

QUARTERLY GROUNDWATER REPORT

**5800 CHRISTIE AVENUE,
EMERYVILLE, CALIFORNIA**

JULY 31, 1993

93 AUG -4 PM 12:58

SUBMITTED TO:

**MR. BRIAN OLIVA
ALAMEDA COUNTY HEALTH CARE SERVICES
HAZARDOUS MATERIALS DIVISION
80 SWAN WAY, ROOM 200
OAKLAND, CALIFORNIA 94621**

PREPARED FOR :

**CROLEY & HERRING INVESTMENT COMPANY
448 THARP DRIVE,
MORAGA, CALIFORNIA 94556**

PREPARED BY:

**ETS ENVIRONMENT & TECHNOLOGY SERVICES
2081 15TH STREET,
SAN FRANCISCO, CALIFORNIA 94114
TELEPHONE: 415-861-0810
FACIMILE: 415-861-3269**

376-3473

ETS ENVIRONMENT & TECHNOLOGY SERVICES

2081 15TH STREET, SAN FRANCISCO, CALIFORNIA 94114
PHONE 415-861-0810 FAX 415-861-3269

July 31, 1993

Mr. Dick Herring
President
Croley & Herring Investment Company
448 Tharp Avenue,
Moraga, California 94556

Subject: Quarterly Groundwater Report
5800 Christie Avenue, Emeryville, California

Dear Mr. Herring:

Enclosed please find a copy of the quarterly groundwater report for the July, 1993 water sampling period at the subject facility.

As per Alameda County Health Care Services and Bay Area Regional Water Quality Control Board request, an additional groundwater monitoring well MW4 was installed downgradient of well EW1. Water level and groundwater quality results of well MW4 are included in the attached report.

Please contact me if you have any question about this report.

Sincerely,

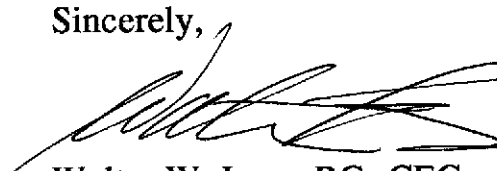

Walter W. Loo, RG CEG
President



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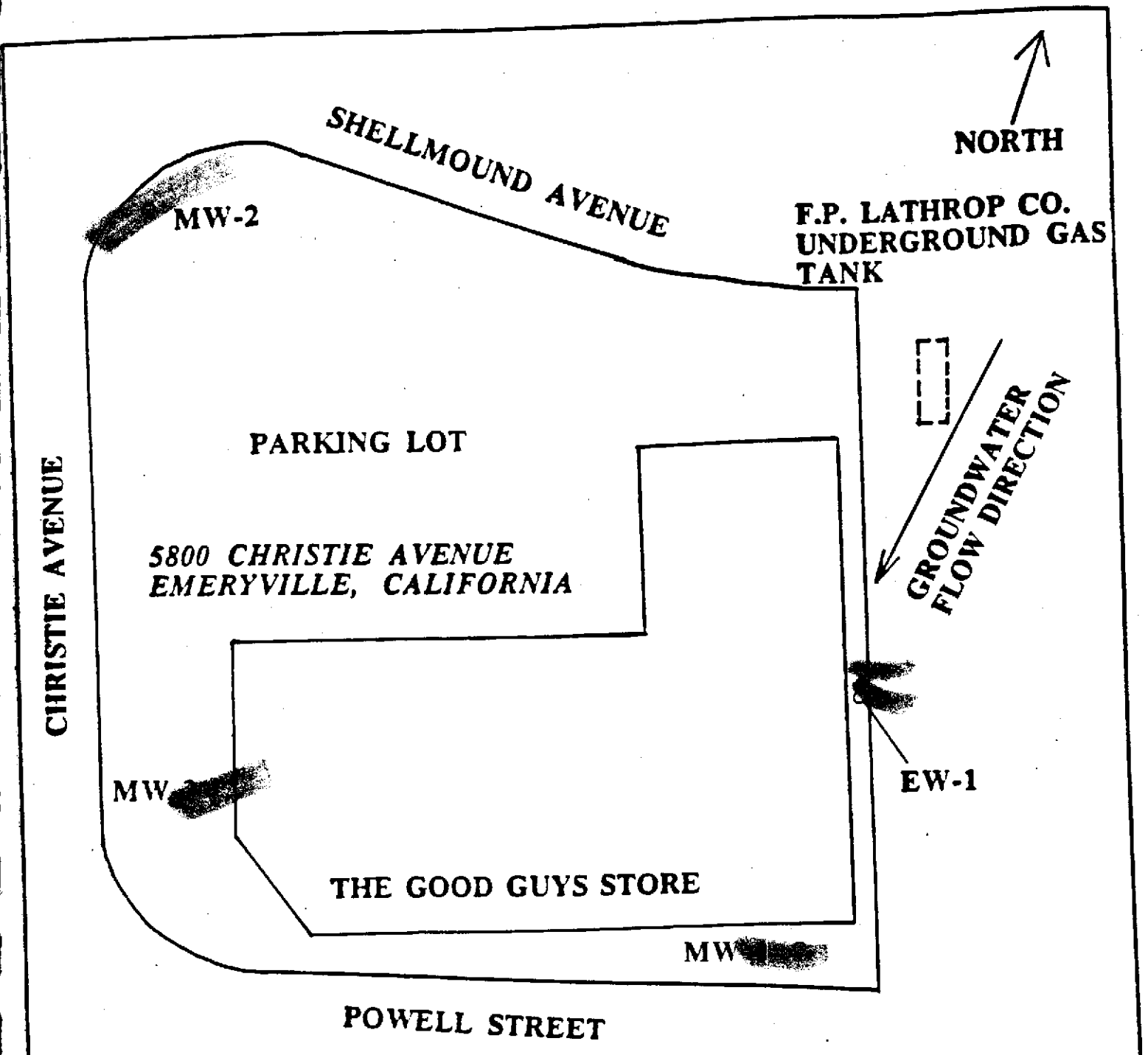
1.0 INTRODUCTION

Environment & Technology Services(ETS) was retained by Croley & Herring Investment Company to perform the 16th quarterly groundwater monitoring for the facility located at 5800 Christie Street in Emeryville, California. The subject facility is currently leased to an electronic merchandise retailer. Prior to leasing, soil contamination was identified at the subject facility. The contaminated soil was removed with the exception of that which was underlying the building because of safety concerns. The removed soil was remediated on-site and properly disposed of with the approval of the regulatory agencies.

A vapor extraction system(VES) was installed immediately adjacent to the northeastern side of the building to mitigate the residual volatile hydrocarbons contained in the soil. The residual volatile organic chemicals(VOCs) were remediated from an average VOCs concentration of about 660 ppm to a satisfactory level at an average of 0.82 ppm in soil. A soil closure plan was submitted(11/15/91) and approval of closure was received on 1/21/92 after submittal of confirmation soil sampling results. The soil vapor extraction system was decommissioned and the Bay Area Air Quality Management District was notified on 12/16/91. The final VES closure report was completed on August 29, 1992. An indoor vapor monitoring system Sierra Monitor Model 5000 was installed by the "Good Guys" electronic store in 1989 through March, 1993. No significant level of methane was detected for the monitoring period. The vapor monitoring system was disconnected in March, 1993 with the concurrence of Mr. Brian Oliva of Alameda County Health Care Services, March 15,1993 correspondence(Appendix A).

As part of the site activities, a quarterly groundwater monitoring program has been implemented. Previous quarterly monitoring events were conducted on November 6, 1989, February 13, 1990, May 15, 1990, August 14, 1990, November 7, 1990, February 13, 1991, May 15, 1991, August 14, 1991, November 7, 1991, February 13, 1992, May 15, 1992, August 14, 1992, November 7, 1992, February 13, 1993, May 15, 1993, August 14, 1993, and November 7, 1993 respectively. This quarterly monitoring event was conducted on July 7, 1993. Water samples were taken from the monitoring wells and sent to a State-certified laboratory for analysis under proper chain-of-custody procedures.

As per Alameda County Health Care Services and Bay Area Regional Water Quality Control Board request, an additional groundwater monitoring well MW4 was installed downgradient of well EW1. This report presents the results of this quarterly groundwater monitoring event on well EW1 and MW4 including laboratory analytical results, groundwater movement analysis, summary of findings, and conclusions and discussions. Boring and well log of MW4 is included in Appendix B.



LEGEND

○ MONITORING WELLS



ETS
ENVIRONMENT & TECHNOLOGY SERVICES

FIGURE 1
LOCATION MAP

2.0 GROUNDWATER MOVEMENT ANALYSIS

Prior to sample collection of this quarterly sampling, depth-to-water table in each of the three existing monitoring wells at the facility was measured for the analysis of groundwater movement. Table 1 presents a summary of the water levels in the three wells (EW1, MW2, MW3 and MW4) from the groundwater monitoring events prepared by ETS.

From the result of the water level measurements on July 7, 1983, water levels were decreased in the three wells, as compared to the results in April 1983. Nevertheless, the groundwater flow direction remains the same direction, flowing towards southwest (Figure 1). The hydraulic gradient was 0.013 feet per horizontal foot.

Groundwater movement across the facility remains in a similar pattern, as compared to the result from the previous sampling event. Data of flow direction and hydraulic gradient are summarized in Table 2.

TABLE 1

SUMMARY OF WATER LEVEL DATA

WELL Name	Elev. of TOC (Ft-MSL)	11/6/89		2/20/90		5/31/90		9/7/90	
		DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.
EW-1	8.62	6.15	2.47	5.93	2.69	5.86	2.76	6.30	2.32
MW-2	7.42	4.37	3.05	4.26	3.16	4.26	3.16	4.60	2.82
MW-3	6.42	5.10	1.32	5.42	1.00	4.93	1.49	5.15	1.17

WELL Name	12/4/90		4/16/91		7/3/91		10/14/91		1/9/92	
	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.
EW-1	7.39	2.23	6.02	2.60	6.20	2.42	6.5	2.12	6.20	2.42
MW-2	4.67	2.75	4.31	3.11	4.52	2.9	3.92	3.5	4.43	3.10
MW-3	5.96	1.35	5.25	1.17	5.33	1.09	4.63	1.79	6.50	-0.08

WELL Name	7/15/92		10/19/92		1/11/93		4/19/93	
	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.	DTW Ft.	SWL Ft.
EW-1	6.10	2.52	6.1	2.52	5.5	3.12	5.95	2.67
MW-2	4.42	3.00	4.77	2.65	2.9	4.92	4.35	3.07
MW-3	5.23	1.19	5.37	1.05	3.6	2.82	5.1	1.32

TABLE 1(continue)
SUMMARY OF WATER LEVEL DATA

WELL Name	Elev. of TOC (Ft-MSL)	7/13/93	
		DTW Ft.	SWL Ft.
EW-1	8.62	6.2	2.42
MW-2	7.42	4.7	2.72
MW-3	6.42	5.35	1.07
MW-4	7.07*	5.75	1.32

* Adjusted elevation

Note: TOC top of casing
 DTW depth to water table
 SWL static water level above MSL
 MSL mean sea level

TABLE 2

GROUNDWATER MOVEMENT ANALYSIS

Date	4/25/89	11/6/89	2/20/90	5/31/90	9/7/90	12/4/90
Flow Towards	SW	S	S	S	S	S
Gradient	0.001	0.012	0.016	0.0125	0.0115	0.045
Date	4/16/91	7/3/91	10/14/91	1/9/92	7/15/92	10/19/92
Flow Towards	S	S	S	SW	S	S
Gradient	0.014	0.013	0.011	0.0238	0.013	0.0127
Date	1/11/93	4/19/93	7/7/93			
Flow Towards	S	SW	SW			
Gradient	0.011	0.013	0.013			

3.0 GROUNDWATER QUALITY

On July 7, 1993, ETS field personnel visited the facility and collected water samples from monitoring well EW1 for laboratory analysis. These groundwater samples were sent to a state-certified laboratory for analyses of halocarbons using EPA method 601, total petroleum hydrocarbons (TPH) as gasoline and gasoline constituents benzene, toluene, ethylbenzene, and total xylenes (BTEX) using EPA method 602.

From the results of the laboratory analysis (Appendix C), water sample taken from well EW1 contained some volatile organic compounds. The VOCs detected in well EW-1 from the July 7, 1993 sampling episode are presented in Table 3.

In addition groundwater monitoring well MW4 was installed downgradient of well EW1 as per Alameda County Health Care Services and Bay Area Regional Water Quality Control Board request. Water level and groundwater quality results of well MW4 are included in Table 1, 2, 3 and 4 of this report.

TABLE 3

SUMMARY OF QUARTERLY GROUNDWATER QUALITY RESULTS OF WELL EW-1
5800 CHRISTIE AVENUE,
EMERYVILLE, CALIFORNIA

CONCENTRATIONS IN MG/L

COMPOUNDS	5/8/89	11/6/89	2/20/90	5/31/90	9/7/90	12/4/90	4/6/91	7/3/91	10/12/92	1/8/92	4/8/92
TPH as GASOLINE	NA	0.74	12.0	24.0	25.0	7.4	51.0	23.0	39.0	<5.0	12.0
BENZENE	ND	0.18	1.3	0.056	1.1	0.18	3.0	0.65	ND	ND	4.0
TOLUENE	0.19	0.039	3.6	6.1	0.8	3.2	12.0	8.7	1.3	0.58	ND
XYLENES	0.17	0.067	0.047	0.14	0.042	ND	ND	ND	ND	ND	ND
ETHYLBENZENE	ND	0.0008	0.0071	0.017	ND	ND	ND	ND	ND	ND	ND
HALOCARBONS	0.718	1.1861	4.701	6.876	6.661	3.762	10.6	6.49	2.794	4.459	6.8
TCE	0.64	0.74	1.1	0.83	0.49	1.5	1.3	0.13	0.73	1.7	2.8
1,1 DCE	0.078	0.0023	0.014	0.069	0.036	ND	ND	ND	ND	ND	ND
1,2 DCE	ND	0.35	2.5	0.11	2.4	1.5	3.7	2.0	0.62	1.52	ND
1,1,1 TCA	ND	0.026	0.55	1.2	0.51	0.072	2.9	0.2	0.47	0.089	ND
1,1 DCA	ND	0.034	0.46	1.9	1.3	0.46	1.8	2.0	0.63	0.42	1.3
1,2 DCA	ND	0.0048	0.034	0.033	0.053	ND	ND	ND	0.12	0.25	2.7
VINYL CHLORIDE	ND	0.029	ND	2.6	1.7	0.23	0.9	1.99	0.17	0.48	ND
CHLOROETHANE	ND	ND	0.029	0.094	0.15	ND	ND	0.17	0.054	ND	ND
MET. CHLORIDE	ND	ND	0.014	0.04	0.022	ND	ND	ND	ND	ND	ND
TOTAL VOCs	1.078	1.9261	16.701	30.876	31.661	11.162	61.6	29.49	41.794	<9.459	18.8

NA NOT ANALYSED

ND NOT DETECTED OR BELOW DETECTION LIMITS

VOCs VOLATILE ORGANIC COMPOUNDS (TPH PLUS TOX)

TABLE 3(CONTINUE)

SUMMARY OF QUARTERLY GROUNDWATER QUALITY RESULTS OF WELL EW-1
5800 CHRISTIE AVENUE,
EMERYVILLE, CALIFORNIA

CONCENTRATIONS IN [REDACTED]

COMPOUNDS	7/15/92	10/19/92	1/11/93	3/29/93	7/7/93
TPH as GASOLINE	100.0	26.0	20.0	15.0	[REDACTED]
BENZENE	ND	ND	ND	ND	ND
TOLUENE	4.7	12.5	7.5	12.0	[REDACTED]
XYLENES	ND	ND	0.075	ND	ND
ETHYLBENZENE	ND	ND	ND	ND	ND
HALOCARBONS	2.461	5.07	0.065	2.5	1.7
PCE	ND	ND	0.042	ND	ND
TCE	0.68	0.27	0.023	2.0	ND
1,1 DCE	ND	4.8	ND	0.5	ND
1,2 DCE	0.6	ND	ND	ND	ND
1,1,1 TCA	0.42	ND	ND	ND	ND
1,1 DCA	0.6	ND	ND	ND	1.7
1,2 DCA	0.11	ND	ND	ND	ND
VINYL CHLORIDE	0.15	ND	ND	ND	ND
CHLOROETHANE	ND	ND	ND	ND	ND
MET. CHLORIDE	ND	ND	ND	ND	ND
TOTAL VOCs	102.461	31.07	20.065	17.5	41.7

NA NOT ANALYSED

ND NOT DETECTED OR BELOW DETECTION LIMITS

VOCs VOLATILE ORGANIC COMPOUNDS (TPH PLUS TOX)

TABLE 4

SUMMARY OF QUARTERLY GROUNDWATER QUALITY RESULTS OF WELL MW-4
 5800 CHRISTIE AVENUE,
 EMERYVILLE, CALIFORNIA
 CONCENTRATIONS IN MG/L

COMPOUNDS	7/13/93
TPH as GASOLINE	<100.0*
BENZENE	0.8
TOLUENE	0.28
XYLENES	0.3
ETHYLBENZENE	0.27
HALOCARBONS	ND
PCE	ND
TCE	ND
1,1 DCE	ND
1,2 DCE	ND
1,1,1 TCA	ND
1,1 DCA	ND
1,2 DCA	ND
VINYL CHLORIDE	ND
CHLOROETHANE	ND
MET. CHLORIDE	ND
TOTAL VOCs	<100*

* TO BE CONFIRMED WITH LABORATORY
 NA NOT ANALYSED
 ND NOT DETECTED OR BELOW DETECTION LIMITS
 VOCs VOLATILE ORGANIC COMPOUNDS (TPH PLUS TOX)

4.0 SUMMARY OF FINDINGS

Table 3 presents a summary of analytical results of well EW1 in time series. Table 4 presents the groundwater quality of well MW4. There are several factors that affect the changes in the hydrocarbon concentration. These factors are variations in water table, chemical breakdown due to biodegradation, and unidentified off-site sources.

