutadiene	ND	200
2-Hexanone	ND	10000
Isopropylbenzene	ND	100
4-Isopropyltoluene	ND	200
Methylene Chloride	ND	1000

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-2

 Lab Sample ID:
 720-17838-4
 Date Sampled:
 01/26/2009 0950

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 200 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1754 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		10000
Naphthalene	ND		200
N-Propylbenzene	ND		200
Styrene	ND		100
1,1,1,2-Tetrachloroethane	ND		100
1,1,2,2-Tetrachloroethane	ND		100
Tetrachloroethene	12000		100
Toluene	ND		100
1,2,3-Trichlorobenzene	ND		200
1,2,4-Trichlorobenzene	ND		200
1,1,1-Trichloroethane	ND		100
1,1,2-Trichloroethane	ND		100
Trichloroethene	720		100
Trichlorofluoromethane	ND		200
1,2,3-Trichloropropane	ND		100
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		100
1,2,4-Trimethylbenzene	ND		100
1,3,5-Trimethylbenzene	ND		100
Vinyl acetate	ND		10000
Vinyl chloride	ND		100
Xylenes, Total	ND		200
2,2-Dichloropropane	ND		100
Surrogate	%Rec	Acceptance	Limits
4-Bromofluorobenzene	95	74 - 131	
1,2-Dichloroethane-d4 (Surr)	109	76 - 132	
Toluene-d8 (Surr)	95	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-5

 Lab Sample ID:
 720-17838-6
 Date Sampled:
 01/26/2009 1015

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1612 Final Weight/Volume: 40 mL

Methyl tert-butyl ether ND 5.0 Acetone ND 50 Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chloroethane ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chlorotoluene ND 0.50
Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 5.0 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chloromethane ND 1.0
Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 1.0 Carbon tetrachloride ND 5.0 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chloromethane ND 1.0
Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chloromethane ND 1.0
Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chloromethane ND 1.0
Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chloromethane ND 1.0
Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0 Chloromethane ND 1.0
n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
Chlorobenzene ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chloromethane ND 1.0
ChloroethaneND1.0ChloroformND1.0ChloromethaneND1.0
ChloroformND1.0ChloromethaneND1.0
Chloromethane ND 1.0
2-Chlorotoluene ND 0.50
4-Chlorotoluene ND 0.50
Chlorodibromomethane ND 0.50
1,2-Dichlorobenzene ND 0.50
1,3-Dichlorobenzene ND 0.50
1,4-Dichlorobenzene ND 0.50
1,3-Dichloropropane ND 1.0
1,1-Dichloropropene ND 0.50
1,2-Dibromo-3-Chloropropane ND 1.0
Ethylene Dibromide ND 0.50
Dibromomethane ND 0.50
Dichlorodifluoromethane ND 0.50
1,1-Dichloroethane ND 0.50
1,2-Dichloroethane ND 0.50
1,1-Dichloroethene ND 0.50
cis-1,2-Dichloroethene ND 0.50
trans-1,2-Dichloroethene ND 0.50
1,2-Dichloropropane ND 0.50
cis-1,3-Dichloropropene ND 0.50
trans-1,3-Dichloropropene ND 0.50
Ethylbenzene ND 0.50
Hexachlorobutadiene ND 1.0
2-Hexanone ND 50
Isopropylbenzene ND 0.50
4-Isopropyltoluene ND 1.0
Methylene Chloride ND 5.0

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-5

 Lab Sample ID:
 720-17838-6
 Date Sampled:
 01/26/2009 1015

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1612 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	37		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	ND		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	%Rec	Acceptance	e Limits
4-Bromofluorobenzene	100	74 - 131	
1,2-Dichloroethane-d4 (Surr)	103	76 - 132	
Toluene-d8 (Surr)	95	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-DUP

 Lab Sample ID:
 720-17838-7
 Date Sampled:
 01/26/2009 1050

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 200 Initial Weight/Volume: 40 mL

Date Analyzed: 01/29/2009 1828 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
Methyl tert-butyl ether	ND		1000
Acetone	ND		10000
Benzene	ND		100
Dichlorobromomethane	ND		100
Bromobenzene	ND		200
Chlorobromomethane	ND		200
Bromoform	ND		200
Bromomethane	ND		200
2-Butanone (MEK)	ND		10000
n-Butylbenzene	ND		200
sec-Butylbenzene	ND		200
tert-Butylbenzene	ND		200
Carbon disulfide	ND		1000
Carbon tetrachloride	ND		100
Chlorobenzene	ND		100
Chloroethane	ND		200
Chloroform	ND		200
Chloromethane	ND		200
2-Chlorotoluene	ND		100
4-Chlorotoluene	ND		100
Chlorodibromomethane	ND		100
1,2-Dichlorobenzene	ND		100
1,3-Dichlorobenzene	ND		100
1,4-Dichlorobenzene	ND		100
1,3-Dichloropropane	ND		200
1,1-Dichloropropene	ND		100
1,2-Dibromo-3-Chloropropane	ND		200
Ethylene Dibromide	ND		100
Dibromomethane	ND		100
Dichlorodifluoromethane	ND		100
1,1-Dichloroethane	ND		100
1,2-Dichloroethane	ND		100
1,1-Dichloroethene	ND		100
cis-1,2-Dichloroethene	770		100
trans-1,2-Dichloroethene	ND		100
1,2-Dichloropropane	ND		100
cis-1,3-Dichloropropene	ND		100
trans-1,3-Dichloropropene	ND		100
Ethylbenzene	ND		100
Hexachlorobutadiene	ND		200
2-Hexanone	ND		10000
Isopropylbenzene	ND		100
4-Isopropyltoluene	ND		200
Methylene Chloride	ND		1000

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-DUP

 Lab Sample ID:
 720-17838-7
 Date Sampled:
 01/26/2009 1050

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 200 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1828 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		10000
Naphthalene	ND		200
N-Propylbenzene	ND		200
Styrene	ND		100
1,1,1,2-Tetrachloroethane	ND		100
1,1,2,2-Tetrachloroethane	ND		100
Tetrachloroethene	12000		100
Toluene	ND		100
1,2,3-Trichlorobenzene	ND		200
1,2,4-Trichlorobenzene	ND		200
1,1,1-Trichloroethane	ND		100
1,1,2-Trichloroethane	ND		100
Trichloroethene	730		100
Trichlorofluoromethane	ND		200
1,2,3-Trichloropropane	ND		100
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		100
1,2,4-Trimethylbenzene	ND		100
1,3,5-Trimethylbenzene	ND		100
Vinyl acetate	ND		10000
Vinyl chloride	ND		100
Xylenes, Total	ND		200
2,2-Dichloropropane	ND		100
Surrogate	%Rec	Acceptance	Limits
4-Bromofluorobenzene	104	74 - 131	
1,2-Dichloroethane-d4 (Surr)	101	76 - 132	
Toluene-d8 (Surr)	96	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-4

 Lab Sample ID:
 720-17838-8
 Date Sampled:
 01/26/2009 1052

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1645 Final Weight/Volume: 40 mL

Methyl tert-butyl ether ND 5.0 Acetone ND 50 Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 Sendanone (MEK) ND 1.0 Seuthylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 1.0 Carbon disulfide ND 0.50 Chloroethane ND 0.50 Chloroethane ND 1.0 Chloroethane ND 1.0 Chlorotoform ND 1.0 Chlorotofuene ND 0.50 Chlorotoluene ND 0.50 Chlorotoluene ND 0.50 1,2-Dichlorobenzene ND 0.50
Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 1.0 Carbon disulfide ND 0.50 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorosthane ND 0.50 Chlorosthane ND 1.0 Chlorostoluene ND 0.50 Chlorostoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 1,2-Dichlorobenzene ND 0.50 </td
Dichlorobromomethane ND 1.0 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorobenzene ND 1.0 Chlorotothane ND 1.0 Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,3-Dichloropropane ND 0.50 1,2-Dichloropropane ND <
Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorobenzene ND 0.50 Chlorothane ND 1.0 Chloroform ND 1.0 Chlorotoluene ND 1.0 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 Chlorodibromomethane ND 0.50 1,2-Dichlorobenzene ND 0.50 1,4-Dichloropropane ND 0.50 1,3-Dichloropropane ND 0.50 1,2-Dibromo-3-Chloropropane ND
Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorothane ND 1.0 Chlorothane ND 1.0 Chlorothane ND 1.0 Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 Chlorodibromomethane ND 0.50 1,2-Dichlorobenzene ND 0.50 1,4-Dichlorobenzene ND 0.50 1,4-Dichloropropane ND 0.50 1,2-Dibromo-3-Chloropropane ND 0.50 1,2-Dibromo-3-Chloropropane
Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorotethane ND 1.0 Chlorotemane ND 1.0 Chlorotethane ND 1.0 Chlorotethane ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotobenzene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,4-Dichloropenzene ND 0.50 1,4-Dichloropropane ND 0.50 1,1-Dichloropropene ND 0.50 1,2-Dibromo-3-Chloropropane ND
Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorotethane ND 1.0 Chloroform ND 1.0 Chlorothane ND 1.0 2-Chlorotoluene ND 1.0 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotobenzene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,3-Dichloropropane ND 0.50 1,4-Dichloropropane ND 0.50 1,2-Dibromo-3-Chloropropane ND 0.50 1,2-Dibromo-3-Chloropropane ND 0.50 Dibromomethane <
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1,4-Dichlorobenzene ND 0.50 1,3-Dichloropropane ND 1.0 1,1-Dichloropropene ND 0.50 1,2-Dibromo-3-Chloropropane ND 1.0 Ethylene Dibromide ND 0.50 Dibromomethane ND 0.50 Dichlorodifluoromethane ND 0.50 1,1-Dichloroethane ND 0.50 1,2-Dichloroethane ND 0.50
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1,2-Dichloroethane ND 0.50
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1.1-DIGHIGHO UCHICHE U
cis-1,2-Dichloroethene 4.3 0.50
trans-1,2-Dichloroethene ND 0.50
1,2-Dichloropropane ND 0.50
cis-1,3-Dichloropropene ND 0.50
trans-1,3-Dichloropropene ND 0.50
Ethylbenzene ND 0.50
Hexachlorobutadiene ND 1.0
2-Hexanone ND 50
Isopropylbenzene ND 0.50
4-Isopropyltoluene ND 1.0
Methylene Chloride ND 5.0