It is obvious that there is no sign of downgradient or off site migration of the chlorinated solvents as indicated by NDs in Table 4.

MW4 detected elevated levels of BTEX compounds in the initial sample and analysis. The suspected sources of the BTEX compounds may have been originated from upgradient closed underground storage tank or from upgradient asphalt manufacturing plant. Well MW4 is located very close to underground utility lines along Powell Street which may serve as migration conduits from upgradient sources. During the construction of well MW4, asphaltic material of unknown origin (may be from upgradient asphalt manufacturing plant) was detected between 2 to 6 feet below grade.

However, there is still strong microbial activity in the subsurface as indicated in the microbiology reports on EW1 and MW4 prepared by Microbe Inotech Laboratories (Appendix D).

APPENDIX A

**ALAMEDA COUNTY HEALTH CARE SERVICES
MARCH 15, 1993 CORRESPONDENCE**

ALAMEDA COUNTY
HEALTH CARE SERVICES

AGENCY
DAVID J. KEARS, Agency Director



RAFAT A. SHAHID, Assistant Agency Director

DEPARTMENT OF ENVIRONMENTAL HEALTH
Hazardous Materials Division
80 Swan Way, Rm. 200
Oakland, CA 94621
(510) 271-4320

March 15, 1993

Crowley & Herring Investment Co.
448 Tharp Drive
Moraga, CA 94556

Subject: 5800 Christie, Emeryville, CA 94608

Dear Mr. Herring:

This letter is in response to your request for this office to concur with the discontinuation of the methane gas monitoring in the "Good Guys" electronics store located at the above site. In light of the fact that since the installation of the Sierra Monitor Model 5000 Methane Gas Detection System in 1989, that the system has never indicated any significant levels of methane, this office concurs with your contention that the monitoring may be discontinued.

Please be advised that such monitoring is strictly voluntary, and has never be regulated by this office. The monitoring has taken place at the request of your lessee, the "Good Guys". If you have any questions concerning any possible health risks at the site I suggest you contact an industrial hygienist.

If, however, there is any change in the conditions noted at the site, please contact this office immediately. The number is (510) 271-4320.

Sincerely,

A handwritten signature in cursive script that reads "Brian P. Oliva".

Brian P. Oliva, REHS, REA
Hazardous Materials Specialist

cc: Frank Verni, % The Good Guys, 5800 Christie, Emeryville, CA
Ed Howell/files

Handwritten initials, possibly "EJ", written in cursive.

TELSTAR

1717 Solano Way • Unit 34 • Concord, California 94520 • (510) 671-2888 • FAX No. (510) 671-9507

February 10, 1993

The Good Guys
5800 Christie Ave.
Emeryville, CA 94608

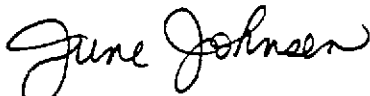
Attn: Frank Verni
Subj: Gas Detection Calibrations
Ref: SR 5159

Dear Frank:

As you know, Telstar has been calibrating your Sierra Monitor Model 5000 Methane Gas Detection system at the Emeryville facility since November, 1989. We have experienced no problems with the system, other than the failure of the power supply that was repaired in November of 1991. We were never called out for any alarms on the system, and basically during our calibrations found no problems, other than the power supply.

If you have any questions, please contact me at 510-671-2888.

Sincerely,



for Dan Mensing
Regional Manager

DM/jj

cc: Croley & Herring Inv. Co. % Dick Herring
Alameda County Health Dept. % Brian Oliva

APPENDIX B




BORING AND WELL LOG MW4

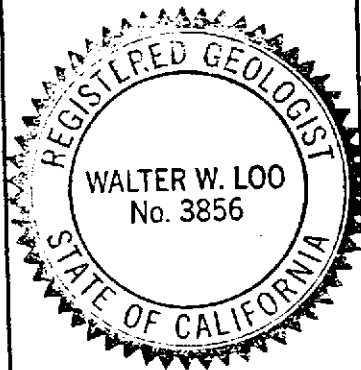
SOIL BORING & WELL LOG

SB/MW #: MW4

Page 1 of 1

PROJECT CHIC LOCATION: 5800 CHRISTIE STREET, EMERYVILLE, CA.
 ELEVATION +7.0 MSL est. MONITORING DEVICE WELL MW4
 DRILLING DATE(S) JULY 10, 1993
 DRILLING METHOD HOLLOW STEM AUGER SUBCONTRACTOR GREGG DRILLING

Depth Below Surface (ft.)	Penetration		Sampler Depth Interval (ft.)	Sample ID #	Hnu reading (ppm)	Soil Description Color, Texture, Moisture, Etc.	Unified Classification	Graphic Log	Sampled Depth	Borehole Well Construction Details
	Blows 6"-6"-6"	BPF								
0						Fill and backfill soil.	SM			10 feet PVC blank casing 2-inch dia.
5						Asphaltic material	OL			
5						Water level at 5 feet.				
10						Bay Mud				10 feet PVC slotted screen 2-inch dia. 0.010" slots
10						Gray to dark, soft silty and clayey	OH			
15										
20										Total Depth 20 Feet
										12/16 sand pack
										1 foot bentonite pellets at 1 foot above well screen to prevent migration of asphaltic material



[Signature]
SIGNATURE OF FIELD SUPERVISOR

ENVIRONMENT & TECHNOLOGY SERVICES

APPENDIX C

GROUNDWATER LABORATORY ANALYSIS REPORT



mail to ETS.
by Dick Hennig.
on 7/22/93

CKY incorporated Environmental Services

Date: 07/15/93
N9307-28

CHIC
448 Tharp Drive
Moraga, CA 94556

Attn: Mr. Walter Loo

Subject: Laboratory Report
Project:

~~CONFIDENTIAL~~

Enclosed is the laboratory report for samples received on 07/08/93. The samples were received in coolers with ice and intact; the chain-of-custody forms were properly filled out. The data reported includes:

<u>Method</u>	<u>No. of Analysis</u>
EPA 8010	1 Water
M8015 (Gas)	1 Water
EPA 8020	1 Water

The results are summarized on the following pages.

Please feel free to call if you have any questions concerning these results.

Sincerely,

Danny Hoang
Laboratory Director

EPA METHODS - 601

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=====
CLIENT:      CHIC
PROJECT:
SAMPLE ID:   Blank
CONTROL NO:  N93007-28
DATE REC'D:  07/08/93
DATE ANALYZED: 07/14/93
MATRIX TYPE:  Water
=====

```

<u>PARAMETERS (601)</u>	<u>RESULTS</u> <u>(ug/L)</u>	<u>DETECTION LIMIT</u> <u>(ug/L)</u>
Dichlorodifluoromethane	ND	5
Chloromethane	ND	5
Vinyl Chloride	ND	5
Bromomethane	ND	5
Chloroethane	ND	1
Trichlorofluoromethane	ND	1
1,1-Dichloroethene	ND	1
Methylene Chloride	ND	1
Trans-1,2-Dichloroethene	ND	1
cis 1,2 -dichloroethene	ND	1
1,1-Dichloroethane	ND	1
Chloroform	ND	1
1,1,1-Trichloroethane	ND	1
Carbon Tetrachloride	ND	1
1,2-Dichloroethane	ND	1
Trichloroethene	ND	1
1,2-Dichloropropane	ND	1
Bromodichloromethane	ND	1
2-Chloroethylvinylether	ND	1
Trans-1,3-Dichloropropene	ND	1
Cis-1,3-Dichloropropene	ND	1
1,1,2-Trichloroethane	ND	1
Tetrachloroethene	ND	1
Dibromochloromethane	ND	1
Chlorobenzene	ND	1
Bromoform	ND	1
1,1,2,2-Tetrachloroethane	ND	1
M-Dichlorobenzene	ND	1
P-Dichlorobenzene	ND	1
O-Dichlorobenzene	ND	1

EPA METHODS - 601

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=====
CLIENT:      CHIC
PROJECT:
SAMPLE ID:   EW-1
CONTROL NO:  N9307-28-1
DATE REC'D:  07/08/93
DATE ANALYZED: 07/14/93
MATRIX TYPE:  Water
=====
  
```

<u>PARAMETERS (601)</u>	<u>RESULTS (ug/L)</u>	<u>DETECTION LIMIT (ug/L)</u>
Dichlorodifluoromethane	ND	5
Chloromethane	ND	5
Vinyl Chloride	ND	5
Bromomethane	ND	5
Chloroethane	ND	1
Trichlorofluoromethane	ND	1
1,1-Dichloroethene	ND	1
Methylene Chloride	ND	1
Trans-1,2-Dichloroethene	ND	1
cis 1,2 -dichloroethene	ND	1
1,1-Dichloroethane	ND	1
Chloroform	ND	1
1,1,1-Trichloroethane	ND	1
Carbon Tetrachloride	ND	1
1,2-Dichloroethane	ND	1
Trichloroethene	ND	1
1,2-Dichloropropane	ND	1
Bromodichloromethane	ND	1
2-Chloroethylvinylether	ND	1
Trans-1,3-Dichloropropene	ND	1
Cis-1,3-Dichloropropene	ND	1
1,1,2-Trichloroethane	ND	1
Tetrachloroethene	ND	1
Dibromochloromethane	ND	1
Chlorobenzene	ND	1
Bromoform	ND	1
1,1,2,2-Tetrachloroethane	ND	1
M-Dichlorobenzene	ND	1
P-Dichlorobenzene	ND	1
O-Dichlorobenzene	ND	1

=====

DILUTION FACATOR = 100

EPA METHOD 5030/Mod. 8015

TOTAL PETROLEUM HYDROCARBONS BY PURGE & TRAP

=====

CLIENT:	CHIC	DATE REC'D:	07/08/93
PROJECT:		DATE ANALYZED:	07/12/93
CONTROL NO:	N9307-28	MATRIX:	Water

=====

<u>SAMPLE ID:</u>	<u>CONTROL NO:</u>	<u>RESULTS</u> <u>(mg/L)</u>	<u>DET. LIMIT</u> <u>(mg/L)</u>	<u>% SURRO</u> <u>RECOVERY</u>
Blank	N9307-28	ND	1.0	114
	N9307-28-1		1.0	91

=====



QUALITY CONTROL DATA

CLIENT: CHIC
 PROJECT:
 CONTROL NO: N9307-28 DATE ANALYZED: 07/12/93

METHOD EPA 8010
 MATRIX: Water

SAMPLE ID: Blank

<u>COMPOUND</u>	<u>SAMPLE RESULTS</u> (mg/L)	<u>AMOUNT SPIKED</u> (mg/L)	<u>% REC.</u>	<u>DUP. % REC.</u>	<u>RPD</u>
1,1-DCE	ND	50	84	92	9
TCE	ND	50	86	96	11
Chlorobenzene	ND	50	114	102	11



QUALITY CONTROL DATA

CLIENT: CHIC
PROJECT:
CONTROL NO: N9307-28 DATE ANALYZED: 07/13/93

METHOD EPA M8015G
MATRIX: Water

SAMPLE ID: Blank

<u>COMPOUND</u>	<u>SAMPLE RESULTS</u> (mg/L)	<u>AMOUNT SPIKED</u> (mg/L)	<u>% REC.</u>	<u>DUP. % REC.</u>	<u>RPD</u>
Gas	ND	2.0	106	120	12



QUALITY CONTROL DATA

CLIENT: CHIC
 PROJECT:
 CONTROL NO: N9307-28

DATE ANALYZED: 07/13/93

METHOD: EPA 8020
 MATRIX: Water

SAMPLE ID: Blank

<u>COMPOUND</u>	<u>SAMPLE RESULTS</u> (ug/L)	<u>AMOUNT SPIKED</u> (ug/L)	<u>% REC.</u>	<u>DUP. % REC.</u>	<u>RPD</u>
Benzene	ND	10	110	130	17
Toluene	ND	10	110	120	9
Ethyl Benzene	ND	10	100	110	10
Xylene	ND	20	100	115	14





CKY incorporated Environmental Services

Date: 07/20/93
N9307-100

CHIC
448 Tharp Drive
Moraga, CA 94556

Attn: Mr. Walter Loo

Subject: Laboratory Report
Project:

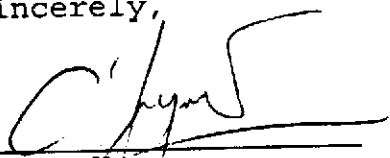
Enclosed is the laboratory report for samples received on 07/13/93. The samples were received in coolers with ice and intact; the chain-of-custody forms were properly filled out. The data reported includes:

<u>Method</u>	<u>No. of Analysis</u>
EPA 8020	1 Water
M8015 (Gas)	1 Water
EPA 8010	1 Water

The results are summarized on the following pages.

Please feel free to call if you have any questions concerning these results.

Sincerely,



Danny Hoang
Laboratory Director

EPA METHOD - 8020
BTEX

=====

CLIENT:	CHIC	DATE REC'D:	07/13/93
PROJECT:		DATE ANALYZED:	07/16/93
CONTROL NO:	N9307-100	MATRIX TYPE:	Water

=====

SAMPLE ID:	CONTROL NO:	RESULTS (ug/L)				% SURRO RECOVERY
		Benz	Tol	Et Benz	Xyls	
Blank	N9307-100	ND	ND	ND	ND	84
MW4	*N9307-100-1	800	280	270	200	
DETECTION LIMIT		1	1	1	1	

* DETECTION LIMIT FOR ALL COMPOUNDS - 100 ug/L

=====

EPA METHOD 5030/Mod. 8015
 TOTAL PETROLEUM HYDROCARBONS BY PURGE & TRAP

```

=====
CLIENT:      CHIC                      DATE REC'D:   07/13/93
PROJECT:                                           DATE ANALYZED: 07/16/93
CONTROL NO:  N9307-100                       MATRIX:      Water
=====
    
```

<u>SAMPLE ID:</u>	<u>CONTROL NO:</u>	<u>RESULTS</u> <u>(mg/L)</u>	<u>DET. LIMIT</u> <u>(mg/L)</u>	<u>% SURRO</u> <u>RECOVERY</u>
Blank	N9307-100	ND	1.0	84
MW4*	N9307-100-1	ND*	100*	72

* Sample could not be run at a lower detection limit due to substantial interference. There is however a presence of compounds not identifiable as gasoline below the 100 mg/L detection limit.