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-4

 Lab Sample ID:
 720-17838-8
 Date Sampled:
 01/26/2009 1052

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Preparation: 5030B Lab File ID: e:\data\200901\012909\SA-

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 1645 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	ND		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	4.9		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	%Rec	Acceptance	e Limits
4-Bromofluorobenzene	101	74 - 131	
1,2-Dichloroethane-d4 (Surr)	103	76 - 132	
Toluene-d8 (Surr)	102	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-6

 Lab Sample ID:
 720-17838-9
 Date Sampled:
 01/26/2009 1120

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Preparation: 5030B Lab File ID: e:\200901\013009\SA-WA

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/30/2009 1426 Final Weight/Volume: 40 mL

Acetone ND 50 Benzene ND 0.50 Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromomethane ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroform ND 1.0 Chloroform ND 1.0 Chloroform ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 1,2-Dichlorobenzene ND 0.50 <tr< th=""><th>Analyte</th><th>Result (ug/L)</th><th>Qualifier</th><th>RL</th></tr<>	Analyte	Result (ug/L)	Qualifier	RL
Benzene	Methyl tert-butyl ether			
Dichlorobromomethane ND 0.50 Bromobenzene ND 1.0 Chlorobromomethane ND 1.0 Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 1.0 Carbon tetrachloride ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chloroform ND 1.0 Chloroform ND 1.0 Chloroform ND 1.0 Chloroform ND 1.0 Chlorodiuene ND 0.50 Chlorodibromomethane ND 0.50 Chlorodibromomethane ND 0.50 Chlorodibromomethane ND 0.50 1,4-Dichloropropene ND 0.50 1,2-Dichloropropene ND 0.50	Acetone			
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Chlorobromomethane	Dichlorobromomethane			
Bromoform ND 1.0 Bromomethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tert-Butylbenzene ND 1.0 Carbon disulfide ND 0.50 Carbon tetrachloride ND 0.50 Chlorotenzene ND 0.50 Chlorotethane ND 0.50 Chlorotethane ND 1.0 Chlorotoform ND 1.0 Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotoluene ND 0.50 4-Chlorotobenzene ND 0.50 4-Chlorotobenzene ND 0.50 1,3-Dichlorobenzene ND 0.50 1,3-Dichloropropane ND 0.50 1,3-Dichloropropane ND 0.50 1,1-Dichloropropane ND 0.50 Dibromo-Achloropropane ND	Bromobenzene	ND		1.0
Brommethane ND 1.0 2-Butanone (MEK) ND 50 n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 carbon disulfide ND 1.0 Carbon disulfide ND 0.50 Chloroel tetrachloride ND 0.50 Chloroethane ND 0.50 Chloroethane ND 1.0 Chloroform ND 1.0 Chlororofure ND 1.0 Chlorotoluene ND 1.0 4-Chlorotoluene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,2-Dichlorobenzene ND 0.50 1,3-Dichloropropane ND 0.50 1,1-Dichloropropane ND	Chlorobromomethane	ND		1.0
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n-Butylbenzene ND 1.0 sec-Butylbenzene ND 1.0 tetr-Butylbenzene ND 1.0 tetr-Butylbenzene ND 1.0 Carbon disulfide ND 5.0 Carbon tetrachloride ND 0.50 Chlorobenzene ND 0.50 Chlorobenzene ND 0.50 Chlorobenzene ND 1.0 Chloroform ND 1.0 Chlorodilune ND 0.50 Chlorodilune ND 0.50 Chlorodibromomethane ND 0.50	Bromomethane	ND		1.0
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IsopropylbenzeneND0.504-IsopropyltolueneND1.0				
4-Isopropyltoluene ND 1.0				
• • •				
Methylene Chloride ND 5.0	Methylene Chloride			5.0

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-6

 Lab Sample ID:
 720-17838-9
 Date Sampled:
 01/26/2009 1120

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Preparation: 5030B Lab File ID: e:\200901\013009\SA-WA

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/30/2009 1426 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	ND		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	ND		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	%Rec	Acceptance	Limits
4-Bromofluorobenzene	104	74 - 131	
1,2-Dichloroethane-d4 (Surr)	103	76 - 132	
Toluene-d8 (Surr)	104	82 - 120	

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-7

 Lab Sample ID:
 720-17838-11
 Date Sampled:
 01/26/2009
 1145

 Client Matrix:
 Water
 Date Received:
 01/26/2009
 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Preparation: 5030B Lab File ID: e:\200901\013009\SA-WA

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/30/2009 1459 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
Methyl tert-butyl ether	ND		5.0
Acetone	ND		50
Benzene	ND		0.50
Dichlorobromomethane	ND		0.50
Bromobenzene	ND		1.0
Chlorobromomethane	ND		1.0
Bromoform	ND		1.0
Bromomethane	ND		1.0
2-Butanone (MEK)	ND		50
n-Butylbenzene	ND		1.0
sec-Butylbenzene	ND		1.0
tert-Butylbenzene	ND		1.0
Carbon disulfide	ND		5.0
Carbon tetrachloride	ND		0.50
Chlorobenzene	ND		0.50
Chloroethane	ND		1.0
Chloroform	ND		1.0
Chloromethane	ND		1.0
2-Chlorotoluene	ND		0.50
4-Chlorotoluene	ND		0.50
Chlorodibromomethane	ND		0.50
1,2-Dichlorobenzene	ND		0.50
	ND		
	ND		0.50
	ND		1.0
	ND		
	ND		
	ND		
•			
1.1-Dichloroethane	ND		
1,2-Dichloroethane	ND		0.50
•			0.50
•	ND		0.50
	ND		
	ND		
=			
1,3-Dichlorobenzene 1,4-Dichlorobenzene 1,3-Dichloropropane 1,1-Dichloropropene 1,2-Dibromo-3-Chloropropane Ethylene Dibromide Dibromomethane Dichlorodifluoromethane	ND N		0.50 0.50 1.0 0.50 1.0 0.50 0.50 0.50 0.50 0.50

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Client Sample ID: MW-7

 Lab Sample ID:
 720-17838-11
 Date Sampled:
 01/26/2009 1145

 Client Matrix:
 Water
 Date Received:
 01/26/2009 1545

8260B Volatile Organic Compounds (GC/MS)

Method: 8260B Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Preparation: 5030B Lab File ID: e:\200901\013009\SA-WA

Dilution: 1.0 Initial Weight/Volume: 40 mL Date Analyzed: 01/30/2009 1459 Final Weight/Volume: 40 mL

Analyte	Result (ug/L)	Qualifier	RL
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	9.9		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	ND		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	%Rec		Acceptance Limits
4-Bromofluorobenzene	102		74 - 131
1,2-Dichloroethane-d4 (Surr)	97		76 - 132
Toluene-d8 (Surr)	99		82 - 120

DATA REPORTING QUALIFIERS

Lab Section Qualifier Description

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

QC Association Summary

		Report			
Lab Sample ID	Client Sample ID	Basis	Client Matrix	Method	Prep Batch
GC/MS VOA					
Analysis Batch:720-46	316				
LCS 720-46316/2	Lab Control Spike	Т	Water	8260B	
LCSD 720-46316/1	Lab Control Spike Duplicate	Т	Water	8260B	
MB 720-46316/3	Method Blank	Т	Water	8260B	
720-17838-1	TRIP BLANK	Т	Water	8260B	
720-17838-2	MW-3	Т	Water	8260B	
720-17838-3	MW-1	Т	Water	8260B	
720-17838-4	MW-2	Т	Water	8260B	
720-17838-6	MW-5	Т	Water	8260B	
720-17838-7	MW-DUP	Т	Water	8260B	
720-17838-8	MW-4	Т	Water	8260B	
Analysis Batch:720-46	397				
LCS 720-46397/2	Lab Control Spike	Т	Water	8260B	
LCSD 720-46397/1	Lab Control Spike Duplicate	Т	Water	8260B	
MB 720-46397/3	Method Blank	Т	Water	8260B	
720-17838-9	MW-6	Т	Water	8260B	
720-17838-11	MW-7	Т	Water	8260B	

Report Basis T = Total

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Method Blank - Batch: 720-46316 Method: 8260B Preparation: 5030B

Lab Sample ID: MB 720-46316/3 Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\data\200901\012909\MB

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 0855 Final Weight/Volume: 40 mL

Date Prepared: 01/29/2009 0855

Analyte	Result	Qual	RL
Methyl tert-butyl ether	ND		5.0
Acetone	ND		50
Benzene	ND		0.50
Dichlorobromomethane	ND		0.50
Bromobenzene	ND		1.0
Chlorobromomethane	ND		1.0
Bromoform	ND		1.0
Bromomethane	ND		1.0
2-Butanone (MEK)	ND		50
n-Butylbenzene	ND		1.0
sec-Butylbenzene	ND		1.0
tert-Butylbenzene	ND		1.0
Carbon disulfide	ND		5.0
Carbon tetrachloride	ND		0.50
Chlorobenzene	ND		0.50
Chloroethane	ND		1.0
Chloroform	ND		1.0
Chloromethane	ND		1.0
2-Chlorotoluene	ND		0.50
4-Chlorotoluene	ND		0.50
Chlorodibromomethane	ND		0.50
1,2-Dichlorobenzene	ND		0.50
1,3-Dichlorobenzene	ND		0.50
1,4-Dichlorobenzene	ND		0.50
1,3-Dichloropropane	ND		1.0
1,1-Dichloropropene	ND		0.50
1,2-Dibromo-3-Chloropropane	ND		1.0
Ethylene Dibromide	ND		0.50
Dibromomethane	ND		0.50
Dichlorodifluoromethane	ND		0.50
1,1-Dichloroethane	ND		0.50
1,2-Dichloroethane	ND		0.50
1,1-Dichloroethene	ND		0.50
cis-1,2-Dichloroethene	ND		0.50
trans-1,2-Dichloroethene	ND		0.50
1,2-Dichloropropane	ND		0.50
cis-1,3-Dichloropropene	ND		0.50
trans-1,3-Dichloropropene	ND		0.50
Ethylbenzene	ND		0.50
Hexachlorobutadiene	ND		1.0
2-Hexanone	ND		50

Calculations are performed before rounding to avoid round-off errors in calculated results.