EPA METHODS - 601

```

=====
CLIENT:      CHIC                      DATE REC'D:    07/13/93
PROJECT:
SAMPLE ID:   MW4                      DATE ANALYZED: 07/16/93
CONTROL NO:  N9307-100-1             MATRIX TYPE:   Water
=====
    
```

<u>PARAMETERS (601)</u>	<u>RESULTS (ug/L)</u>	<u>DETECTION LIMIT (ug/L)</u>
Dichlorodifluoromethane	ND	500
Chloromethane	ND	500
Vinyl Chloride	ND	500
Bromomethane	ND	500
Chloroethane	ND	500
Trichlorofluoromethane	ND	100
1,1-Dichloroethene	ND	100
Methylene Chloride	ND	100
Trans-1,2-Dichloroethene	ND	100
cis 1,2 -dichloroethene	ND	100
1,1-Dichloroethane	ND	100
Chloroform	ND	100
1,1,1-Trichloroethane	ND	100
Carbon Tetrachloride	ND	100
1,2-Dichloroethane	ND	100
Trichloroethene	ND	100
1,2-Dichloropropane	ND	100
Bromodichloromethane	ND	100
2-Chloroethylvinylether	ND	100
Trans-1,3-Dichloropropene	ND	100
Cis-1,3-Dichloropropene	ND	100
1,1,2-Trichloroethane	ND	100
Tetrachloroethene	ND	100
Dibromochloromethane	ND	100
Chlorobenzene	ND	100
Bromoform	ND	100
1,1,2,2-Tetrachloroethane	ND	100
M-Dichlorobenzene	ND	100
O-Dichlorobenzene	ND	100
O-Dichlorobenzene	ND	100

* HIGHER DET. LIMIT DUE TO SAMPLE MATRIX.

EPA METHODS - 601

=====

CLIENT: CHIC
 PROJECT:
 SAMPLE ID: Blank
 CONTROL NO: N9307-100

=====

DATE REC'D: 07/13/93
 DATE ANALYZED: 07/16/93
 MATRIX TYPE: Water

=====

<u>PARAMETERS (601)</u>	<u>RESULTS (ug/L)</u>	<u>DETECTION LIMIT (ug/L)</u>
Dichlorodifluoromethane	ND	5
Chloromethane	ND	5
Vinyl Chloride	ND	5
Bromomethane	ND	5
Chloroethane	ND	5
Trichlorofluoromethane	ND	1
1,1-Dichloroethene	ND	1
Methylene Chloride	ND	1
Trans-1,2-Dichloroethene	ND	1
cis 1,2 -dichloroethene	ND	1
1,1-Dichloroethane	ND	1
Chloroform	ND	1
1,1,1-Trichloroethane	ND	1
Carbon Tetrachloride	ND	1
1,2-Dichloroethane	ND	1
Trichloroethene	ND	1
1,2-Dichloropropane	ND	1
Bromodichloromethane	ND	1
2-Chloroethylvinylether	ND	1
Trans-1,3-Dichloropropene	ND	1
Cis-1,3-Dichloropropene	ND	1
1,1,2-Trichloroethane	ND	1
Tetrachloroethene	ND	1
Dibromochloromethane	ND	1
Chlorobenzene	ND	1
Bromoform	ND	1
1,1,2,2-Tetrachloroethane	ND	1
M-Dichlorobenzene	ND	1
P-Dichlorobenzene	ND	1
O-Dichlorobenzene	ND	1

=====

QUALITY CONTROL DATA

CLIENT: CHIC
 PROJECT:
 CONTROL NO: N9307-100

DATE EXTCD: N/A
 DATE ANALYZED: 07/16/93

METHOD EPA 8020
 MATRIX: Water

SAMPLE ID: N9307-127-5

<u>COMPOUND</u>	<u>SAMPLE RESULTS</u> (ug/L)	<u>AMOUNT SPIKED</u> (ug/L)	<u>% REC.</u>	<u>DUP. % REC.</u>	<u>RPD</u>
Benzene	ND	10	70	80	13
Toluene	ND	10	80	90	12
Ethyl Benzene	ND	10	70	80	13
Xylene	ND	20	80	90	12

QUALITY CONTROL DATA

CLIENT: CHIC
 PROJECT: DATE EXTRACTED: N/A
 CONTROL NO: N9307-100 DATE ANALYZED: 07/16/93

METHOD EPA M8015G
 MATRIX: Water

SAMPLE ID: n9307-127-5

<u>COMPOUND</u>	<u>SAMPLE RESULTS</u> (mg/L)	<u>AMOUNT SPIKED</u> (mg/L)	<u>% REC.</u>	<u>DUP. % REC.</u>	<u>RPD</u>
Gas	ND	1.0	100	100	0

QUALITY CONTROL DATA

CLIENT: CHIC
 PROJECT: DATE EXTRACTED: N/A
 CONTROL NO: N9307-100 DATE ANALYZED: 07/16/93

METHOD EPA 8010
 MATRIX: Water

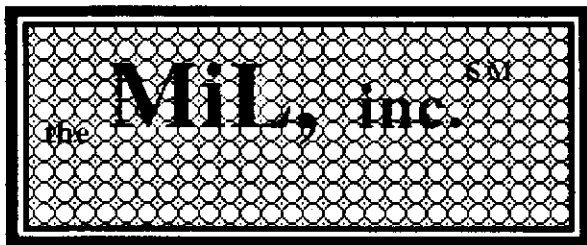
SAMPLE ID: N9307-128-4

<u>COMPOUND</u>	<u>SAMPLE RESULTS</u> (mg/L)	<u>AMOUNT SPIKED</u> (mg/L)	<u>% REC.</u>	<u>DUP. % REC.</u>	<u>RPD</u>
TCE	ND	20	100	115	14
Chlorobenzene	ND	20	100	85	16



APPENDIX D

MICROBIOLOGY ANALYSIS



**Total
Heterotrophic
Plate Count
Analysis &
Microbial
Identification**

**Microbe
Inotech
Laboratories,
inc.**

1840 Craig Road
St. Louis, MO
63146-4712
U.S.A.

Telephone: (314) 878-6626
(800) 688-9144

FAX: (314) 878-9376

E-mail: Bruce C. Hemming
76177.204@compuserve.com

Report Prepared For:

**Environmental & Technical
Services**

**ATTN: Walter Loo, R.G.,
C.E.G.**

**2081 15th Street
San Francisco, CA 94114**

Client Phone (415) 861-0810

Client Fax (415) 861-3269

Report No. MILB—2305

PO Number none

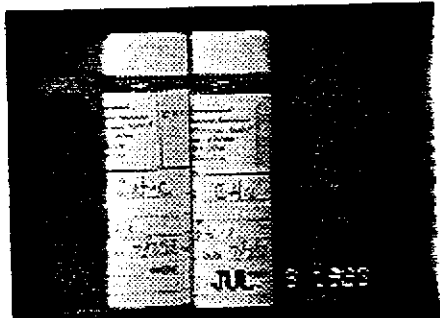
July 19, 1993

Summary Report of Analysis
[No. 2305]

Environmental & Technical Services
ATTN: Walter Loo, R.G., C.E.G.
2081 15th Street
San Francisco, CA 94114

Description:

Fri, July 9, 1993 2:30 p.m. : Received by 2nd day Fedex one water sample labeled EW-1 in two 40 ml vials. Analyses requested are Total Plate Count and Bacterial ID. MiL #2305 jk. pict file. Strains may be tested for co-metabolic study—to be confirmed later.



Chain of Custody Record Information

Purchase Order No.—none
MiL, Inc. REPORT & Invoice No.: MILB-2305

Processing:

[Standard Bacterial Plate Count - serial dilution method and direct spread plate count] Within 20 minutes of reception aliquots from the sample was checked for volume and then serially diluted. Each dilution was sterilely transferred in a laminar flow biological cabinet and placed on previously prepared and dried trypticase soy broth agar (TSBA) medium in Petri plates. Observations for colony forming units (CFU) were made at 24 and 48 hrs. of incubation at 28°C for the sample. Colony differentiation was noted at 48 hrs.

Summary Final Results—Total Heterotrophic Plate Count:

DATA:	Direct Count: Colony Forming Units (CFU/ g) on TSBA Medium	
	24 Hrs.	48 Hrs.
Sample: EW-1	4.59 x 10 ⁵	4.64 x 10 ⁵
<u>Distinct morphological Colony Types at 48 Hrs. in Sample</u>		
3		

Summary of GC-FAME/Biolog Analyses							
Strain Name	Primary Identification by GC	Sim. Coef.	Dist. Coef.	Primary ID by Biolog™	Plate Type	Sim. Coef	Dist. Coef
2305-1	<i>Staphylococcus xylosus</i>	0.013	9.561	<i>Staphylococcus hominis</i>	GP	0.994	0.51
2305-2	<i>Bacillus maroccanus</i>	0.287	5.104	CDC Group D-2	GP	0.286	9.994
2305-3	<i>Comamonas testosteroni</i>	0.13	6.533	<i>Alcaligenes faecalis type II</i>	GN	0.769	2.432

GC-FAME & Biolog™ Processing:

Following isolation the strains were individually streaked out onto TSBA. The TSBA plates were processed after 24 hr incubation by [Method 1 - Standard GC-FAME]. The strains were examined against both the newly installed Aerobe (TSBA [rev. 3.70]) and Clinical Aerobe (CLIN [rev.3.70]) GC-FAME databases. Subsequently the strains were prepared for Biolog™ analysis by suspending them in sterile saline and loading the solutions into the appropriate microtiter plates (Gram negative or Gram positive). The plates were incubated for 24 hours and then examined against version 3.0 of the Biolog™ database using an automated microplate reader.

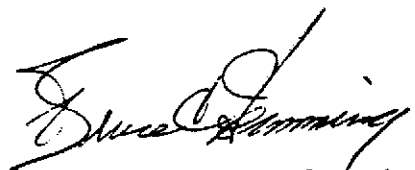
Similarity and Distance Coefficient

In order to create the database that we use to identify your organisms, thousands of species of bacteria had to be tested. In fact each species itself had to be tested hundreds of times to determine a set of characteristics unique to it. The species characteristics that are in our database are an "average" of the characteristics of hundres of tested bacteria of the same species. The Similarity and Distance Coefficient of your organism refers to the similarity and distance to the hypothetical 'mean' organism in the database. The database organism has a similarity coefficient of one and a distance of zero. So the closer your strain is to one and zero the more closely it matches the mean organism in the database.

A good match is one with a similarity coefficient greater than .5 and a distance coefficient of less than 7.

Thank you from the staff on project.

Julie Milke - Laboratory Manager



Dr. Bruce C. Hemming - Operations Director

SECRET

CALIB. 1 CALIBRATION STANDARD (AEROBE) 15-JUL-93 00:31:25 Area: 644208 % Named: 90
** GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0022. **

CALIB. 1 CALIBRATION STANDARD (AEROBE) 15-JUL-93 01:01:44 Area: 541700 % Named: 90
** GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0012. **

2 2305-1 L00 (AEROBE) 15-JUL-93 02:02:25 Area: 102908 % Named: 100
TSBA [Rev 3.70] Staphylococcus 0.013
S. xylosus 0.013
CLIN [Rev 3.70] * NO MATCH *

3 2305-2 L00 (AEROBE) 15-JUL-93 02:32:46 Area: 1048 % Named: 100
TOTAL AREA LESS THAN 50000.
TSBA [Rev 3.70] Bacillus 0.287 (not an approved name)
B. maroccanus 0.287 (not an approved name)
ELIN [Rev 3.70] * NO MATCH *

16-JUL-93 20:05:51

CALIB. 1 CALIBRATION STANDARD (AEROBE) 16-JUL-93 01:38:29 Area: 475000 % Named: 100
** GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0019. **

CALIB. 1 CALIBRATION STANDARD (AEROBE) 17-JUL-93 02:08:52 Area: 484904 % Named: 100
** GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0013. **

1 2305-3 (AEROBE) 17-JUL-93 02:39:07 Area: 108840 % Named: 90
TSBA [Rev 3.70] Comamonas 0.068 (Pseudomonas testosteroni)
C. testosteroni 0.068 (Pseudomonas testosteroni)
Variovorax 0.050 (Ralcaligenes paradoxus)
U. paradoxus 0.050 (Ralcaligenes paradoxus)
U. p. GC subgroup B 0.050 (Ralcaligenes paradoxus)
U. p. GC subgroup A 0.028 (Ralcaligenes paradoxus)
Pseudomonas 0.046
P. putida 0.046
P. p. biotype B 0.046
P. syringae 0.035
P. s. syringae 0.035
P. s. savastanoi pv. fraxinus 0.028
P. s. phaseolicola 0.019
P. corrugata 0.033
CLIN [Rev 3.70] Comamonas 0.130 (Pseudomonas testosteroni)
C. testosteroni 0.130 (Pseudomonas testosteroni)

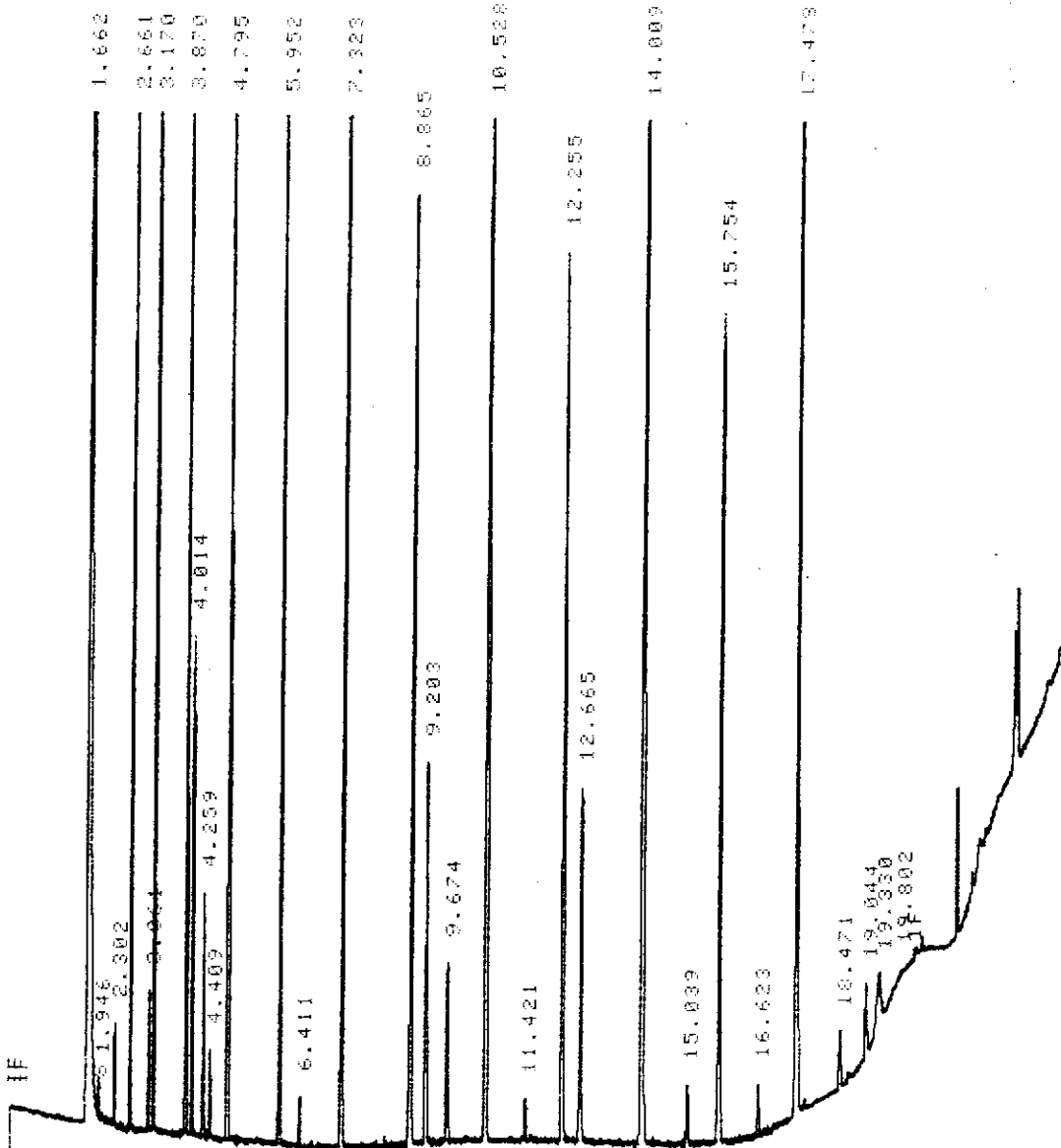
ID: 1 CALIBRATION STANDARD Date of run: 15-JUL-93 01:01:44
 Bottle: 1 CALIBRATION (ACROBE)

RT	Area	Ar/Ht Respon	ECL	Name	%	Comment 1	Comment 2
1.674	135145472	0.023	7.041	SOLVENT PEAK		< min rt	
1.956	504	0.020	7.595			< min rt	
2.312	1536	0.022	8.295				
2.671	27664	0.024	9.000	9:0	5.13		
3.071	2496	0.024	9.786				
3.130	56688	0.026	10.000	10:0	10.08	Peak match -0.0004	
3.880	29528	0.029	11.000	11:0	5.05	Peak match -0.0007	
4.024	11688	0.030	11.156	10:0 20H	2.03	Peak match 0.0010	
4.269	6056	0.031	11.421	10:0 30H	1.02	Peak match 0.0019	
4.419	2048	0.030	11.583				
4.805	61904	0.032	12.000	12:0	10.27	Peak match -0.0010	
5.962	31552	0.036	13.000	13:0	5.09	Peak match -0.0001	
6.420	1472	0.035	13.334				
7.333	63864	0.039	14.000	14:0	10.06	Peak match 0.0004	
8.875	32904	0.044	15.000	15:0	5.24	Peak match -0.0016	
9.212	13864	0.044	15.203	14:0 20H	2.13	Peak match 0.0023	
9.685	6944	0.045	15.487	Sum In Feature 3	1.06	Peak match 0.0015	14:0 30H/16:1 ISO I
10.537	66720	0.044	16.000	16:0	10.14	Peak match -0.0010	
11.433	1384	0.041	16.519				
12.265	33608	0.046	17.000	17:0	5.04	Peak match -0.0003	
12.675	14240	0.048	17.234	16:0 20H	2.13	Peak match 0.0012	
14.019	68624	0.048	18.000	18:0	10.20	Peak match -0.0005	
15.050	2576	0.051	18.590				
15.765	34280	0.049	19.000	19:0	5.05	Peak match 0.0003	
16.631	2032	0.047	19.502				
17.489	69944	0.050	20.000	20:0	10.25		
18.480	2400	0.050	20.575				
19.052	10560	0.118	20.907			> max ar/ht	
19.342	21560	0.216	21.075			> max ar/ht	
19.814	31168	0.312	21.349			> max ar/ht	
*****	6944			SUMMED FEATURE 3	1.06	12:0 ALDE ?	unknown 10.928
*****						16:1 ISO I/14:0 30H	14:0 30H/16:1 ISO I

Solvent Ar Total Area Named Area % Named Total Amt Nbr Ref ECL Deviation Ref ECL Shift
 135145472 643272 631264 98.13 630859 0
 GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0012.