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Method Blank - Batch: 720-46316 Method: 8260B Preparation: 5030B

Lab Sample ID: MB 720-46316/3 Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\data\200901\012909\MB

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL

Date Analyzed: 01/29/2009 0855 Final Weight/Volume: 40 mL Date Prepared: 01/29/2009 0855

Analyte	Result	Qual	RL
Isopropylbenzene	ND		0.50
4-Isopropyltoluene	ND		1.0
Methylene Chloride	ND		5.0
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	ND		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	ND		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	% Rec	Acceptance Limits	
4-Bromofluorobenzene	99	74 - 131	
1,2-Dichloroethane-d4 (Surr)	103	76 - 132	
Toluene-d8 (Surr)	98	82 - 120	

Calculations are performed before rounding to avoid round-off errors in calculated results.

82 - 120

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Lab Control Spike/ Method: 8260B
Lab Control Spike Duplicate Recovery Report - Batch: 720-46316 Preparation: 5030B

LCS Lab Sample ID: LCS 720-46316/2 Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\data\200901\012909\LS-

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL

Date Analyzed: 01/29/2009 0748 Final Weight/Volume: 40 mL Date Prepared: 01/29/2009 0748

LCSD Lab Sample ID: LCSD 720-46316/1 Analysis Batch: 720-46316 Instrument ID: Varian 3900G

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\data\200901\012909\LD-V

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL Date Analyzed: 01/29/2009 0822 Final Weight/Volume: 40 mL

<u>% R</u>ec. LCS **LCSD RPD** RPD Limit LCS Qual LCSD Qual Limit Analyte Benzene 87 81 70 - 130 6 20 104 109 70 - 130 20 Chlorobenzene 4 1.1-Dichloroethene 107 102 70 - 130 5 20 Toluene 70 - 130 20 98 93 5 70 - 130 Trichloroethene 82 87 6 20 LCS % Rec LCSD % Rec Surrogate Acceptance Limits 4-Bromofluorobenzene 94 104 74 - 131 1,2-Dichloroethane-d4 (Surr) 105 109 76 - 132

93

97

Calculations are performed before rounding to avoid round-off errors in calculated results.

Date Prepared:

Toluene-d8 (Surr)

01/29/2009 0822

Job Number: 720-17838-1 Client: Geosyntec Consultants, Inc..

Method Blank - Batch: 720-46397 Method: 8260B Preparation: 5030B

Lab Sample ID: MB 720-46397/3 Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\200901\013009\MB-WA

Units: ug/L Initial Weight/Volume: 40 mL Dilution: 1.0 Date Analyzed: 01/30/2009 1319 Final Weight/Volume: 40 mL

Date Prepared: 01/30/2009 1319

Analyte	Result	Qual	RL
Methyl tert-butyl ether	ND		5.0
Acetone	ND		50
Benzene	ND		0.50
Dichlorobromomethane	ND		0.50
Bromobenzene	ND		1.0
Chlorobromomethane	ND		1.0
Bromoform	ND		1.0
Bromomethane	ND		1.0
2-Butanone (MEK)	ND		50
n-Butylbenzene	ND		1.0
sec-Butylbenzene	ND		1.0
tert-Butylbenzene	ND		1.0
Carbon disulfide	ND		5.0
Carbon tetrachloride	ND		0.50
Chlorobenzene	ND		0.50
Chloroethane	ND		1.0
Chloroform	ND		1.0
Chloromethane	ND		1.0
2-Chlorotoluene	ND		0.50
4-Chlorotoluene	ND		0.50
Chlorodibromomethane	ND		0.50
1,2-Dichlorobenzene	ND		0.50
1,3-Dichlorobenzene	ND		0.50
1,4-Dichlorobenzene	ND		0.50
1,3-Dichloropropane	ND		1.0
1,1-Dichloropropene	ND		0.50
1,2-Dibromo-3-Chloropropane	ND		1.0
Ethylene Dibromide	ND		0.50
Dibromomethane	ND		0.50
Dichlorodifluoromethane	ND		0.50
1,1-Dichloroethane	ND		0.50
1,2-Dichloroethane	ND		0.50
1,1-Dichloroethene	ND		0.50
cis-1,2-Dichloroethene	ND		0.50
trans-1,2-Dichloroethene	ND		0.50
1,2-Dichloropropane	ND		0.50
cis-1,3-Dichloropropene	ND		0.50
trans-1,3-Dichloropropene	ND		0.50
Ethylbenzene	ND		0.50
Hexachlorobutadiene	ND		1.0
2-Hexanone	ND		50

Calculations are performed before rounding to avoid round-off errors in calculated results.

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Method Blank - Batch: 720-46397 Method: 8260B Preparation: 5030B

Lab Sample ID: MB 720-46397/3 Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\200901\013009\MB-WA

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL

Date Analyzed: 01/30/2009 1319 Final Weight/Volume: 40 mL Date Prepared: 01/30/2009 1319

Analyte	Result	Qual	RL
Isopropylbenzene	ND		0.50
4-Isopropyltoluene	ND		1.0
Methylene Chloride	ND		5.0
4-Methyl-2-pentanone (MIBK)	ND		50
Naphthalene	ND		1.0
N-Propylbenzene	ND		1.0
Styrene	ND		0.50
1,1,1,2-Tetrachloroethane	ND		0.50
1,1,2,2-Tetrachloroethane	ND		0.50
Tetrachloroethene	ND		0.50
Toluene	ND		0.50
1,2,3-Trichlorobenzene	ND		1.0
1,2,4-Trichlorobenzene	ND		1.0
1,1,1-Trichloroethane	ND		0.50
1,1,2-Trichloroethane	ND		0.50
Trichloroethene	ND		0.50
Trichlorofluoromethane	ND		1.0
1,2,3-Trichloropropane	ND		0.50
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		0.50
1,2,4-Trimethylbenzene	ND		0.50
1,3,5-Trimethylbenzene	ND		0.50
Vinyl acetate	ND		50
Vinyl chloride	ND		0.50
Xylenes, Total	ND		1.0
2,2-Dichloropropane	ND		0.50
Surrogate	% Rec	Acceptance Limits	
4-Bromofluorobenzene	104	74 - 131	
1,2-Dichloroethane-d4 (Surr)	99	76 - 132	
Toluene-d8 (Surr)	107	82 - 120	

Calculations are performed before rounding to avoid round-off errors in calculated results.

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Lab Control Spike/ Method: 8260B
Lab Control Spike Duplicate Recovery Report - Batch: 720-46397 Preparation: 5030B

LCS Lab Sample ID: LCS 720-46397/2 Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\200901\013009\LS-WA

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL Date Analyzed: 01/30/2009 1213 Final Weight/Volume: 40 mL

Date Analyzed: 01/30/2009 1213 Final Weight/Volume: 40 mL

Date Prepared: 01/30/2009 1213

LCSD Lab Sample ID: LCSD 720-46397/1 Analysis Batch: 720-46397 Instrument ID: Varian 3900F

Client Matrix: Water Prep Batch: N/A Lab File ID: e:\200901\013009\LD-WA

Dilution: 1.0 Units: ug/L Initial Weight/Volume: 40 mL Date Analyzed: 01/30/2009 1246 Final Weight/Volume: 40 mL

% Rec. LCS **RPD** LCSD Limit RPD Limit LCS Qual LCSD Qual Analyte Benzene 113 111 70 - 130 2 20 70 - 130 20 Chlorobenzene 110 116 5 1.1-Dichloroethene 105 103 70 - 130 2 20 Toluene 70 - 130 20 92 101 9 70 - 130 2 Trichloroethene 97 95 20 LCS % Rec LCSD % Rec Surrogate Acceptance Limits 4-Bromofluorobenzene 92 109 74 - 131 1,2-Dichloroethane-d4 (Surr) 95 104 76 - 132 95 104 82 - 120 Toluene-d8 (Surr)

Calculations are performed before rounding to avoid round-off errors in calculated results.

Date Prepared:

01/30/2009 1246

720-17838

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Login Sample Receipt Check List

Client: Geosyntec Consultants, Inc.. Job Number: 720-17838-1

Login Number: 17838 List Source: TestAmerica San Francisco

Creator: Bullock, Tracy List Number: 1

Question T / F/ NA Comment Radioactivity either was not measured or, if measured, is at or below N/A background The cooler's custody seal, if present, is intact. N/A The cooler or samples do not appear to have been compromised or True tampered with. Samples were received on ice. True True Cooler Temperature is acceptable. Cooler Temperature is recorded. True True COC is present. COC is filled out in ink and legible. True COC is filled out with all pertinent information. True There are no discrepancies between the sample IDs on the containers and True the COC. True Samples are received within Holding Time. True Sample containers have legible labels. Containers are not broken or leaking. True Sample collection date/times are provided. True Appropriate sample containers are used. True Sample bottles are completely filled. True There is sufficient vol. for all requested analyses, incl. any requested True MS/MSDs VOA sample vials do not have headspace or bubble is <6mm (1/4") in True diameter. If necessary, staff have been informed of any short hold time or quick TAT True needs Multiphasic samples are not present. True

True

Samples do not require splitting or compositing.



Friday, January 30, 2009 2:54:14PM

Melissa Brewer TestAmerica San Francisco 1220 Quarry Lane Pleasanton, CA 94566-4756

RE: Work Sharing Agreement Work Order: MSA0515

Enclosed are the results of analyses for samples received by the laboratory on 01/26/09 18:40. If you have any questions concerning this report, please feel free to contact me.

Sincerely,

Angel Pitts Project Manager

CA ELAP Certificate # 2705

The Chain(s) of Custody, 2 pages, are included and are an integral part of this report.

The report shall not be reproduced except in full, without the written approval of the laboratory. The client, by accepting this report, also agrees not to alter any reports whether in the hard copy or electronic format and to use reasonable efforts to preserve the reports in the form and substance originally provided by TestAmerica.

For Volatile Analysis a trip blank is required to be provided. If trip blank results are not included in the report, then either the trip blank was not submitted or requested to be analyzed.

The reported results were obtained in compliance with the 2003 NELAC standards unless otherwise noted.





TestAmerica San FranciscoProject:Work Sharing AgreementMSA05151220 Quarry LaneProject Number:720-17838Reported:Pleasanton CA, 94566-4756Project Manager:Melissa Brewer01/30/09 14:53

ANALYTICAL REPORT FOR SAMPLES

Sample ID	Laboratory ID	Matrix	Date Sampled	Date Received
MW-3	MSA0515-01	Water	01/26/09 10:02	01/26/09 18:40
MW-1	MSA0515-02	Water	01/26/09 11:23	01/26/09 18:40
MW-2	MSA0515-03	Water	01/26/09 14:00	01/26/09 18:40
MW-DUP	MSA0515-04	Water	01/26/09 14:30	01/26/09 18:40



TestAmerica San FranciscoProjectWork Sharing AgreementMSA05151220 Quarry LaneProject Number:720-17838Reported:Pleasanton CA, 94566-4756Project Manager:Melissa Brewer01/30/09 14:53

Anions by EPA Method 300.0 TestAmerica Morgan Hill

				8					
Analyte	Result	Reporting Limit	Units	Dilution	Batch	Prepared	Analyzed	Method	Notes
MW-3 (MSA0515-01) Water	Sampled: 01/26/09 10:02 R	eceived: 01/26/09	18:40						
Chloride	120	10	mg/l	10	9A27013	01/27/09	01/27/09	EPA 300.0	
Nitrate as NO3	13	0.50	"	1	"	"	01/27/09 08:52	"	
Nitrite as NO2	ND	0.50	"	"	"	"	"	"	
Sulfate as SO4	140	5.0	"	10	"	"	01/27/09	"	
MW-1 (MSA0515-02) Water	Sampled: 01/26/09 11:23 R	eceived: 01/26/09	18:40						
Chloride	83	10	mg/l	10	9A27013	01/27/09	01/27/09	EPA 300.0	
Nitrate as NO3	ND	0.50	"	1	"	"	01/27/09 09:56	"	
Nitrite as NO2	ND	0.50	"	"	"	"	"	"	
Sulfate as SO4	130	5.0	"	10	"	"	01/27/09	"	
MW-2 (MSA0515-03) Water	Sampled: 01/26/09 14:00 R	eceived: 01/26/09	18:40						
Chloride	110	10	mg/l	10	9A27013	01/27/09	01/27/09	EPA 300.0	
Nitrate as NO3	3.9	0.50	"	1	"	"	01/27/09 10:17	"	
Nitrite as NO2	ND	0.50	"	"	"	"	"	"	
Sulfate as SO4	140	5.0	"	10	"	"	01/27/09	"	
MW-DUP (MSA0515-04) Wa	ter Sampled: 01/26/09 14:30	Received: 01/20	6/09 18:40						
Chloride	110	10	mg/l	10	9A27013	01/27/09	01/27/09	EPA 300.0	
Nitrate as NO3	3.9	0.50	"	1	"	"	01/27/09 10:59	"	
Nitrite as NO2	ND	0.50	"	"	"	"	"	"	
Sulfate as SO4	140	5.0	"	10	"	"	01/27/09	"	



TestAmerica San FranciscoProject:Work Sharing AgreementMSA05151220 Quarry LaneProject Number:720-17838Reported:Pleasanton CA, 94566-4756Project Manager:Melissa Brewer01/30/09 14:53

Anions by EPA Method 300.0 - Quality Control TestAmerica Morgan Hill

		Reporting		Spike	Source		%REC		RPD		ĺ
Analyte	Result	Limit	Units	Level	Result	%REC	Limits	RPD	Limit	Notes	ĺ

Batel 7/12/013 General Treparation / E.	1112000									
Blank (9A27013-BLK1)				Prepared &	Analyzed:	01/27/09				
Chloride	ND	1.0	mg/l							
Nitrate as NO3	ND	0.50	"							
Nitrite as NO2	ND	0.50	"							
Sulfate as SO4	ND	0.50	"							
Laboratory Control Sample (9A27013-BS1)				Prepared &	Analyzed:	01/27/09				
Sulfate as SO4	10.6	0.50	mg/l	10.0		106	90-110			
Chloride	10.4	1.0	"	10.0		104	90-110			
Nitrate as NO3	10.3	0.50	"	10.0		103	90-110			
Nitrite as NO2	5.19	0.50	"	5.00		104	90-110			
Matrix Spike (9A27013-MS1)	Source: MSA	0515-01		Prepared &	Analyzed:	01/27/09				
Nitrate as NO3	20.9	5.0	mg/l	10.0	13.5	74	80-120			M2
Nitrite as NO2	3.10	0.50	"	5.00	ND	62	80-120			M2
Chloride	115	10	"	10.0	117	0	80-120			M2
Sulfate as SO4	139	5.0	"	10.0	144	0	80-120			M2
Matrix Spike Dup (9A27013-MSD1)	Source: MSA	0515-01		Prepared &	Analyzed:	01/27/09				
Chloride	116	10	mg/l	10.0	117	0	80-120	0.4	20	M2
Nitrate as NO3	21.2	5.0	"	10.0	13.5	77	80-120	2	20	M2
Nitrite as NO2	3.23	0.50	"	5.00	ND	65	80-120	4	20	M2
Sulfate as SO4	138	5.0	"	10.0	144	0	80-120	0.8	20	M2





TestAmerica San FranciscoProject:Work Sharing AgreementMSA05151220 Quarry LaneProject Number:720-17838Reported:Pleasanton CA, 94566-4756Project Manager:Melissa Brewer01/30/09 14:53

Notes and Definitions

M2	The MS and/or MSD were below the acceptance limits due to sample matrix interferen	ce See Blank Snike (LCS)
1V12	The MS and/of MSD were below the acceptance mints due to sample matrix interioren	cc. See Blank Spike (LCS).

DET Analyte DETECTED

ND Analyte NOT DETECTED at or above the reporting limit or MDL, if MDL is specified

NR Not Reported

dry Sample results reported on a dry weight basis

RPD Relative Percent Difference

MSA0523 MSA03159

TESTAMERICA San Francisco Chain of Custody 1220 Quarry Lane • Pleasariton CA 94566-4756

Phone: (925) 484-1919 • Fax: (925) 600-3002

P/u 1/26/09 Date SUB TO: MH THE LEADER IN ENVIRONMENTAL TESTING Report To Analysis Request 608 608 Low Level Metals by EPA 200.0/6020 (ICP-MS): C RCRA Company: TEST AMERICA EPA 8081 EPA 8082 Address: Metals: 🗅 Lead 🗅 LUFT CAM17 Metals (EPA 6010/7470/7471) Semivolatiles GC/MS □ EPA 8270 □ 625 W.E.T (STLC) TCLP Phone: Email Volatile Organics (□ EPA 8260B □ Bill To: Sampled By: Pesticides PCBs Phone: Attn: Sample ID 10:02 W NON 11:43 14:00 1) Relinquished by: Project Info. Sample Receipt 3) Relinquished by: # of Containers Project Name: Signature Time Printed Name Date 720-17838 Credit Card# Conforms to record: Company 3) Received by: 72h Other 48h 4.0 1840 Dav Report: ☐ Routine ☐ Level 3 ☐ Evel 4 ☐ EDD ☐ State Tank Fund EDF Signature Signature Time ☐ Global ID AGARIL 26/00 PEOSYNTEL Date Printed Name Printed Name Date "JYMT is and Couditions on reverse Company Company dica SF reports 8515M hora € -C₂₄ industry norm). Default for 8015B is Rev 06/04 **TEST AMERICA SAMPLE RECEIPT LOG**

CLIENT NAME: TAST REC. BY (PRINT) WORKORDER: Lagged in 1/27		DATE REC'D AT LAB: TIME REC'D AT LAB: DATE LOGGED IN:	126 109 1940 -128109	1/27/00			WAST	KING W	/ATER
CIRCLE THE APPROPRIATE RESPONS	E LAB SAMPLE#	. CLIĘNT ID	CONTAINER DESCRIPTION	PRESER VATIVE	рН**	SAMPLE MATRIX	DATE SAMPLED	Temp. >6°C	REMARKS: CONDITION
Custody Seal(s) Present / Absent Intact / Broken*								***************************************	
2. Chain-of-Custody Present / Absent*									
Traffic Reports or Packing List: Present / Absent	·								
4. Airbill / Sticker - Present / Absent Tracking #									
5. Sample Condition: Intact/Leaking*/Broken*									
6. Samples labeled Yes No*									
7. Sample ID's listed on COC Yes No*									
8. Does information on COC and sample									
labels agree? (Yes No*						,			
9. Sample received within	· · · · · · · · · · · · · · · · · · ·				3	,			
hold time: Yes No*				5:/(/				
10. Adequate sample volume received Yes / No*			Jet J	$\frac{2}{\sqrt{2}}$		-c/			
11. Proper preservatives used Yes No*					Ċ) 			
12. Trip Blank / Temp Blank Received?		٠,	v. /	22	7				
(circle which if yes) Yes / No	/	ال	*	4					<u> </u>
13. Thermometer Used : IR-1 / IR-2/ Backup		J 1 /							
14. Cooler RT*** CF*** CT***		201				-			
1 <u>3·8 ~-i·) 고영</u>		. /		•					
2		_nt	a						
3		Ø /							
4									
5									
15. Is/Are corrected temp 0-6°C? (es / No*									
**Exception (if any): Metals / Perchlorate /							,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		
Win 24hrs of sampling-on ice / Problem COC		LED, CONTACT PROJECT							

SAMPLERECEIPTLOG

Page ____ of ___

^{**}CHECK SAMPLE PREP LOG IF NOT INDICATED

^{***} Read Temperature/Correction Factor/Corrected Temperature



3/30/2009

Ms. Angela Liang GeoSyntec Consultants 475 14th Street Suite 400 Oakland CA 94612

Project Name: Hopyard Project #: WR0574/SVE Workorder #: 0903353

Dear Ms. Angela Liang

The following report includes the data for the above referenced project for sample(s) received on 3/13/2009 at Air Toxics Ltd.