CALIBRATION STANDARD

RUN # 2 JUL 15, 1993 01:01:44
START



STOP

RUN # 2 JUL 15, 1993 01:01:44
START-NO PLOT
END OF SIGNAL

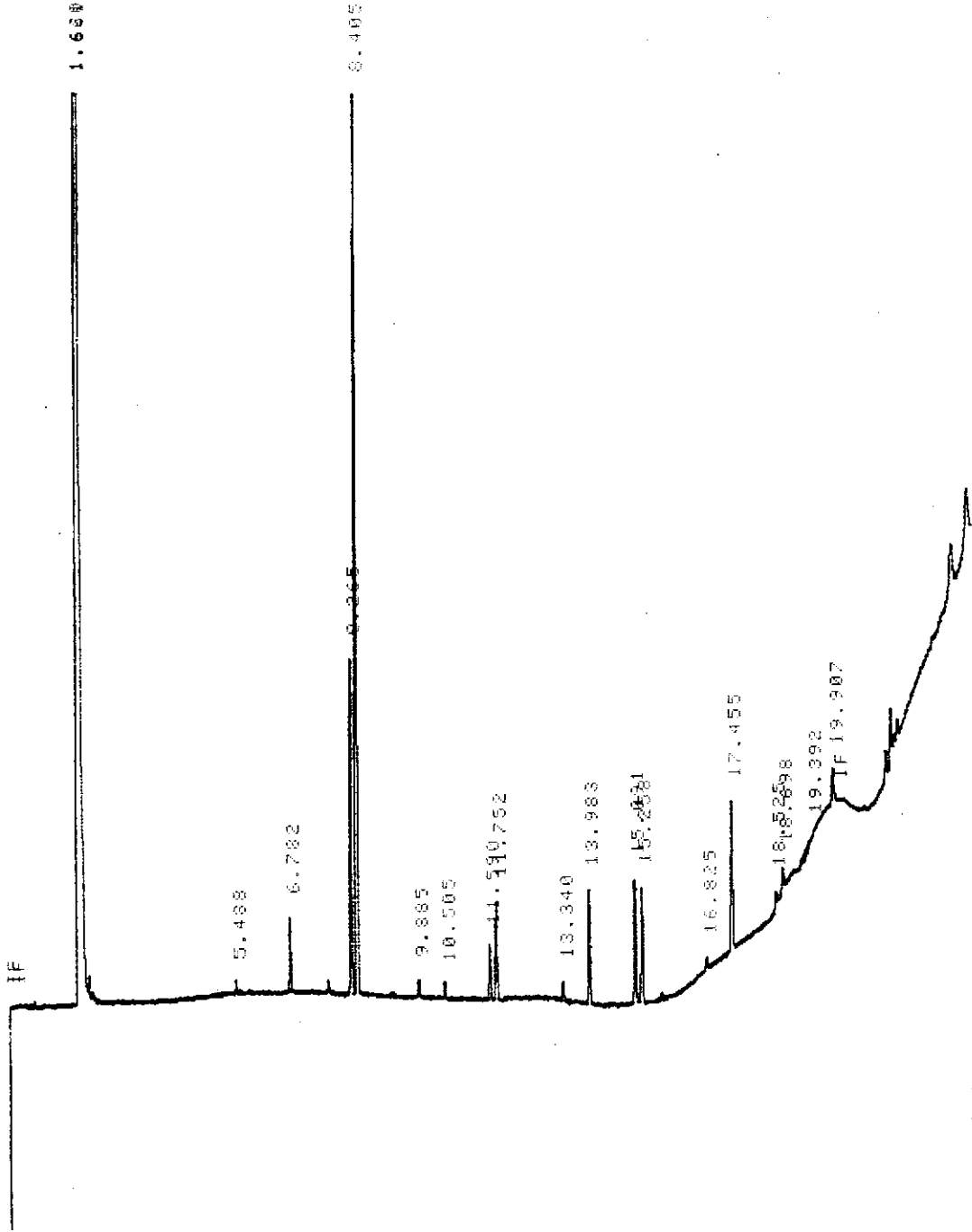
#

BOTTLE: 3 ID#: 2THU 15-JUL-93 02:49:15

FILE DATA:F93715046

2305-1 L00

RUN # 4 JUL 15, 1993 02:02:25
START



STOP

RUN # 4 JUL 15, 1993 02:02:25
START-No plot
END OF SIGNAL

#

FILE DATA 7793710006

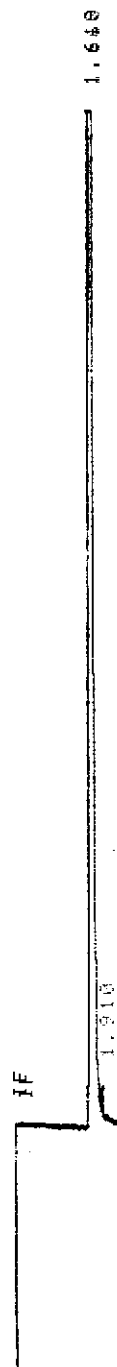
3THU 15 JUL 1993 02:19:51

BOTTLE: 400

2305-2 L00

RUN # 5 JUL 15, 1993 02:32:46

START



1.910

8.400

19.365
IF 19.909

STOP

RUN # 5 JUL 15, 1993 02:32:46

START-NO #101

END OF SIGNAL

BREAK

*

ID: 1 CALIBRATION STANDARD Date of run: 16-JUL-93 01:38:39

Bottle: 1 CALIBRATION LAEROBEC

RT	Area	Ar/Ht	Respon	ECL	Name	Z	Comment 1	Comment 2
1.671	133543744	0.023	...	7.033	SOLVENT PEAK	...	< min rt	
2.667	20720	0.024	1.171	9.000	9:0	5.11		
3.175	42680	0.026	1.121	10.000	10:0	10.07	Peak match -0.0013	
3.876	22360	0.029	1.079	11.000	11:0	5.08	Peak match -0.0031	
4.019	9112	0.030	1.073	11.155	10:0 20H	2.06	Peak match 0.0026	
4.264	4672	0.032	1.063	11.420	10:0 30H	1.05	Peak match 0.0010	
4.900	46800	0.032	1.044	12.000	12:0	10.29	Peak match -0.0030	
5.956	23880	0.036	1.015	13.000	13:0	5.11	Peak match -0.0016	
7.326	48376	0.039	0.992	14.000	14:0	10.11	Peak match -0.0021	
8.868	24680	0.042	0.974	15.000	15:0	5.06	Peak match -0.0018	
9.206	10496	0.044	0.971	15.203	14:0 20H	2.15	Peak match 0.0016	
9.679	5224	0.045	0.966	15.488	Sum In Feature 3	1.06	Peak match 0.0015	14:0 30H/16:1 ISO 1
10.531	50408	0.044	0.960	16.000	16:0	10.19	Peak match -0.0023	
12.260	25480	0.046	0.949	17.000	17:0	5.09	Peak match -0.0013	
12.670	10776	0.049	0.947	17.234	16:0 20H	2.15	Peak match 0.0014	
14.015	51392	0.048	0.940	18.000	18:0	10.18	Peak match -0.0013	
15.763	25920	0.049	0.933	19.000	19:0	5.10	Peak match -0.0001	
17.486	52008	0.049	0.927	20.000	20:0	10.16		
*****	5224	SUMMED FEATURE 3	1.06	12:0 ALDE ?	unknown 10.926
*****	16:1 ISO 1/14:0 30H	14:0 30H/16:1 ISO 1

Solvent Ar	Total Area	Named Area	% Named	Total Amt	Nbr Ref	ECL	Deviation	Ref ECL	Shift
133543744	425000	425000	100.00	424805	0				

GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0019.

BOTTLE: 1 ID#: 1FRI 16-JUL-93 23:35:52

FILE: DATA:F93716849

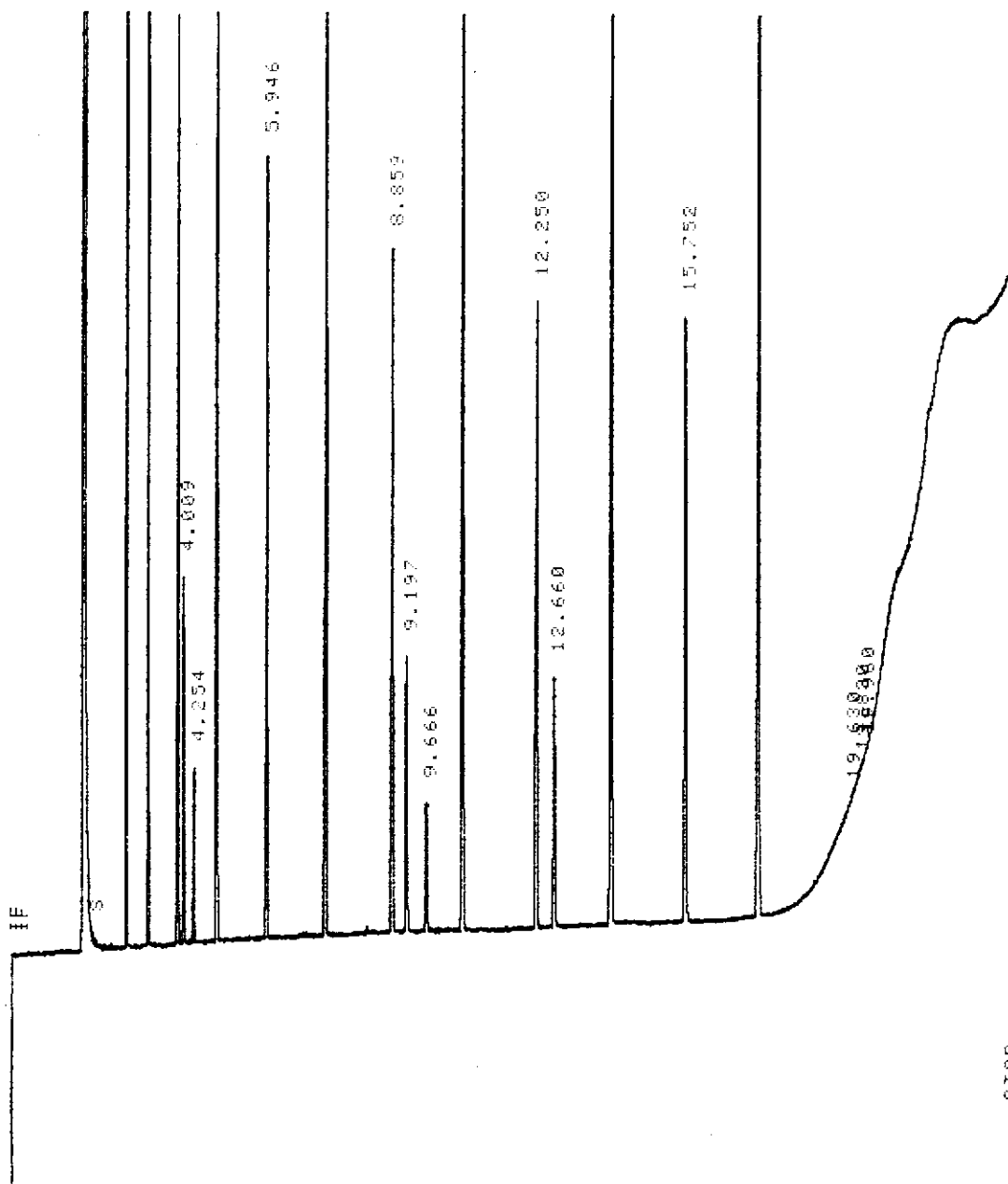
CALIBRATION STANDARD

RUN # 1 JUL 16, 1993 01:38:39

START

IF

1.660
2.652
3.165
3.866
4.790
5.946
7.316
8.859
9.197
9.666
10.521
12.250
12.660
14.005
15.792
17.475



STOP

RUN # 1 JUL 16, 1993 01:38:39

START-No Plot

END OF SIGNAL

#

ID: 1 2305-B Date of run: 17-JUL-93 02:39:07
 Bottle: 2 SAMPLE [AERROBE]

RT	Area	Area%	Response	ECL	Name	%	Comment 1	Comment 2
1.669	122711872	0.028	...	7.046	SOLVENT PEAK	...	< min rt	
1.995	1240	0.026	...	7.686		...	< min rt	
4.260	4184	0.032	1.076	11.421	10:0 30H	4.31	ECL deviates -0.002	
4.794	4128	0.032	1.055	12.000	12:0	4.17	ECL deviates 0.000	Reference -0.003
4.960	1736	0.036	...	12.145		...		
6.569	956	0.041	1.018	13.454	12:0 30H	0.93	ECL deviates -0.001	
7.316	680	0.042	1.008	14.000	14:0	0.66	ECL deviates -0.000	Reference -0.004
10.215	31448	0.045	0.979	15.818	16:1 w7c	29.50	ECL deviates 0.001	
10.517	28216	0.044	0.976	16.000	16:0	26.39	ECL deviates -0.000	Reference -0.004
12.053	7560	0.047	0.961	16.889	17:0 CYCLO	6.97	ECL deviates 0.001	Reference -0.003
12.247	840	0.055	0.959	17.001	17:0	0.77	ECL deviates 0.001	Reference -0.003
12.330	3240	0.057	0.958	17.048	16:1 20H	2.97	ECL deviates 0.001	
12.655	3664	0.047	0.955	17.233	16:0 20H	3.35	ECL deviates -0.002	
13.597	3816	0.051	0.943	17.770	18:1 w9c	3.45	ECL deviates 0.001	
13.690	16048	0.049	0.942	17.824	Sum in feature ?	14.49	ECL deviates -0.001	18:1 w9c/w12t/w7c
13.997	1320	0.056	0.937	17.999	18:0	1.19	ECL deviates -0.001	Reference -0.005
15.575	1096	0.062	0.911	18.901	19:0 CYCLO w8c	0.96	ECL deviates 0.001	Reference -0.002
19.567	28688	0.573	...	21.215		...	> max rt	
19.869	45672	0.297	...	21.390		...	> max rt	
*****	16048		SUMMED FEATURE ?	14.48	18:1 w7c/w9t/w12t	18:1 w9c/w12t/w7c
*****					18:1 w12t/w9t/w7c	