The data and associated QC analyzed by Modified TO-15 are compliant with the project requirements or laboratory criteria with the exception of the deviations noted in the attached case narrative.

Thank you for choosing Air Toxics Ltd. for you air analysis needs. Air Toxics Ltd. is committed to providing accurate data of the highest quality. Please feel free to contact the Project Manager: Kyle Vagadori at 916-985-1000 if you have any questions regarding the data in this report.

Regards,

Kyle Vagadori Project Manager

Kya Vych



WORK ORDER #: 0903353

Work Order Summary

CLIENT: Ms. Angela Liang BILL TO: Ms. Angela Liang

GeoSyntec Consultants GeoSyntec Consultants

Oakland, CA 94612 Oakland, CA 94612

PHONE: 510-836-3034 **P.O.** # WR0574

FAX: 510-836-3036 **PROJECT #** WR0574/SVE Hopyard

DATE RECEIVED: 03/13/2009 **CONTACT:** Kyle Vagadori **DATE COMPLETED:** 03/27/2009

			RECEIPT	FINAL
FRACTION #	<u>NAME</u>	<u>TEST</u>	VAC./PRES.	PRESSURE
01A	SVE-Infl	Modified TO-15	3.8 "Hg	15 psi
02A	Lab Blank	Modified TO-15	NA	NA
03A	CCV	Modified TO-15	NA	NA
04A	LCS	Modified TO-15	NA	NA

CERTIFIED BY:

Sinda d. Fruman

DATE: 03/30/09

Laboratory Director

Certification numbers: CA NELAP - 02110CA, LA NELAP/LELAP- AI 30763, NJ NELAP - CA004 NY NELAP - 11291, UT NELAP - 9166389892, AZ Licensure AZ0719

Name of Accrediting Agency: NELAP/Florida Department of Health, Scope of Application: Clean Air Act,

Accreditation number: E87680, Effective date: 07/01/08, Expiration date: 06/30/09

Air Toxics Ltd. certifies that the test results contained in this report meet all requirements of the NELAC standards

This report shall not be reproduced, except in full, without the written approval of Air Toxics Ltd.



LABORATORY NARRATIVE Modified TO-15 GeoSyntec Consultants Workorder# 0903353

One 1 Liter Summa Canister sample was received on March 13, 2009. The laboratory performed analysis via modified EPA Method TO-15 using GC/MS in the full scan mode. The method involves concentrating up to 0.2 liters of air. The concentrated aliquot is then flash vaporized and swept through a water management system to remove water vapor. Following dehumidification, the sample passes directly into the GC/MS for analysis.

This workorder was independently validated prior to submittal using 'USEPA National Functional Guidelines' as generally applied to the analysis of volatile organic compounds in air. A rules-based, logic driven, independent validation engine was employed to assess completeness, evaluate pass/fail of relevant project quality control requirements and verification of all quantified amounts.

Method modifications taken to run these samples are summarized in the table below. Specific project requirements may over-ride the ATL modifications.

Requirement	TO-15	ATL Modifications
Daily CCV	= 30% Difference</td <td><!--= 30% Difference; Compounds exceeding this criterion and associated data are flagged and narrated.</p--></td>	= 30% Difference; Compounds exceeding this criterion and associated data are flagged and narrated.</p
Sample collection media	Summa canister	ATL recommends use of summa canisters to insure data defensibility, but will report results from Tedlar bags at client request
Method Detection Limit	Follow 40CFR Pt.136 App. B	The MDL met all relevant requirements in Method TO-15 (statistical MDL less than the LOQ). The concentration of the spiked replicate may have exceeded 10X the calculated MDL in some cases

Receiving Notes

There were no receiving discrepancies.

Analytical Notes

There were no analytical discrepancies.

Definition of Data Qualifying Flags

Eight qualifiers may have been used on the data analysis sheets and indicates as follows:

- B Compound present in laboratory blank greater than reporting limit (background subtraction no performed).
 - J Estimated value.
 - E Exceeds instrument calibration range.
 - S Saturated peak.
 - O Exceeds quality control limits.
 - U Compound analyzed for but not detected above the reporting limit.



- UJ- Non-detected compound associated with low bias in the CCV
- N The identification is based on presumptive evidence.

File extensions may have been used on the data analysis sheets and indicates as follows:

- a-File was requantified
- b-File was quantified by a second column and detector
- r1-File was requantified for the purpose of reissue



Summary of Detected Compounds MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

Client Sample ID: SVE-Infl

Lab ID#: 0903353-01A

Compound	Rɒt. Limit (ppbv)	Amount (ppbv)	Rpt. Limit (ug/m3)	Amount (ug/m3)
Acetone	6.2	7.9	15	19
2-Butanone (Methyl Ethyl Ketone)	1.5	2.6	4.5	7.5
Tetrahydrofuran	1.5	2.9	4.5	8.6
Trichloroethene	1.5	6.8	8.3	36
Tetrachloroethene	1.5	250	10	1700



Client Sample ID: SVE-Infl Lab ID#: 0903353-01A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name: y032029 Date of Collection: 3/12/09 9:12:00 AM
Dil. Factor: 3.08 Date of Analysis: 3/21/09 05:12 AM

DII. Factor:	3.08	Date	Date of Analysis: 3/21/09 05:12 AW				
Compound	Rpt. Limit (ppbv)	Amount (ppbv)	Rpt. Limit (ug/m3)	Amount (ug/m3)			
Freon 12	1.5	Not Detected	7.6	Not Detected			
Freon 114	1.5	Not Detected	11	Not Detected			
Chloromethane	6.2	Not Detected	13	Not Detected			
Vinyl Chloride	1.5	Not Detected	3.9	Not Detected			
1,3-Butadiene	1.5	Not Detected	3.4	Not Detected			
Bromomethane	1.5	Not Detected	6.0	Not Detected			
Chloroethane	1.5	Not Detected	4.1	Not Detected			
Freon 11	1.5	Not Detected	8.6	Not Detected			
Ethanol	6.2	Not Detected	12	Not Detected			
Freon 113	1.5	Not Detected	12	Not Detected			
1,1-Dichloroethene	1.5	Not Detected	6.1	Not Detected			
Acetone	6.2	7.9	15	19			
2-Propanol	6.2	Not Detected	15	Not Detected			
Carbon Disulfide	1.5	Not Detected	4.8	Not Detected			
3-Chloropropene	6.2	Not Detected	19	Not Detected			
Methylene Chloride	1.5	Not Detected	5.4	Not Detected			
Methyl tert-butyl ether	1.5	Not Detected	5.6	Not Detected			
rans-1,2-Dichloroethene	1.5	Not Detected	6.1	Not Detected			
Hexane	1.5	Not Detected	5.4	Not Detected			
1,1-Dichloroethane	1.5	Not Detected	6.2	Not Detected			
2-Butanone (Methyl Ethyl Ketone)	1.5	2.6	4.5	7.5			
cis-1,2-Dichloroethene	1.5	Not Detected	6.1	Not Detected			
Tetrahydrofuran	1.5	2.9	4.5	8.6			
Chloroform	1.5	Not Detected	7.5	Not Detected			
1,1,1-Trichloroethane	1.5	Not Detected	8.4	Not Detected			
Cyclohexane	1.5	Not Detected	5.3	Not Detected			
Carbon Tetrachloride	1.5	Not Detected	9.7	Not Detected			
2,2,4-Trimethylpentane	1.5	Not Detected	7.2	Not Detected			
Benzene	1.5	Not Detected	4.9	Not Detected			
1,2-Dichloroethane	1.5	Not Detected	6.2	Not Detected			
Heptane	1.5	Not Detected	6.3	Not Detected			
Trichloroethene	1.5	6.8	8.3	36			
1,2-Dichloropropane	1.5	Not Detected	7.1	Not Detected			
1,4-Dioxane	6.2	Not Detected	22	Not Detected			
Bromodichloromethane	1.5	Not Detected	10	Not Detected			
cis-1,3-Dichloropropene	1.5	Not Detected	7.0	Not Detected			
4-Methyl-2-pentanone	1.5	Not Detected	6.3	Not Detected			
Toluene	1.5	Not Detected	5.8	Not Detected			
trans-1,3-Dichloropropene	1.5	Not Detected	7.0	Not Detected			



Client Sample ID: SVE-Infl Lab ID#: 0903353-01A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name:	y032029	Date of Collection: 3/12/09 9:12:00 AM
Dil. Factor:	3.08	Date of Analysis: 3/21/09 05:12 AM

Compound	Rpt. Limit (ppbv)	Amount (ppbv)	Rpt. Limit (ug/m3)	Amount (ug/m3)
1,1,2-Trichloroethane	1.5	Not Detected	8.4	Not Detected
Tetrachloroethene	1.5	250	10	1700
2-Hexanone	6.2	Not Detected	25	Not Detected
Dibromochloromethane	1.5	Not Detected	13	Not Detected
1,2-Dibromoethane (EDB)	1.5	Not Detected	12	Not Detected
Chlorobenzene	1.5	Not Detected	7.1	Not Detected
Ethyl Benzene	1.5	Not Detected	6.7	Not Detected
m,p-Xylene	1.5	Not Detected	6.7	Not Detected
o-Xylene	1.5	Not Detected	6.7	Not Detected
Styrene	1.5	Not Detected	6.6	Not Detected
Bromoform	1.5	Not Detected	16	Not Detected
Cumene	1.5	Not Detected	7.6	Not Detected
1,1,2,2-Tetrachloroethane	1.5	Not Detected	10	Not Detected
Propylbenzene	1.5	Not Detected	7.6	Not Detected
4-Ethyltoluene	1.5	Not Detected	7.6	Not Detected
1,3,5-Trimethylbenzene	1.5	Not Detected	7.6	Not Detected
1,2,4-Trimethylbenzene	1.5	Not Detected	7.6	Not Detected
1,3-Dichlorobenzene	1.5	Not Detected	9.2	Not Detected
1,4-Dichlorobenzene	1.5	Not Detected	9.2	Not Detected
alpha-Chlorotoluene	1.5	Not Detected	8.0	Not Detected
1,2-Dichlorobenzene	1.5	Not Detected	9.2	Not Detected
1,2,4-Trichlorobenzene	6.2	Not Detected	46	Not Detected
Hexachlorobutadiene	6.2	Not Detected	66	Not Detected

Container Type: 1 Liter Summa Canister

,		Method	
Surrogates	%Recovery	Limits	
Toluene-d8	102	70-130	
1,2-Dichloroethane-d4	104	70-130	
4-Bromofluorobenzene	95	70-130	



Client Sample ID: Lab Blank Lab ID#: 0903353-02A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name:	y032005	Date of Collection: NA
Dil. Factor:	1.00	Date of Analysis: 3/20/09 10:40 AM

Dil. Factor:	1.00	Date	Date of Analysis: 3/20/09 10:40 AM		
Compound	Rpt. Limit (ppbv)	Amount (ppbv)	Rpt. Limit (ug/m3)	Amount (ug/m3)	
Freon 12	0.50	Not Detected	2.5	Not Detected	
Freon 114	0.50	Not Detected	3.5	Not Detected	
Chloromethane	2.0	Not Detected	4.1	Not Detected	
Vinyl Chloride	0.50	Not Detected	1.3	Not Detected	
1,3-Butadiene	0.50	Not Detected	1.1	Not Detected	
Bromomethane	0.50	Not Detected	1.9	Not Detected	
Chloroethane	0.50	Not Detected	1.3	Not Detected	
Freon 11	0.50	Not Detected	2.8	Not Detected	
Ethanol	2.0	Not Detected	3.8	Not Detected	
Freon 113	0.50	Not Detected	3.8	Not Detected	
1,1-Dichloroethene	0.50	Not Detected	2.0	Not Detected	
Acetone	2.0	Not Detected	4.8	Not Detected	
2-Propanol	2.0	Not Detected	4.9	Not Detected	
Carbon Disulfide	0.50	Not Detected	1.6	Not Detected	
3-Chloropropene	2.0	Not Detected	6.3	Not Detected	
Methylene Chloride	0.50	Not Detected	1.7	Not Detected	
Methyl tert-butyl ether	0.50	Not Detected	1.8	Not Detected	
trans-1,2-Dichloroethene	0.50	Not Detected	2.0	Not Detected	
Hexane	0.50	Not Detected	1.8	Not Detected	
1,1-Dichloroethane	0.50	Not Detected	2.0	Not Detected	
2-Butanone (Methyl Ethyl Ketone)	0.50	Not Detected	1.5	Not Detected	
cis-1,2-Dichloroethene	0.50	Not Detected	2.0	Not Detected	
Tetrahydrofuran	0.50	Not Detected	1.5	Not Detected	
Chloroform	0.50	Not Detected	2.4	Not Detected	
1,1,1-Trichloroethane	0.50	Not Detected	2.7	Not Detected	
Cyclohexane	0.50	Not Detected	1.7	Not Detected	
Carbon Tetrachloride	0.50	Not Detected	3.1	Not Detected	
2,2,4-Trimethylpentane	0.50	Not Detected	2.3	Not Detected	
Benzene	0.50	Not Detected	1.6	Not Detected	
1,2-Dichloroethane	0.50	Not Detected	2.0	Not Detected	
Heptane	0.50	Not Detected	2.0	Not Detected	
Trichloroethene	0.50	Not Detected	2.7	Not Detected	
1,2-Dichloropropane	0.50	Not Detected	2.3	Not Detected	
1,4-Dioxane	2.0	Not Detected	7.2	Not Detected	
Bromodichloromethane	0.50	Not Detected	3.4	Not Detected	
cis-1,3-Dichloropropene	0.50	Not Detected	2.3	Not Detected	
4-Methyl-2-pentanone	0.50	Not Detected	2.0	Not Detected	
Toluene	0.50	Not Detected	1.9	Not Detected	
trans-1,3-Dichloropropene	0.50	Not Detected	2.3	Not Detected	



Client Sample ID: Lab Blank Lab ID#: 0903353-02A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name:	y032005 Date of Collection: NA			
Dil. Factor:	1.00	Date	of Analysis: 3/20/	09 10:40 AM
Compound	Rpt. Limit (ppbv)	Amount (ppbv)	Rpt. Limit (ug/m3)	Amount (ug/m3)
1,1,2-Trichloroethane	0.50	Not Detected	2.7	Not Detected
Tetrachloroethene	0.50	Not Detected	3.4	Not Detected
2-Hexanone	2.0	Not Detected	8.2	Not Detected
Dibromochloromethane	0.50	Not Detected	4.2	Not Detected
1,2-Dibromoethane (EDB)	0.50	Not Detected	3.8	Not Detected
Chlorobenzene	0.50	Not Detected	2.3	Not Detected
Ethyl Benzene	0.50	Not Detected	2.2	Not Detected
m,p-Xylene	0.50	Not Detected	2.2	Not Detected
o-Xylene	0.50	Not Detected	2.2	Not Detected
Styrene	0.50	Not Detected	2.1	Not Detected
Bromoform	0.50	Not Detected	5.2	Not Detected
Cumene	0.50	Not Detected	2.4	Not Detected
1,1,2,2-Tetrachloroethane	0.50	Not Detected	3.4	Not Detected
Propylbenzene	0.50	Not Detected	2.4	Not Detected
4-Ethyltoluene	0.50	Not Detected	2.4	Not Detected
1,3,5-Trimethylbenzene	0.50	Not Detected	2.4	Not Detected
1,2,4-Trimethylbenzene	0.50	Not Detected	2.4	Not Detected
1,3-Dichlorobenzene	0.50	Not Detected	3.0	Not Detected
1,4-Dichlorobenzene	0.50	Not Detected	3.0	Not Detected
alpha-Chlorotoluene	0.50	Not Detected	2.6	Not Detected
1,2-Dichlorobenzene	0.50	Not Detected	3.0	Not Detected
1,2,4-Trichlorobenzene	2.0	Not Detected	15	Not Detected
Hexachlorobutadiene	2.0	Not Detected	21	Not Detected
Container Type: NA - Not Applicable				
Surrogates		%Recovery		Method Limits
Toluene-d8		99		70-130
1,2-Dichloroethane-d4		109		70-130
4-Bromofluorobenzene		98		70-130



Client Sample ID: CCV Lab ID#: 0903353-03A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name: y032003 Date of Collection: NA
Dil. Factor: 1.00 Date of Analysis: 3/20/09 09:12 AM

Compound	%Recovery
Freon 12	96
Freon 114	98
Chloromethane	99
Vinyl Chloride	95
1,3-Butadiene	104
Bromomethane	112
Chloroethane	102
Freon 11	98
Ethanol	102
Freon 113	98
1,1-Dichloroethene	95
Acetone	99
2-Propanol	104
Carbon Disulfide	106
3-Chloropropene	112
Methylene Chloride	100
Methyl tert-butyl ether	88
trans-1,2-Dichloroethene	100
Hexane	97
1,1-Dichloroethane	105
2-Butanone (Methyl Ethyl Ketone)	108
cis-1,2-Dichloroethene	104
Tetrahydrofuran	103
Chloroform	102
1,1,1-Trichloroethane	112
Cyclohexane	101
Carbon Tetrachloride	107
2,2,4-Trimethylpentane	101
Benzene	102
1,2-Dichloroethane	102
Heptane	102
Trichloroethene	102
1,2-Dichloropropane	102
1,4-Dioxane	103
Bromodichloromethane	105
cis-1,3-Dichloropropene	110
4-Methyl-2-pentanone	109
Toluene	104
trans-1,3-Dichloropropene	112



Client Sample ID: CCV Lab ID#: 0903353-03A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name: y032003 Date of Collection: NA
Dil. Factor: 1.00 Date of Analysis: 3/20/09 09:12 AM