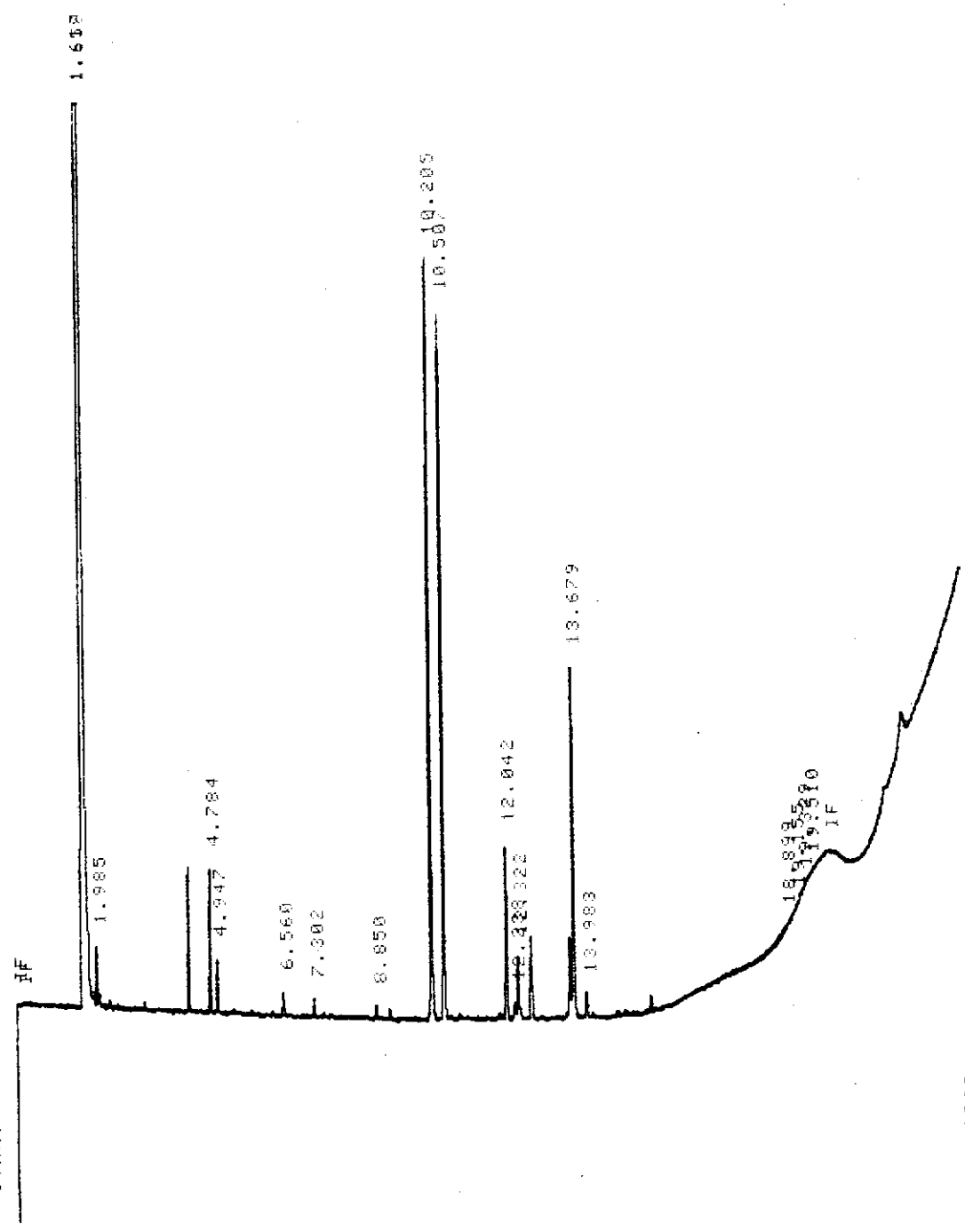
Solvent Rt	Total Area	Named Area	% Named	Total Amt	Nbr Ref	ECL Deviation	Ref ECL Shift
122711872	108940	107104	98.40	104390	7	0.001	0.004

TSBR [Rev 3.70]	Comamonas	0.068	(Pseudomonas testosteroni)
	C. testosteroni	0.068	(Pseudomonas testosteroni)
	Uariovorax	0.050	(Alcaligenes paradoxus)
	U. paradoxus	0.050	(Alcaligenes paradoxus)
	U. p. GC subgroup B	0.050	(Alcaligenes paradoxus)
	U. p. GC subgroup A	0.028	(Alcaligenes paradoxus)
	Pseudomonas	0.046	
	P. putida	0.046	
	P. p. biotype B	0.046	
	P. syringae	0.035	
	P. s. syringae	0.035	
	P. s. savastanoi pv. fraxinus	0.020	
	P. s. phaseolicola	0.019	
	P. corrugata	0.033	
CLIN [Rev 3.70]	Comamonas	0.130	(Pseudomonas testosteroni)
	C. testosteroni	0.130	(Pseudomonas testosteroni)

BOTTLE: 2 ID#: 1 SAT 17-JUL-93 00:35:06
FILE DATA:F93715849

2005-3

RUN # 3 JUL 17, 1993 02:39:07
START



STOP

RUN # 3 JUL 17, 1993 02:39:07
START-No plot
END OF SIGNAL

#

Biolog Dan
24 Hr Time Point



the **MiL, inc.**

**Interpretation of the Carbon Source
Pattern Recognition Data using a
Multi-well Plate Method (Biolog
Microplate System™) -- Contact Us:
314-878-6626 or Fax 314-878-9376**

The MiL, inc. utilizes the Biolog Microplate System™ for microbial identification and characterization by carbon source pattern recognition. The microplate technique allows for characterization by 95 different tests yielding a potential of 4×10^{28} patterns generated from a single microplate. Each strain of micro-organism yields a distinct pattern and the different species of bacteria will give distinct families of patterns that can be recognized by the Biolog MicroLog™ Software. Microplates are available for Gram Negative (GN), Gram Positive (GP) and E.coli/Salmonella (ES) Analysis. Custom analysis are performed by the MiL, inc. and can be particularly useful in biodegradation or additional selective media development studies. Additional interpretative instructions are provided with such custom services.

To characterize a given microbial isolate the organism is streaked onto a nutrient medium that will support vigorous growth (for example, Nutrient agar, tryptic soy agar or tryptic soy agar supplemented with 5% sheep red blood cells). The more fastidious organisms may require chocolate or BHI agar for growth, whereas many environmental organisms grow better in more minimal media. The culture plates are incubated at 28 to 35° C for 4-18 hours (environmental isolates are typically grown at 28° C with thermophyllic strains often incubated at 50° C). After incubation colonies are removed from the culture plate using a saline moistened cotton swab. A suspension of uniform turbidity is prepared in 0.85% saline by comparison with a

known turbidimetric standard. The bacterial suspension is inoculated into the microplate wells (150 µl per well) and the plate covered with the microplate lid. The covered plates are incubated at 28-35° C for 4 hours or overnight (16-24 hours). Should other diluents be requested or used, such changes will be noted.

Microplates may be read at 4 or 24 hours because some organisms give results at 4 hours and may become unreadable at 24 hours. The plates are read in our microplate reader at 590 nm. The absorbance or transmittance (i.e. color) in each well is referenced against the negative control well (A-1) so that any purple color recorded above this control level is read as a positive utilization of the given carbon source. The data are reported as the percent color change as compared to well A-1 utilizing the following formula.

$$\text{Percent color change} = \frac{\text{OD}_{590}(\text{well}) - \text{OD}_{590}(\text{well A-1})}{\text{OD}_{590}(\text{well A-1})}$$

Positive results will be reported in brackets ([]), generally if the Percent Color Change is equal to or greater than 40, the reaction in the given well is considered to be "positive" however the parameters for each substrate may be different and a positive test below a value of 40 is possible. The reported results will be otherwise considered negative. The computer algorithms employed provide standardization of settings ensuring repeatability and avoidance of operator bias. Names of all carbon source substrates employed are provided in the results regardless of response.

We, the MiL's microbiologists, find these methods to be excellent for strain characterization or differentiation between isolates. However, we urge caution in acceptance of the putative identifications to the commercial database and suggest these tests be conducted in conjunction with other methods (we recommend our GC-FAME analyses) when strain identifications are sought.

GN MicroPlate™

A1 water	A2 α-cyclodextrin	A3 dextrin	A4 glycogen	A5 tween 40	A6 tween 80	A7 N-acetyl-D-galactosamine	A8 N-acetyl-D-glucosamine	A9 adonitol	A10 L-arabinose	A11 D-arabitol	A12 cellobiose
B1 D-erythritol	B2 D-fructose	B3 L-tucose	B4 D-galactose	B5 gentiobiose	B6 α-D-glucose	B7 m-inositol	B8 α-D-lactose	B9 lactulose	B10 maltose	B11 D-mannitol	B12 D-mannose
C1 D-melibiose	C2 β-methyl D-glucoside	C3 D-psicose	C4 D-raffinose	C5 L-rhamnose	C6 D-sorbitol	C7 sucrose	C8 D-trehalose	C9 turannose	C10 xylytol	C11 methyl pyruvate	C12 mono-methyl succinate
D1 acetic acid	D2 cis-aconitic acid	D3 citric acid	D4 formic acid	D5 D-galactonic acid lactone	D6 D-galacturonic acid	D7 D-gluconic acid	D8 D-glucosaminic acid	D9 D-glucuronic acid	D10 α-hydroxybutyric acid	D11 β-hydroxybutyric acid	D12 γ-hydroxybutyric acid
E1 p-hydroxy phenylacetic acid	E2 itaconic acid	E3 α-keto butyric acid	E4 α-keto glutaric acid	E5 α-keto valeric acid	E6 D,L-lactic acid	E7 malonic acid	E8 propionic acid	E9 quinic acid	E10 D-saccharic acid	E11 sebacic acid	E12 succinic acid
F1 bromo succinic acid	F2 succinamic acid	F3 glucuronamide	F4 alaninamide	F5 D-alanine	F6 L-alanine	F7 L-alanyl-glycine	F8 L-asparagine	F9 L-aspartic acid	F10 L-glutamic acid	F11 glycyl-L-aspartic acid	F12 glycyl-L-glutamic acid
G1 L-histidine	G2 hydroxy L-proline	G3 L-leucine	G4 L-ornithine	G5 L-phenylalanine	G6 L-proline	G7 L-pyrogutamic acid	G8 D-serine	G9 L-serine	G10 L-threonine	G11 D,L-carnitine	G12 γ-amino butyric acid
H1 urocanic acid	H2 inosine	H3 uridine	H4 thymidine	H5 phenyl ethylamine	H6 putrescine	H7 2-amino ethanol	H8 2,3-butanediol	H9 glycerol	H10 D,L-α-glycerol phosphate	H11 glucose-1-phosphate	H12 glucose-6-phosphate

GP MicroPlate™

A1 water	A2 α-cyclodextrin	A3 β-cyclodextrin	A4 dextrin	A5 glycogen	A6 inulin	A7 mannan	A8 tween 40	A9 tween 80	A10 N-acetyl-D-glucosamine	A11 N-acetyl-D-mannosamine	A12 amygdalin
B1 L-arabinose	B2 D-arabitol	B3 arbutin	B4 cellobiose	B5 D-fructose	B6 L-lucose	B7 D-galactose	B8 D-galacturonic acid	B9 gentiobiose	B10 D-gluconic acid	B11 α-D-glucose	B12 m-inositol
C1 α-D-lactose	C2 lactulose	C3 maltose	C4 maltotriose	C5 D-mannitol	C6 D-mannose	C7 D-melezitose	C8 D-melibiose	C9 α-methyl D-galactoside	C10 β-methyl D-galactoside	C11 3-methyl glucose	C12 α-methyl D-glucoside
D1 β-methyl D-glucoside	D2 α-methyl D-mannoside	D3 palatinose	D4 D-psicose	D5 D-raffinose	D6 L-rhamnose	D7 D-ribose	D8 salicin	D9 sedoheptulosan	D10 D-sorbitol	D11 stachyose	D12 sucrose
E1 D-tagatose	E2 D-trehalose	E3 turannose	E4 xylytol	E5 D-xylose	E6 acetic acid	E7 α-hydroxybutyric acid	E8 β-hydroxybutyric acid	E9 γ-hydroxybutyric acid	E10 p-hydroxyphenyl acetic acid	E11 α-keto glutaric acid	E12 α-keto valeric acid
F1 lactamide	F2 D-lactic acid methyl ester	F3 L-lactic acid	F4 D-malic acid	F5 L-malic acid	F6 methyl pyruvate	F7 mono-methyl succinate	F8 propionic acid	F9 pyruvic acid	F10 succinamic acid	F11 succinic acid	F12 N-acetyl L-glutamic acid
G1 alaninamide	G2 D-alanine	G3 L-alanine	G4 L-alanyl-glycine	G5 L-asparagine	G6 L-glutamic acid	G7 glycyl L-glutamic acid	G8 L-pyrogutamic acid	G9 L-serine	G10 putrasone	G11 2,3-butanediol	G12 glycerol
H1 adenosine	H2 2-deoxy adenosine	H3 inosine	H4 thymidine	H5 uridine	H6 adenosine-5-monophosphate	H7 thymidine-5-monophosphate	H8 uridine-5-monophosphate	H9 fructose-6-phosphate	H10 glucose-1-phosphate	H11 glucose-6-phosphate	H12 D-L-α-glycerol phosphate

Date : 15/07/93
 Hour : 24
 Plate Type : GP
 Plate # : 1
 Strain Name : 2305-1
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GP

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=>" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=>" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A 1	0	10	34	<190>	<205>	22	-8	36	-4	-6	20	-33
B 1	-23	-11	-11	33	<222>	11	4	35	38	<94>	<283>	-25
C 1	-3	8	<258>	<231>	13	<58>	<210>	-5	9	-9	48	-33
D 1	24	13	42	<146>	0	15	<82>	<76>	-15	19	14	<92>
E 1	8	<205>	<224>	-2	30	15	<54>	-2	-17	-4	21	-17
F 1	46	<274>	<184>	10	2	<254>	1	-2	<175>	7	15	-33
G 1	17	<171>	<195>	<168>	24	13	21	4	<284>	35	<50>	<272>
H 1	-9	-1	-4	18	23	-3	5	21	-1	23	32	-10

BIO-NUMBER : 0600-0206-1440-0461-3000-3110-3411-0000

SPECIES IDENTIFICATION : STAPHYLOCOCCUS HOMINIS

	CLOSEST SPECIES	SIM.....	DIST....	AVG.....	MA
X					
=> 1)	STAPHYLOCOCCUS HOMINIS	0.994	0.510	0.750	2.7
37					
2)	STAPHYLOCOCCUS AURICULARIS	0.000	3.260	1.000	3.0
63					
3)	STAPHYLOCOCCUS HAEMOLYTICUS	0.000	3.287	1.375	4.3
75					
4)	STAPHYLOCOCCUS CORNII	0.000	4.462	0.938	5.0
25					
5)	STAPHYLOCOCCUS CAPITIS SS UREOLYTICUS	0.000	5.324	1.000	2.3
13					
6)	STAPHYLOCOCCUS EQUORUM	0.000	5.964	1.188	2.4
19					
7)	STAPHYLOCOCCUS ARLETTAE	0.000	6.685	0.656	6.1
31					
8)	ROTHIA DENTROCARIOSIA	0.000	6.735	0.438	1.3
50					
9)	STAPHYLOCOCCUS WARNERI	0.000	7.107	0.438	3.4
56					
10)	STAPHYLOCOCCUS SAPROPHYTICUS	0.000	7.459	0.729	1.8
94					
other :		-----	-----	-----	---