1,1,2-Trichloroethane Tetrachloroethene 2-Hexanone Dibromochloromethane 1,2-Dibromoethane (EDB) Chlorobenzene Ethyl Benzene m,p-Xylene o-Xylene Styrene Bromoform Cumene 1,1,2,2-Tetrachloroethane Propylbenzene 4-Ethyltoluene 1,3,5-Trimethylbenzene 1,2,4-Trimethylbenzene	102 103 103 110 108
2-Hexanone Dibromochloromethane 1,2-Dibromoethane (EDB) Chlorobenzene Ethyl Benzene m,p-Xylene o-Xylene Styrene Bromoform Cumene 1,1,2,2-Tetrachloroethane Propylbenzene 4-Ethyltoluene 1,3,5-Trimethylbenzene	103 110 108
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Styrene Bromoform Cumene 1,1,2,2-Tetrachloroethane Propylbenzene 4-Ethyltoluene 1,3,5-Trimethylbenzene	102
Bromoform Cumene 1,1,2,2-Tetrachloroethane Propylbenzene 4-Ethyltoluene 1,3,5-Trimethylbenzene	104
Cumene 1,1,2,2-Tetrachloroethane Propylbenzene 4-Ethyltoluene 1,3,5-Trimethylbenzene	117
1,1,2,2-Tetrachloroethane Propylbenzene 4-Ethyltoluene 1,3,5-Trimethylbenzene	114
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4-Ethyltoluene 1,3,5-Trimethylbenzene	107
1,3,5-Trimethylbenzene	105
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1.2.4 Trimothylbonzono	112
1,2,4-11111ettiyibetizette	112
1,3-Dichlorobenzene	109
1,4-Dichlorobenzene	111
alpha-Chlorotoluene	120
1,2-Dichlorobenzene	110
1,2,4-Trichlorobenzene	105
Hexachlorobutadiene	100

Container Type: NA - Not Applicable

26.		Method
Surrogates	%Recovery	Limits
Toluene-d8	102	70-130
1,2-Dichloroethane-d4	100	70-130
4-Bromofluorobenzene	103	70-130



Client Sample ID: LCS Lab ID#: 0903353-04A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name: y032004 Date of Collection: NA
Dil. Factor: 1.00 Date of Analysis: 3/20/09 09:53 AM

Compound	%Recovery
Freon 12	94
Freon 114	97
Chloromethane	92
Vinyl Chloride	94
1,3-Butadiene	99
Bromomethane	86
Chloroethane	80
Freon 11	99
Ethanol	79
Freon 113	109
1,1-Dichloroethene	105
Acetone	115
2-Propanol	106
Carbon Disulfide	105
3-Chloropropene	110
Methylene Chloride	107
Methyl tert-butyl ether	93
trans-1,2-Dichloroethene	103
Hexane	101
1,1-Dichloroethane	112
2-Butanone (Methyl Ethyl Ketone)	114
cis-1,2-Dichloroethene	113
Tetrahydrofuran	105
Chloroform	108
1,1,1-Trichloroethane	118
Cyclohexane	106
Carbon Tetrachloride	111
2,2,4-Trimethylpentane	104
Benzene	106
1,2-Dichloroethane	107
Heptane	106
Trichloroethene	106
1,2-Dichloropropane	104
1,4-Dioxane	102
Bromodichloromethane	110
cis-1,3-Dichloropropene	112
4-Methyl-2-pentanone	113
Toluene	113
trans-1,3-Dichloropropene	113



Client Sample ID: LCS Lab ID#: 0903353-04A

MODIFIED EPA METHOD TO-15 GC/MS FULL SCAN

File Name: y032004 Date of Collection: NA
Dil. Factor: 1.00 Date of Analysis: 3/20/09 09:53 AM

Compound	%Recovery
1,1,2-Trichloroethane	104
Tetrachloroethene	105
2-Hexanone	98
Dibromochloromethane	113
1,2-Dibromoethane (EDB)	107
Chlorobenzene	105
Ethyl Benzene	103
m,p-Xylene	102
o-Xylene	106
Styrene	115
Bromoform	118
Cumene	112
1,1,2,2-Tetrachloroethane	110
Propylbenzene	108
4-Ethyltoluene	108
1,3,5-Trimethylbenzene	109
1,2,4-Trimethylbenzene	110
1,3-Dichlorobenzene	110
1,4-Dichlorobenzene	109
alpha-Chlorotoluene	120
1,2-Dichlorobenzene	109
1,2,4-Trichlorobenzene	102
Hexachlorobutadiene	97

Container Type: NA - Not Applicable

21.		Method	
Surrogates	%Recovery	Limits	
Toluene-d8	103	70-130	
1,2-Dichloroethane-d4	100	70-130	
4-Bromofluorobenzene	101	70-130	



Sample Transportation Notice

Relinquishing signature on this document indicates that sample is being shipped in compliance with 180 BLUE RAVINE ROAD, SUITE B all applicable local, State, Federal, national, and international laws, regulations and ordinances of any kind. Air Toxics Limited assumes no liability with respect to the collection, handling or shipping of these samples. Relinquishing signature also indicates agreement to hold harmless defend, and indome fy Air Toxics Limited against any claim, demand, or action, of any kind, related to the collection, handling, or shipping of samples, D.O.T. Holline (800) 467-4922.

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APPENDIX C

EISB Gene-Trac Laboratory Analytical Report



130 Research Lane, Suite 2 Guelph, Ontario N1G 5G3 Phone (519) 822-2265 Fax (519) 822-3151

Certificate of Analysis: Quantitative Gene-Trac Dehalococcoides Assay

Customer: Angela Liang & Melissa Asher, Geosyntec Consultants SIREM Reference: S-1515

Project: Hopyard Cleaners **Report Issued:** 18-Feb-09

Customer Reference: WR0574 Data Files: QPCR-0443

Table 1: Test Results

Customer Sample II	SiREM Sample ID	Sample Collection Date	Sample Matrix	Percent Dhc ^A	Dehalococcoides Enumeration B
MW-3	DHC-4790	26-Jan-09	Groundwater	0.004-0.01%	1 x 10 ⁵ /liter
MW-1	DHC-4791	26-Jan-09	Groundwater	0.008-0.03%	2 x 10 ⁵ /liter
MW-2	DHC-4792	26-Jan-09	Groundwater	0.005-0.02%	2 x 10 ⁵ /liter
MW-DUP	DHC-4793	26-Jan-09	Groundwater	0.007-0.02%	2 x 10 ⁵ /liter

Notes:

Analyst:

Jennifer Wilkinson

Biotechnology Technologist

Approved:

Ximena Druar, B.Sc.

Molecular Biology Coordinator

^A Percent *Dehalococcoides* (Dhc) in microbial population. This value is calculated by dividing the number of Dhc 16S ribosomal ribonucleic acid (rRNA) gene copies by the total number of bacteria as estimated by the mass of DNA extracted from the sample.

^BBased on quantification of Dhc 16S rRNA gene copies. Dhc are generally reported to contain one 16S rRNA gene copy per cell; therefore, this number is often interpreted to represent the number of Dhc cells present in the sample.



130 Research Lane, Suite 2 Guelph, Ontario N1G 5G3 Phone (519) 822-2265 Fax (519) 822-3151

Certificate of Analysis: Gene-Trac-VC, Vinyl Chloride Reductase (vcrA) Assay

Customer: Angela Liang & Melissa Asher, Geosyntec SiREM Reference: S-1515

Project: Hopyard Cleaners Report Issued: 19-Feb-09

Customer Reference: WR0574 Data Files: VC-QPCR-0179

VC-QPCR check-gel-0202

Table 1: Test Results

Customer Sample ID	SiREM Sample ID	Sample Collection Date	Sample Matrix	Percent <i>vcrA</i> ^A	Vinyl Chloride Reductase (<i>vcrA</i>) Gene Copies
MW-3	VCR-1155	26-Jan-09	Groundwater	NA ⁽¹⁾	Detected ⁽²⁾
MW-1	VCR-1156	26-Jan-09	Groundwater	NA ⁽¹⁾	Detected ⁽²⁾
MW-2	VCR-1157	26-Jan-09	Groundwater	NA ⁽¹⁾	Inconclusive ⁽³⁾
MW-DUP	VCR-1158	26-Jan-09	Groundwater	0.0009-0.003%	3 x 10 ⁴ /liter

Notes:

Analyst:

Jennifer Wilkinson

J. Wilkinson

Biotechnology Technologist

Approved:

Ximena Druar, B.Sc.

Molecular Biology Coordinator

^A Percent *vcrA* in microbial population. This value is calculated by dividing the number of vinyl chloride reductase A (*vcrA*) gene copies quantified by the total number of bacteria estimated to be in the sample based on the mass of DNA extracted from the sample. Range represents normal variation in enumeration of *vcrA*.

¹Not applicable as *vcrA* detected but not quantifiable.

²Detected but not quantifiable. The sample specific quantitation limit is 2 x 10⁴/liter.

 $^{^{3}}$ Inconclusive results may indicate extremely low concentrations of *vcrA* DNA at or below the sample specific quantitation limit (1 x 10^{4} /liter), however, test results were insufficient to assign a conclusive positive result for this sample.



Table 2: Detailed Test Parameters, Gene-Trac Test Reference S-1515

Customer Sample ID	MW-3	MW-1	MW-2	MW-DUP		
SiREM Sample ID	DHC-4790	DHC-4791	DHC-4792	DHC-4793		
Date Received	28-Jan-09	24-Oct-08	24-Oct-08	24-Oct-08		
Sample Temperature	1.4 °C	1.4 °C	1.4 °C	1.4 °C		
Volume Used for DNA Extraction	100 mL	100 mL	100 mL	100 mL		
DNA Extraction Date	18-Feb-09	18-Feb-09	18-Feb-09	18-Feb-09		
DNA Concentration in Sample (extractable)	7043 ng/L	5213 ng/L	6405 ng/L	6353 ng/L		
PCR Amplifiable DNA	Detected	Detected	Detected	Detected		
Dhc qPCR Analysis Date	18-Feb-09	18-Feb-09	18-Feb-09	18-Feb-09		
vcrA qPCR Analysis Date	18-Feb-09	18-Feb-09	18-Feb-09	18-Feb-09		
qPCR Controls (see Table 3 & 4)	Passed	Passed	Passed	Passed		
Comments						

Notes:

Refer to Table 3 for detailed results of controls. ND = not detected

°C = degrees Celsius

PCR = polymerase chain reaction qPCR = quantitative PCR Dhc = Dehalococcoides ng/L = nanograms per liter mL = milliliters

DNA = Deoxyribonucleic acid



Table 3: Gene-Trac-DHC Experimental Control Results, Gene-Trac Test Reference S-1515

Laboratory Control	Analysis Date	Control Description	Spiked Dhc 16S rRNA Gene Copies per Liter	Recovered Dhc 16S rRNA Gene Copies per Liter	Comments
Positive Control Low Concentration	18-Feb-09	qPCR with KB1 genomic DNA (CSLD-0085)	1.7 x 10 ⁶	2.2 x 10 ⁶	
Positive Control High Concentration	18-Feb-09	qPCR with KB1 genomic DNA (CSHD-0085)	2.2 x 10 ⁸	2.4 x 10 ⁸	
DNA Extraction Blank	18-Feb-09	DNA extraction sterile water (DB-0921)	0	8.6 x 10 ²	See Note 1
Negative Control	18-Feb-09	Tris Reagent Blank (TBD-0045)	0	ND	

Notes:

Dhc = Dehalococcoides

DNA = Deoxyribonucleic acid

NA = not applicable

ND = not detected

qPCR = quantitative PCR

16S rRNA = 16S ribosomal ribonucleic acid

¹ Deemed acceptable as test results for relevant samples are greater than 2 orders of magnitude above DNA Extraction Blank test result.