ABBREVIATED NAME : STA.HOM
 FULL NAME : STAPHYLOCOCCUS HOMINIS
 DATA BASE CATEGORY : ENVIRONMENTAL

4 HOUR DATA :

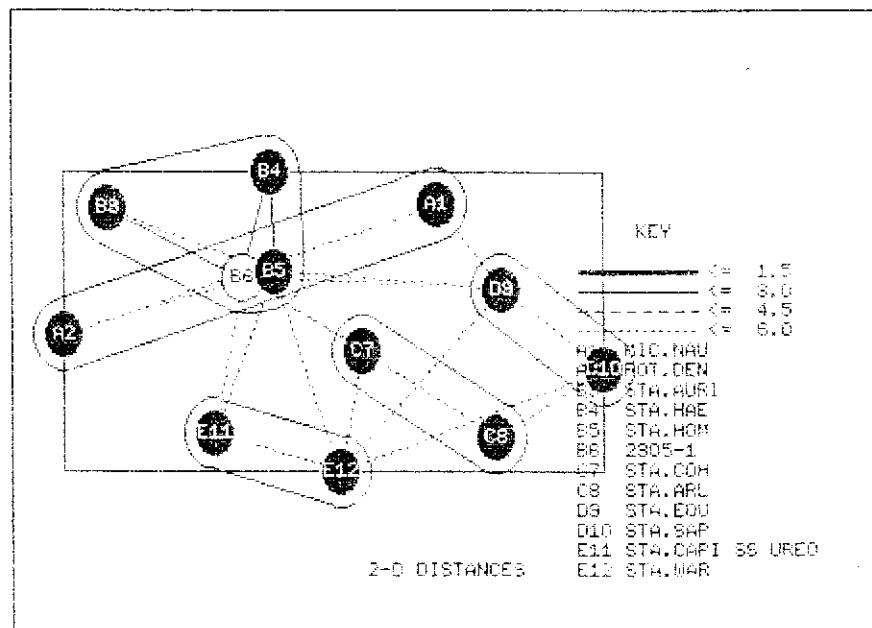
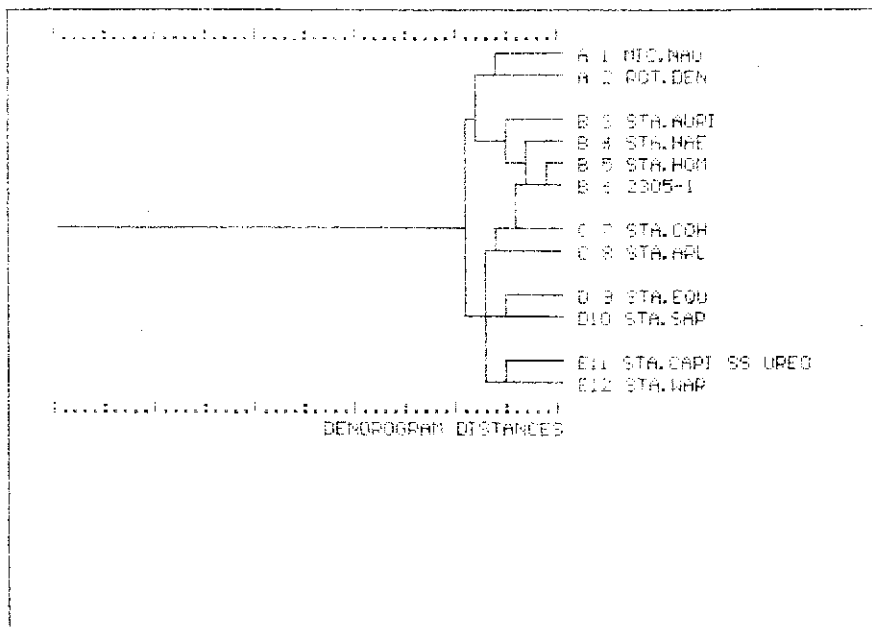
	1	2	3	4	5	6	7	8	9	10	11	12
A I	0	0	0	100	83	0	0	0	0	20	0	0
B I	0	13	0	0	100	0	0	0	0	30	100	0
C I	20	20	100	100	0	38	56	0	0	0	52	0
D I	0	0	0	92	0	0	56	47	0	0	0	100
E I	0	87	100	0	63	0	53	0	0	0	0	0
F I	60	100	92	0	0	100	0	0	100	0	0	0
G I	0	83	83	43	0	0	0	0	75	0	37	90
H I	0	0	0	0	0	0	0	0	31	0	0	0

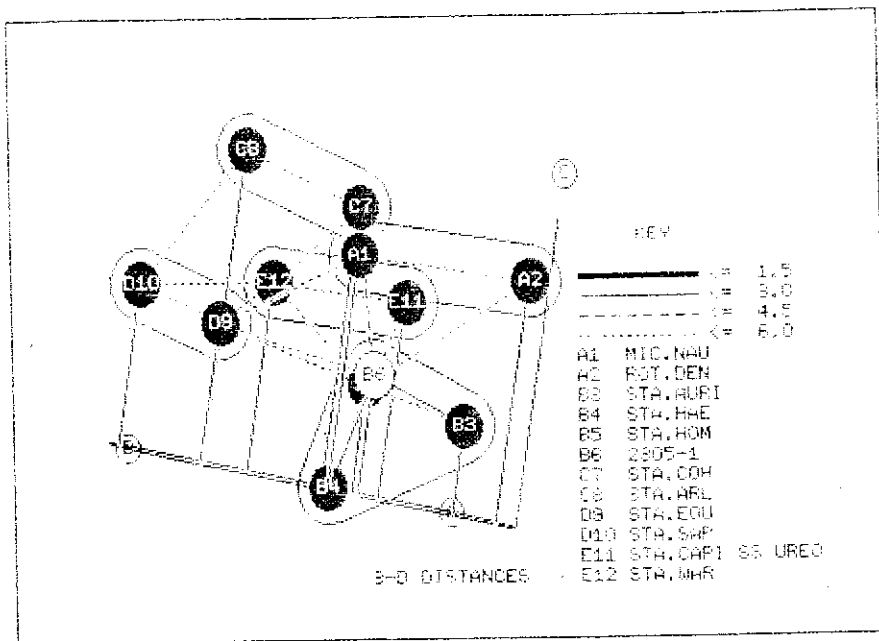
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A I	0	15	0	100	92	0	0	13	0	54	0	13
B I	0	13	0	6	100	0	13	0	0	42	100	0
C I	40	25	100	100	0	38	56	0	0	0	52	0
D I	0	0	6	92	0	0	56	63	0	15	0	100
E I	0	92	100	6	63	0	54	0	0	0	0	10
F I	60	100	92	0	0	100	0	0	100	0	0	21
G I	0	83	83	71	0	21	0	13	75	0	40	92
H I	10	23	0	10	10	0	0	0	31	0	0	13

CLOSEST SPECIES :

- 1) 3.244 : STAPHYLOCOCCUS HAEMOLYTICUS
- 2) 4.949 : STAPHYLOCOCCUS EQUORUM
- 3) 5.897 : STAPHYLOCOCCUS AURICULARIS
- 4) 5.960 : STAPHYLOCOCCUS CAPITIS SS UREOLYTICUS
- 5) 6.171 : ROTHIA DENTROCARIOSEA





Date : 16/07/93
 Hour : 24
 Plate Type : GP
 Plate # : 2
 Strain Name : 2305-2
 Strain # : W.L00
 Other Info : 7
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GP

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus AI control well
 <XXX> = positive, <XXX> = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	-2	-4	-2	-1	1	<409>	<285>	-16	3	9
B	-3	-3	5	-3	5	-10	2	-3	-21	5	5	4
C	-6	-4	-3	-6	-2	-7	-2	-3	-21	-11	6	7
D	-1	0	1	2	3	-7	8	-2	4	-17	5	8
E	8	0	4	2	25	<553>	<673>	<707>	<548>	17	<657>	<434>
F	<49>	<779>	<807>	<671>	<771>	<776>	<710>	<509>	<740>	<732>	<761>	<762>
G	<286>	<321>	<243>	<237>	<791>	<790>	<436>	<714>	<610>	1	<36>	25
H	23	<507>	<656>	16	19	13	15	17	<126>	9	26	25

BIO-NUMBER : 0030-0000-0000-0000-0173-3777-7770-3010

NO IDENTIFICATION

	CLOSEST SPECIES :	SIM.....	DIST.....	AUG.....	MA
X	=> 1) CDC GROUP D-2	0.286	9.994	0.375	1.9
87	2) CDC GROUP B-1 / B-3	0.120	10.251	0.333	3.1
75	3) MICROCOCCUS DIVERSUS	0.000	12.674	0.063	0.1
19	4) BACILLUS SPHAERICUS	0.000	13.105	2.188	4.0
06	5) CORYNEBACTERIUM PSEUDODIPHTHERITICUM	0.000	14.958	1.563	8.4
38	6) BACILLUS BADIUS	0.000	15.260	0.563	1.3
56	7) RHODOCOCOCUS EQUI	0.000	16.837	0.250	2.3
13	8) RHODOCOCOCUS RHODOCHROUS	0.000	16.934	0.063	0.1
19	9) CORYNEBACTERIUM VARIABILIS	0.000	17.007	0.375	3.3
38	10) RHODOCOCOCUS ERYTHROPOLIS	0.000	17.085	0.781	1.0
69	other :	-----	-----	-----	---

ABBREVIATED NAME : CDC D-2
 FULL NAME : CDC GROUP D-2
 DATA BASE CATEGORY : ENVIRONMENTAL

4 HOUR DATA :

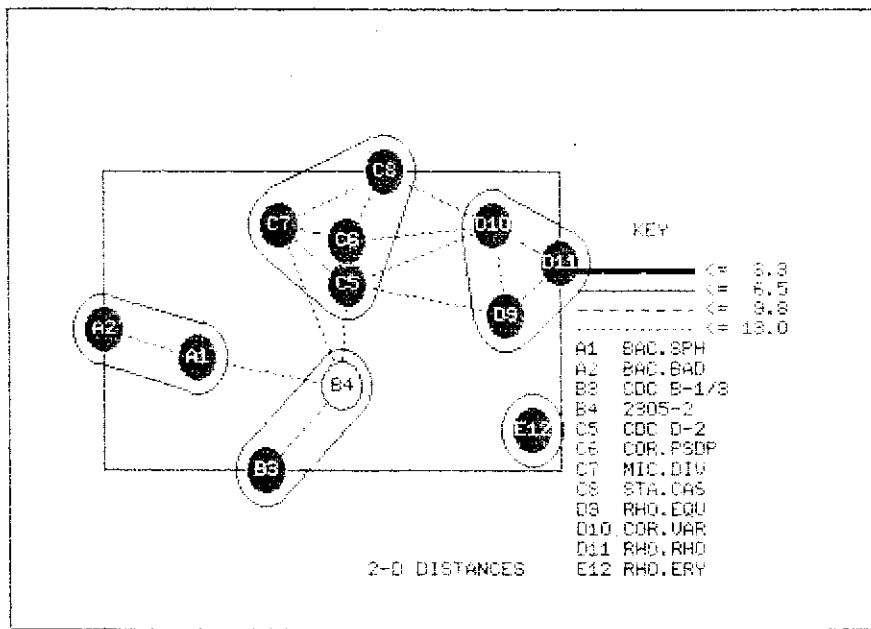
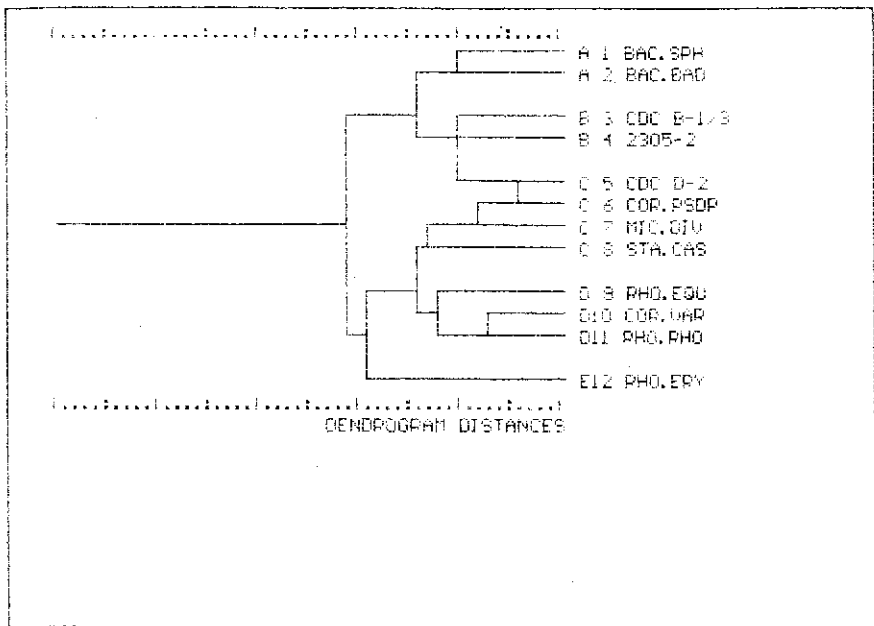
	1	2	3	4	5	6	7	8	9	10	11	12
A :	0	8	0	0	0	0	58	50	50	0	0	0
B :	25	0	0	0	0	50	0	0	0	0	17	0
C :	0	0	0	0	0	17	0	0	0	0	17	0
D :	0	0	0	0	0	42	75	8	0	0	0	0
E :	17	0	0	17	67	100	58	33	17	0	8	0
F :	58	0	58	0	100	100	17	8	100	100	50	100
G :	42	0	17	0	100	100	33	100	67	0	0	8
H :	50	0	0	0	50	0	0	25	0	0	0	0

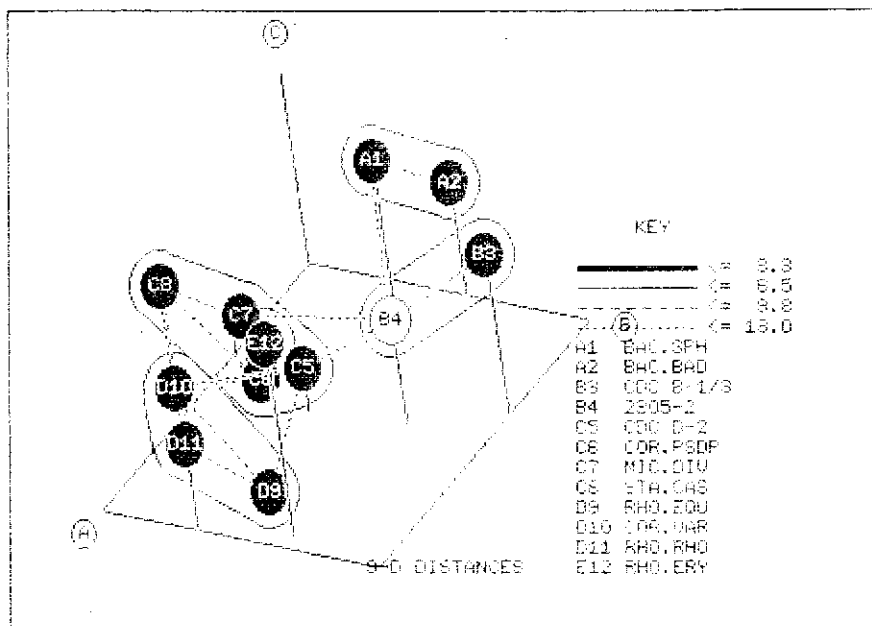
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A :	0	8	0	0	0	0	58	50	50	0	0	0
B :	25	0	0	0	0	50	0	0	0	0	25	0
C :	0	0	0	0	0	17	0	0	0	0	25	0
D :	0	0	0	0	0	42	75	8	0	0	0	0
E :	17	0	0	17	67	100	58	33	17	0	8	0
F :	58	0	58	0	100	100	50	8	100	100	67	100
G :	42	0	25	0	100	100	33	100	67	0	0	8
H :	50	0	42	0	50	0	0	25	0	0	0	0

CLOSEST SPECIES :

- 1) 4.022 : CORYNEBACTERIUM PSEUDODIPHThERITICUM
- 2) 8.129 : CDC GROUP F-1 SUBGROUP 8
- 3) 8.308 : MICROCOCCUS DIVERSUS
- 4) 10.168 : CORYNEBACTERIUM CYSTITIDIS
- 5) 11.044 : BACILLUS BREVIS