Table 4: Gene-Trac-VC Experimental Control Results, Gene-Trac Test Reference S-1515

Laboratory Control	Analysis Date	Control Description	Spiked <i>vcrA</i> reductase Gene Copies per Liter	Recovered vcrA reductase Gene Copies per Liter	Comments
Positive Control Low Concentration	18-Feb-09	qPCR with KB1 genomic DNA (CSLV-0047)	1.1 x 10 ⁶	1.5 x 10 ⁶	
Positive Control High Concentration	18-Feb-09	qPCR with KB1 genomic DNA (CSHV-0047)	1.6 x 10 ⁸	1.4 x 10 ⁸	
DNA Extraction Blank	18-Feb-09	DNA extraction sterile water (DB-0921)	0	ND	
Negative Control	18-Feb-09	Tris Reagent Blank (TBV-0018)	0	ND	

Notes:

NA = not applicable

ND = not detected

qPCR = quantitative PCR

Dhc = Dehalococcoides

DNA = Deoxyribonucleic acid

16S rRNA = 16S ribosomal ribonucleic acid

vcrA = vinyl chloride reductase

¹ Within defined limits of +/- 50%

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Interpretation of Quantitative Gene-Trac Dehalococcoides Test Results

1) Background:

Dehalococcoides group organisms (*Dhc*) are the only known microorganisms capable of complete dechlorination of chloroethenes (i.e., tetrachloroethene, trichloroethene, *cis*-dichloroethene, vinyl chloride to non-toxic ethene. The detection of the *Dhc* 16S ribosomal ribonucleic acid (rRNA) gene has been correlated with the complete biological dechlorination of chlorinated ethenes to ethene at contaminated sites (Hendrickson et. al., 2002, *Applied and Environmental Microbiology*, 68: 485-495). The Quantitative Gene-Trac *Dehalococcoides* test is a quantitative polymerase chain reaction (PCR) test used to determine the concentration of the *Dhc* 16S rRNA gene in soil and groundwater samples.

2) Interpretation of Test Results:

The Quantitative Gene-Trac test reports two types of results, "Dehalococcoides 16S rRNA Gene Copies" is a raw value whereas "% Dehalococcoides in Microbial Population" is the raw value expressed as percentage of total microbial population. A detailed explanation of the two types of results is provided below.

a) Dehalococcoides 16S rRNA Gene Copies

This value is the direct number of *Dhc* 16S rRNA gene copies detected in the sample. Results may be reported either per liter (for groundwater) or per gram (for soil). This number is generally interpreted as equivalent to the number of viable *Dhc* present in the sample when certain reasonable assumptions are made, including that the DNA quantified belongs to viable *Dhc* (i.e., not from dead *Dhc*) and that each *Dhc* cell contains only one 16S rRNA gene. Guidelines for relating this value to observable dechlorination impacts for groundwater samples are provided below.

- Values of 10³ gene copies per liter or lower, indicate the sample contains low concentrations of *Dhc* organisms which may indicate that site conditions are suboptimal for high rates of dechlorination. Increases in *Dhc* concentrations at the site may be possible if conditions are modified (e.g., electron donor addition).
- Values of 10⁴-10⁶ gene copies per liter, indicates the sample contains moderate concentrations of *Dhc* which may, or may not, be associated with observable dechlorination impacts (i.e., ethene).
- Values at or above 10⁷ gene copies per liter, indicate the samples contains high concentrations of *Dhc* which is often associated with high rates of dechlorination and the production of ethene. Test results exceeding 10⁹ gene copies/liter are rarely observed.



b) % Dehalococcoides in Microbial Population (% Dhc)

This value presents the percentage of *Dhc* (% *Dhc*) relative to other microorganisms in the sample based on the formulas below. % *Dhc* is a measure of the predominance of *Dhc* and, in general, the higher this percentage the better.

%
$$Dhc = \frac{Number\ Dhc}{Number\ Dhc + Number\ other\ Bacteria}$$

Where:

Number other Bacteria =
$$\frac{Total\ DNA\ in\ sample\ (ng) - DNA\ attributed\ to\ Dhc\ (ng)}{4.0\ x\ 10^{-6}\ ng\ DNA\ per\ bacterial\ cell}$$

The number of non-Dhc bacteria is estimated by assuming each non-Dhc bacterium contains 4.0 x 10⁻⁶ nanograms (ng) of DNA (Paul and Clark. 1996. Soil Microbiology and Biochemistry). Because the total mass of DNA in a sample is determined (by fluorometry) the total number of bacteria present can be estimated. For perspective, the % Dhc can range from very low fractions of percentages, in samples that have low numbers of Dhc and high numbers of other bacteria (incompletely colonized by Dhc), to greater than 50% in Dhc enriched cultures such as KB-1TM (fully colonized by Dhc).

In addition to determining the predominance of *Dhc*, this value is also used for interpretation of *Dhc* counts from different sampling locations or the same location over time, because it is normalized to total bacteria. In particular, the % *Dhc* value can be used to correct *Dhc* counts where samples are biased low due to non-representative sampling of biomass (bacteria). Example 1 below illustrates a scenario where the % *Dhc* value improves the interpretation of data where one sampling event was biased.

Example 1, use of % Dhc Value to interpret raw data

Example 1 presents results from monitoring well MW-1 sampled in April, May and June. Based on the raw *Dhc* counts alone (*Dehalococcoides* 16S rRNA Gene Copies) it might be assumed that the number of *Dhc* decreased 10-fold between April and May; however, based on the percentage of *Dhc* it is clear that the proportion of *Dhc* actually increased from April to May and that the low count is probably a case of sampling variability (biased low). The higher raw count and the higher percentage of *Dhc* in June confirms the trend of increasing *Dhc* concentrations over time.

Sample	Dehalococcoides 16S rRNA Gene Copies	% Dhc	Interpretation Based on % <i>Dhc</i>
MW-1– April	1.0 x 10 ⁵ /Liter	0.1%	Dhc is a low proportion of total microbial population
MW-1- May	1.0 x 10 ⁴ /Liter	1%	<i>Dhc</i> predominance increased 10-fold from April, low count from low biomass sampled, non-biased sample would be [(1.0/0.1) x 1.0 x10 ⁵] = 10 ⁶ /Liter
MW-1 June	1.0 x 10 ⁷ /Liter	10%	Dhc predominance moderate and has increased 100-fold from April



3) Explanation of Notes

Quantitation limit: The quantitation limit of Gene-Trac test is 2,150 *Dhc* 16S rRNA gene copies per liter. Note, the specific quantitation limit for each test varies depending on the volume of sample used in the DNA extraction process. For example, if only a ½ liter of water was used the quantitation limit would increase two-fold to 4300 gene copies per liter. The specific quantitation limit is provided only where *Dhc* is not detected.

Value is an estimated quantity between the quantitation limit and detection limit: This is applicable in situations where *Dhc* DNA is detected above the detection limit, but below the quantitation limit, of the standard curve. In such cases an estimated value is provided which is based on extrapolation of the standard curve.

Sample inhibited testing: Each Quantitative Gene-Trac test includes a quantification of the amount of DNA extracted from the sample and a second test to determine if the extracted DNA is suitable for *Dhc* testing (PCR with a universal Bacteria primer). If a sample is determined to contain DNA and PCR with universal primers is negative, it suggests that the extracted DNA inhibited the PCR. Inhibition may be caused by compounds present in the original groundwater sample (e.g., humic acids). Where inhibition occurs there is an increased likelihood of false negatives since *Dhc* DNA, if present, may not be detected.

DNA not extracted from the sample: If DNA is not detected in the sample then "DNA not extracted from the sample" is reported. This is commonly due to samples that contain little or no biomass (bacteria). In some cases sampling may not capture bacteria (e.g., when attached bacteria are not dislodged from the aguifer matrix).

4) Converting Standard Gene-Trac to Dhc 16S rRNA Gene Copies/Liter

Quantitative Gene-Trac provides quantitative results in *Dhc* 16S rRNA Gene Copies/Liter, whereas standard Gene-Trac provides semi-quantitative results using a plus scale. Based on parallel analysis of standard versus Quantitative Gene-Trac estimates of the number of *Dhc* gene copies for each + score in the standard test were determined. Note, the conversion factors do not apply in all cases and are meant to be used as a rule of thumb for relating standard Gene-Trac results to Quantitative-Gene-Trac.

Estimated 16S rRNA Gene Copies/Liter for Standard Gene-Trac Intensity Scores

Standard Gene-Trac Intensity Score	Approximate Range of 16S rRNA Gene Copies/Liter
+	10 ³ -10 ⁵
++	10 ⁴ -10 ⁶
+++	10 ⁵ -10 ⁷
++++	10 ⁶ -10 ⁸