Date : 18/07/93
 Hour : 24
 Plate Type : GN
 Plate # : 1
 Strain Name : 2305-3
 Strain # : W.L00
 Other Info : 7
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A 1	0	-10	-12	-8	<323>	<193>	-18	-13	-10	-5	-10	-5
B 1	-8	-8	-16	-12	-13	-16	-12	-14	-12	-14	-12	-11
C 1	-9	-11	-12	-11	-13	-11	-15	-13	-13	-16	<544>	<505>
D 1	<359>	<295>	-1	<306>	-17	-12	<122>	-10	-7	<300>	<501>	<48>
E 1	<83>	-10	<264>	<310>	-7+	<566>	-16	<370>	-6	-7	<317>	<567>
F 1	<407>	<561>	-14	<20+>	<38+>	<57>	<54>	<444>	<479>	<540>	-10	-3
G 1	<58>	-9	<95>	-7	<48>	<208>	<528>	-6	<24>	<143>	-7	-8
H 1	<426>	<375>	-4	-7	-8	-9	-10	<10-	3	0	-5	-7

BIO-NUMBER : 0300-0000-0003-6447-5523-6374-5344-6000

SPECIES IDENTIFICATION : ALCALIGENES FAECALIS TYPE II

	CLOSEST SPECIES	SIM.	DIST.	AVG.	MA
X					
=>	1) ALCALIGENES FAECALIS TYPE II	0.755	2.432	1.938	5.0
94					
2)	COMAMONAS ACIDOVORANS	0.033	3.496	2.250	6.8
63					
3)	COMAMONAS TESTOSTERONI	0.004	4.203	1.688	5.1
81					
4)	ACINETOBACTER SPECIES GROUP B3	0.000	5.452	0.234	4.6
50					
5)	ALCALIGENES DENITRIFICANS/PIECHAUDII	0.000	6.495	0.857	3.8
25					
6)	ALCALIGENES FAECALIS SS FAECALIS	0.000	6.873	0.388	2.9
31					
7)	COMAMONAS TERRIGENA	0.000	7.076	1.938	8.3
44					
8)	PSEUDOMONAS PSEUDOALCALIGENES	0.000	7.176	1.750	7.2
13					
9)	CDC GROUP IVC-2	0.000	7.338	0.101	1.6
44					
10)	ACINETOBACTER GENOSPECIES 11	0.000	7.467	1.313	3.4
69					
other :					

ABBREVIATED NAME : ALC.FAE TYPE II
 FULL NAME : ALCALIGENES FAECALIS TYPE II
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

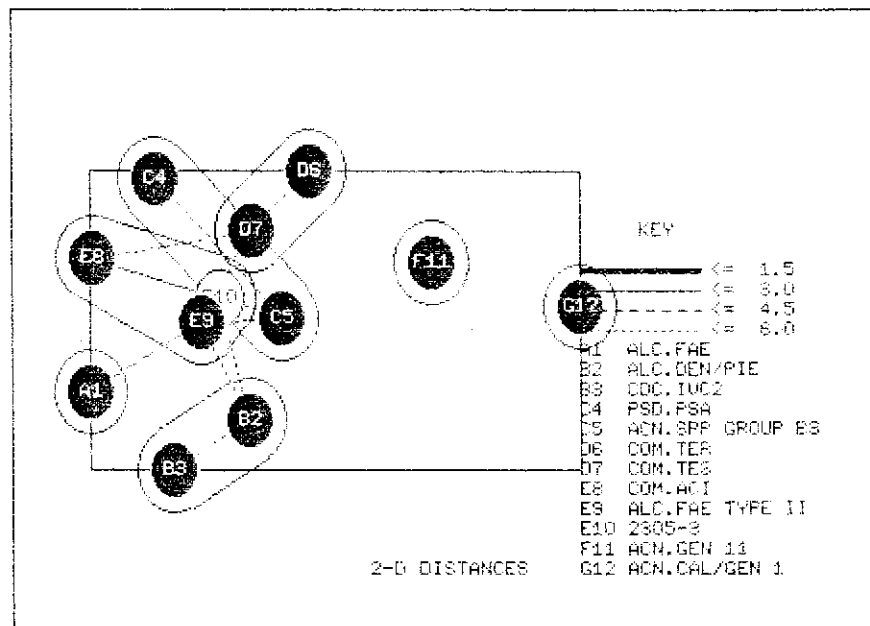
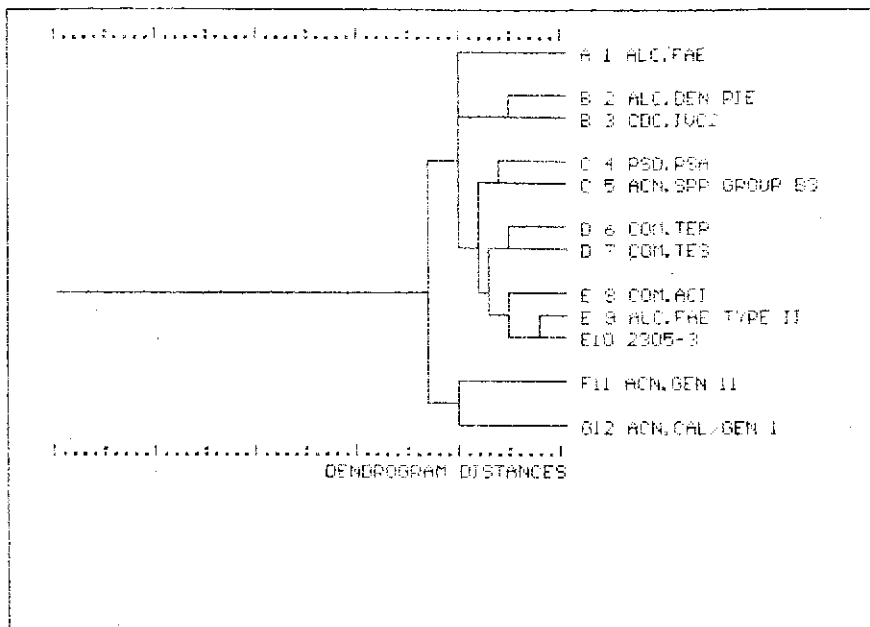
	1	2	3	4	5	6	7	8	9	10	11	12
A :	0	16	16	52	52	52	0	6	0	0	0	0
B :	20	0	0	0	0	0	0	0	0	0	0	0
C :	0	16	8	0	8	0	0	0	0	0	76	76
D :	92	44	28	60	0	8	28	6	16	76	100	0
E :	52	28	92	76	64	100	28	80	24	0	40	92
F :	92	92	16	48	84	76	16	92	84	100	12	40
G :	36	16	84	44	88	88	56	44	60	24	0	28
H :	52	36	12	0	24	12	0	12	8	0	0	0

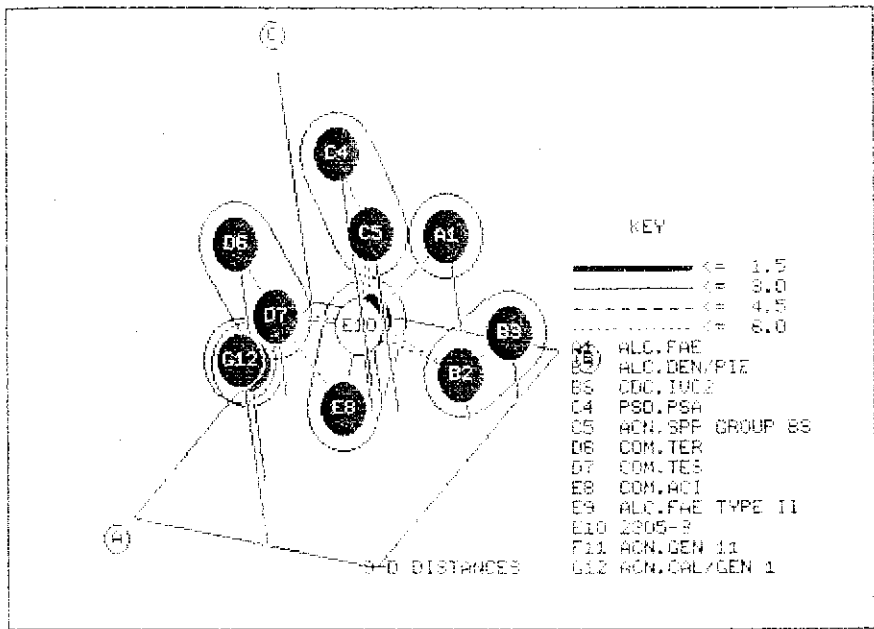
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A :	0	0	20	16	88	88	0	0	0	0	0	0
B :	0	0	0	0	0	0	0	0	0	0	0	0
C :	0	0	0	0	0	0	0	0	0	0	100	100
D :	100	88	52	64	0	20	44	0	20	100	100	24
E :	80	56	100	68	100	100	20	100	36	0	44	100
F :	100	100	20	100	100	100	32	100	92	100	0	16
G :	60	0	92	40	100	100	60	68	80	72	0	40
H :	64	32	8	0	36	8	0	0	40	8	0	0

CLOSEST SPECIES :

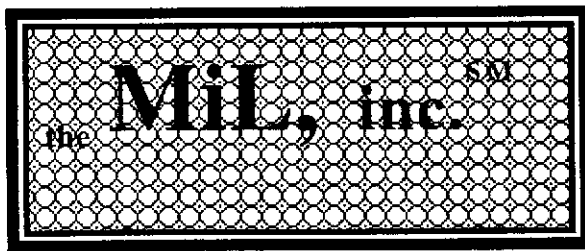
- 1) 4.115 : ALCALIGENES FAECALIS SS FAECALIS
- 2) 4.927 : BORDETELLA BRONCHISEPTICA
- 3) 5.161 : AQUASPIRILLUM PUTRIDICONCHYLUM
- 4) 5.247 : ACINETOBACTER SPECIES GROUP B3
- 5) 5.450 : ALCALIGENES DENITRIFICANS/PIECHAUDII





Warranty and Limits of Liability

In accepting analytical work, we warrant the accuracy of test results under the conditions employed in the laboratory. The foregoing express warranty is exclusive and is given in lieu of all other warranties, expressed or implied. We disclaim any other warranties, expressed or implied, including a Warranty of Fitness for Particular Purpose and Warranty of Merchantability. We accept no legal responsibility for the purposes for which the client uses the test results.



**Microbe
Inotech
Laboratories,
inc.**

1840 Craig Road
St. Louis, MO
63146-4712
U.S.A.

Telephone: (314) 878-6626
(800) 688-9144

FAX: (314) 878-9376

E-mail: Bruce C. Hemming
76177.204@compuserve.com

**Total
Heterotrophic
Plate Count
Analysis &
Microbial
Identification**

Report Prepared For:

**Environmental & Technical
Services**

**ATTN: Walter Loo, R.G.,
C.E.G.**

**2081 15th Street
San Francisco, CA 94114**

Client Phone (415) 861-0810

Client Fax (415) 861-3269

Report No. MILB—2312

PO Number none

July 19, 1993

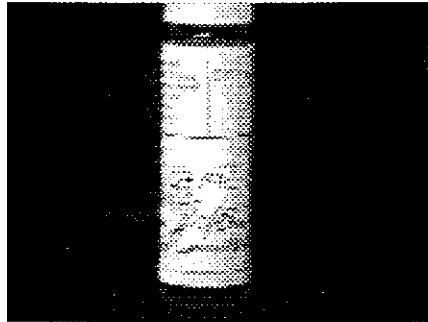
Summary Report of Analysis
[No. 2312]

Environmental & Technical Services
ATTN: Walter Loo, R.G., C.E.G.
2081 15th Street
San Francisco, CA 94114

July 19, 1993

Description:

Wed, Jul 14, 1993 - 1:00 PM : Received by FEDEX one 100 ml water sample labeled CHIC, MW4 taken 7-13-93 at 4:00 pm Analyses requested are Total Plate Count and Bacterial ID. MiL #2312 sac. pix



Chain of Custody Record Information

Purchase Order No.—none
MiL, Inc. REPORT & Invoice No.: MILB-2312

Processing:

[Standard Bacterial Plate Count - serial dilution method and direct spread plate count] Within 20 minutes of reception aliquots from each of the samples was checked for weight and then serially diluted. Each dilution was sterilely transferred in a laminar flow biological cabinet and placed on previously prepared and dried trypticase soy broth agar (TSBA) medium in Petri plates.

Observations for colony forming units (CFU) were made at 24 and 48 hrs. of incubation at 28°C for each sample. Colony differentiation was noted at 48 hrs.

Summary Final Results—Total Heterotrophic Plate Count:

DATA:	Direct Count: Colony Forming Units (CFU/ g) on TSBA Medium	
	24 Hrs.	48 Hrs.
Sample:		
CHIC-MW4	2.32 x 10 ⁶	9.01 x 10 ⁶
	<u>Distinct morphological Colony Types at 48 Hrs. in Sample</u>	
	6	

Percentages of Types In Sample:

Sample	CHIC-MW4	Description
<u>Strain</u>		
1	60%	large clear white & crusty
2	10%	small yellow
3	10%	small white
4	10%	rough, yellowish
5	5%	glossy center, rough edge,
		small yellowish
6	5%	fuzzy large yellow

Summary of GC-FAME/Biolog Analyses							
Strain Name	Primary Identification by GC	Sim. Coef.	Dist. Coef.	Primary ID by Biolog™	Plate Type	Sim. Coef	Dist. Coef
2312-1	<i>Pseudomonas stutzeri</i>	0.772	2.326	Nearest neighbor: <i>Pseudomonas stutzeri</i>	GN	0.23	13.644
2312-2	<i>Pseudomonas stutzeri</i>	0.713	2.659	<i>Pseudomonas alcaligenes A</i>	GN	0.904	1.591
2312-3	<i>Pseudomonas putida</i> biotype B	0.744	2.488	<i>Pseudomonas fluorescens B</i>	GN	0.889	1.307
2312-4	<i>Pseudomonas stutzeri</i>	0.311	4.941	<i>Pseudomonas stutzeri</i>	GN	0.51	8.193
2312-5	<i>Pseudomonas stutzeri</i>	0.666	2.912	Nearest neighbor: 1) <i>Vibrio mimicus</i> 2) <i>P. stutzeri</i>	GN	0.056	17.889 19.963
2312-6	<i>Pseudomonas aeruginosa</i>	0.799	2.168	<i>Pseudomonas aeruginosa</i>	GN	0.621	5.49

Note: *P. stutzeri* isolates shown are distinct strains as demonstrated by distinct metabolic profiles of the Biolog assay, colony morphologies, and significant differences in GCFAME indices.

GC-FAME & Biolog™ Processing:

Following isolation the strains were individually streaked out onto TSBA. The TSBA plates were processed after 24 hr incubation by [Method 1 - Standard GC-FAME]. The strains were examined against both the newly installed Aerobe (TSBA [rev. 3.70]) and Clinical Aerobe (CLIN [rev.3.70]) GC-FAME databases. Subsequently the strains were prepared for Biolog™ analysis by suspending them in sterile saline and loading the solutions into the appropriate microtiter plates (Gram negative or Gram positive). The plates were incubated for 24 hours and then examined against version 3.0 of the Biolog™ database using an automated microplate reader.

Similarity and Distance Coefficient

In order to create the database that we use to identify your organisms, thousands of species of bacteria had to be tested. In fact each species itself had to be tested hundreds of times to determine a set of characteristics unique to it. The species characteristics that are in our database are an "average" of the characteristics of hundreds of tested bacteria of the same species. The Similarity and Distance Coefficient of your organism refers to the similarity and distance to the hypothetical 'mean' organism in the database. The database organism has a similarity coefficient of one and a distance of zero. So the closer your strain is to one and zero the more closely it matches the mean organism in the database.

A good match is one with a similarity coefficient greater than .5 and a distance coefficient of less than 7.

Thank you from the staff on project

Julie Milke - Laboratory Manager

Dr. Bruce C. Hemming - Operations Director

SECRET

CALIB. 1 CALIBRATION STANDARD [AEROBE] 16-JUL-93 01:38:39 Area: 475080 % Named: 100
 ** GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0019. **

CALIB. 1 CALIBRATION STANDARD [AEROBE] 17-JUL-93 02:08:52 Area: 484904 % Named: 100
 ** GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0013. **

2 2312-1 [AEROBE] 17-JUL-93 03:09:29 Area: 160120 % Named: 100
 TSBA [Rev 3.70] Pseudomonas 0.772 (includes P. perfectomarina)
 P. stutzeri 0.772 (includes P. perfectomarina)
 P. alcaligenes 0.479
 P. mendocina 0.444
 CLIN [Rev 3.70] Pseudomonas 0.709
 P. stutzeri 0.709

3 2312-2 [AEROBE] 17-JUL-93 03:39:42 Area: 226680 % Named: 100
 TSBA [Rev 3.70] Pseudomonas 0.331 (includes P. perfectomarina)
 P. stutzeri 0.331 (includes P. perfectomarina)
 CLIN [Rev 3.70] Pseudomonas 0.713
 P. stutzeri 0.713

4 2312-3 [AEROBE] 17-JUL-93 04:09:57 Area: 94416 % Named: 100
 TSBA [Rev 3.70] Pseudomonas 0.633 (Pseudomonas aureofaciens)
 P. chlororaphis 0.633 (Pseudomonas aureofaciens)
 CLIN [Rev 3.70] Pseudomonas 0.744
 P. putida 0.744
 P. p. biotype B 0.744
 P. p. biotype A 0.664
 P. fluorescens 0.695

5 2312-4 [AEROBE] 17-JUL-93 04:40:13 Area: 290112 % Named: 100
 TSBA [Rev 3.70] Pseudomonas 0.101 (includes P. perfectomarina)
 P. stutzeri 0.101 (includes P. perfectomarina)
 P. mendocina 0.054
 Neisseria 0.072
 N. mucosa 0.072
 CLIN [Rev 3.70] Pseudomonas 0.311
 P. stutzeri 0.311

16-JUL-93 23:35:51

6 2312-5 [AEROBE] 17-JUL-93 05:10:33 Area: 212616 % Named: 100
 TSBA [Rev 3.70] Pseudomonas 0.289 (includes P. perfectomarina)
 P. stutzeri 0.289 (includes P. perfectomarina)
 Neisseria 0.187
 N. mucosa 0.187
 CLIN [Rev 3.70] Pseudomonas 0.666
 P. stutzeri 0.666

7 2312-6 [AEROBE] 17-JUL-93 05:40:47 Area: 265376 % Named: 99
 TSBA [Rev 3.70] Pseudomonas 0.657
 P. aeruginosa 0.657
 CLIN [Rev 3.70] Pseudomonas 0.799
 P. aeruginosa 0.799

ID: 1 CALIBRATION STANDARD Date of run: 17-JUL-93 02:09:52
 Bottle: 1 CALIBRATION [AEROBE]

RT	Area	Ar/Ht	Respon	ECL	Name	Z	Comment 1	Comment 2
1.670	137742208	0.023	. . .	7.043	SOLVENT PEAK	. . .	< min rt	
2.666	20648	0.024	1.211	9.000	9:0	5.15		
3.175	42296	0.026	1.144	10.000	10:0	9.97	Peak match	-0.0012
3.874	22384	0.029	1.093	11.000	11:0	5.04	Peak match	-0.0016
4.017	9040	0.030	1.086	11.155	10:0 20H	2.02	Peak match	0.0026
4.262	4672	0.031	1.076	11.420	10:0 30H	1.04	Peak match	0.0014
4.797	47072	0.032	1.055	12.000	12:0	10.23	Peak match	-0.0009
5.952	24096	0.035	1.028	13.000	13:0	5.10	Peak match	0.0062
7.322	48896	0.039	1.008	14.000	14:0	10.15	Peak match	-0.0002
8.863	25000	0.042	0.991	15.000	15:0	5.10	Peak match	-0.0016
9.200	10640	0.044	0.988	15.203	14:0 20H	2.17	Peak match	0.0018
9.672	5296	0.044	0.984	15.487	Sum In Feature 3	1.07	Peak match	0.0020
10.524	50768	0.044	0.976	16.000	16:0	10.21	Peak match	-0.0011
12.252	26016	0.046	0.959	17.000	17:0	5.14	Peak match	-0.0008
12.662	11344	0.050	0.955	17.234	16:0 20H	2.23	Peak match	0.0011
14.005	52112	0.048	0.937	18.000	18:0	10.06	Peak match	-0.0001
15.753	26344	0.049	0.907	19.000	19:0	4.92	Peak match	0.0007
17.477	58280	0.053	0.866	20.000	20:0	10.40		
19.155	16960	0.108	. . .	20.973	> max ar/ht	
19.334	17536	0.165	. . .	21.077	> max ar/ht	
19.625	25728	0.196	. . .	21.246	> max ar/ht	
*****	5296	SUMMED FEATURE 3	1.07	12:0 ALDE ?	unknown 10.928
*****	16:1 ISO I/14:0 30H	14:0 30H/16:1 ISO I

Solvent Ar	Total Area	Named Area	X Named	Total Amt	Nbr Ref	ECL	Deviation	Ref ECL	Shift
137742208	484904	484904	100.00	485552	0

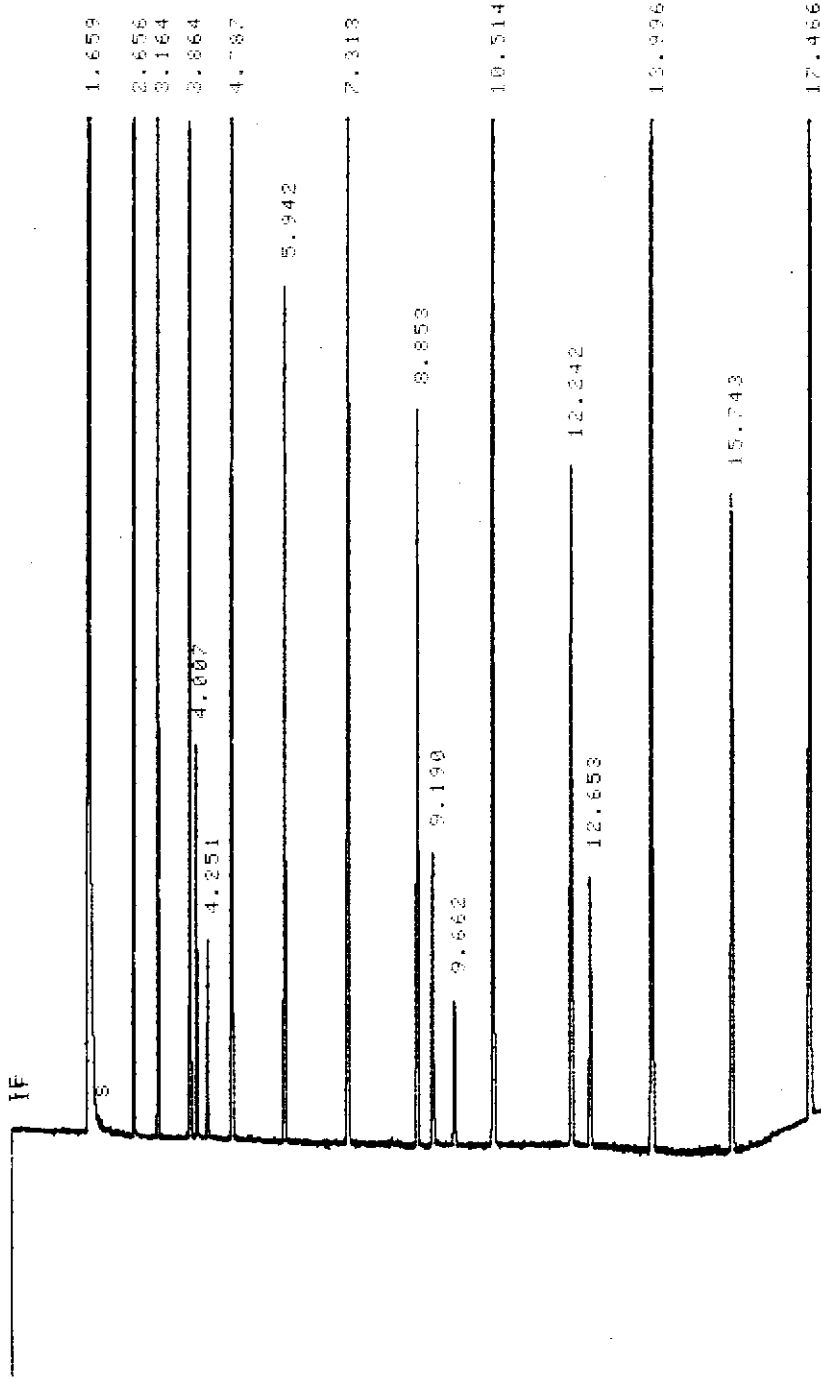
GOOD PEAK MATCHING: PEAK POSITION MATCHING ERROR (RMS) IS 0.0013.

BOTTLE: 1 ID#: 1SAT 17-JUL-93 00:05:47

FILE DATA:F93716849

CALIBRATION STANDARD

RUN # 2 JUL 17, 1993 02:08:52
START



STOP

RUN # 2 JUL 17, 1993 02:08:52
START-No plot
END OF SIGNAL

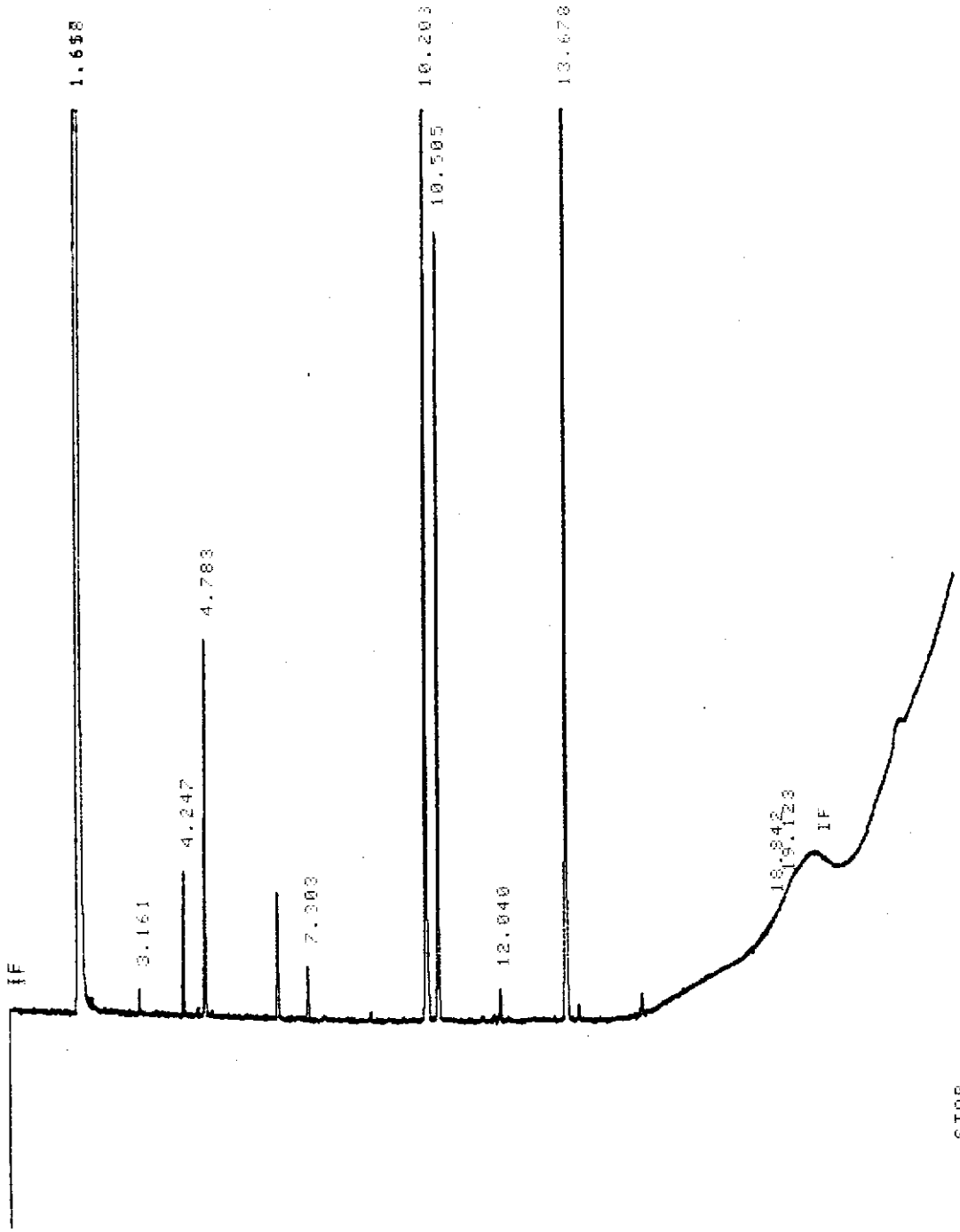
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=====

BOTTLE: 3 ID#: 2SAT 17-JUL-93 01:07:09 FILE DATA:F93716849

2312-1

RUN # 4 JUL 17, 1993 03:09:29
START



STOP

RUN # 4 JUL 17, 1993 03:09:29
START-H0 PLOT
END OF SIGNAL

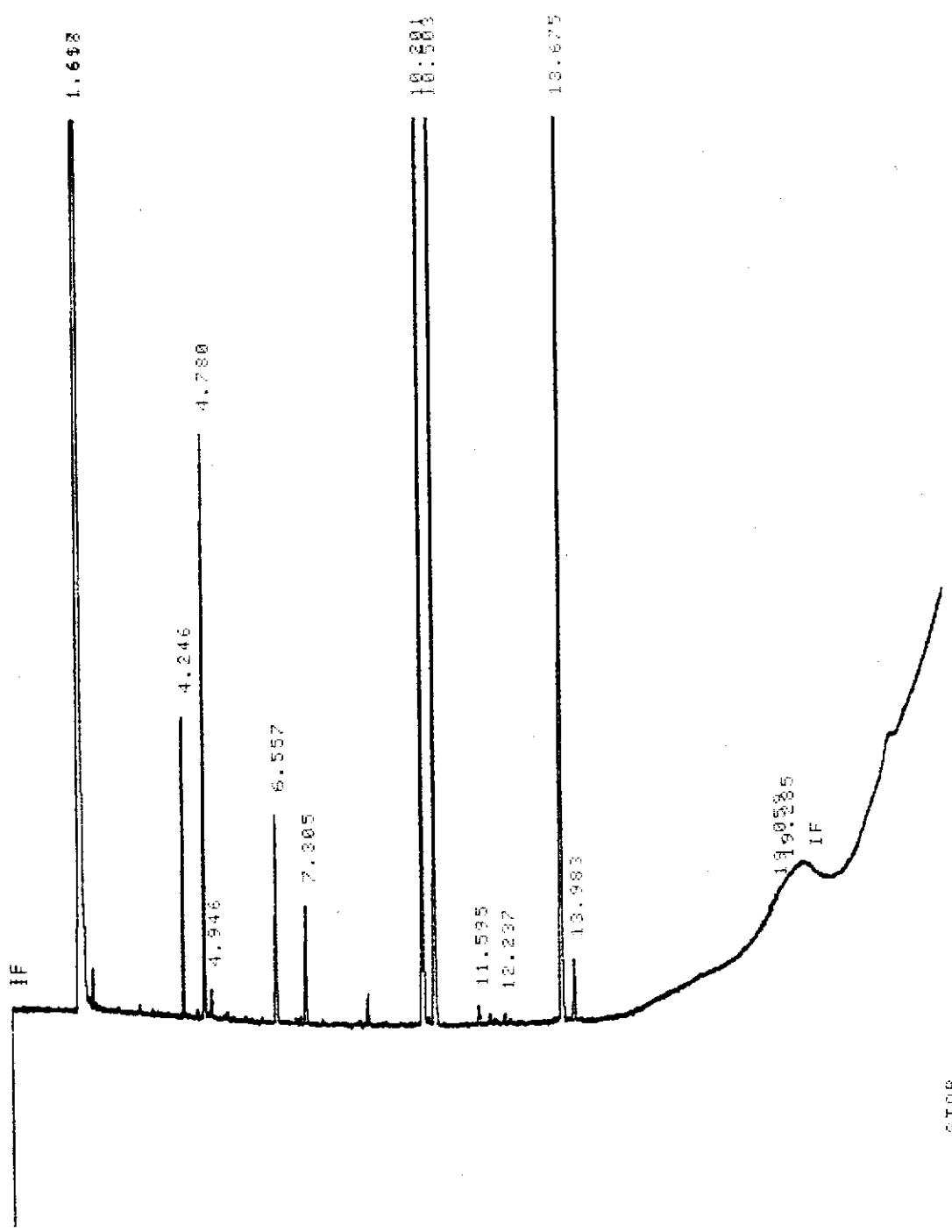
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BOTTLE: 4 ION: 3SAT 17-JUL-93 01:36:56 FILE DATA:F93016849

2312-2

RUN # 5 JUL 17, 1993 03:39:42

START



STOP

RUN # 5 JUL 17, 1993 03:39:42

START-H0 P107

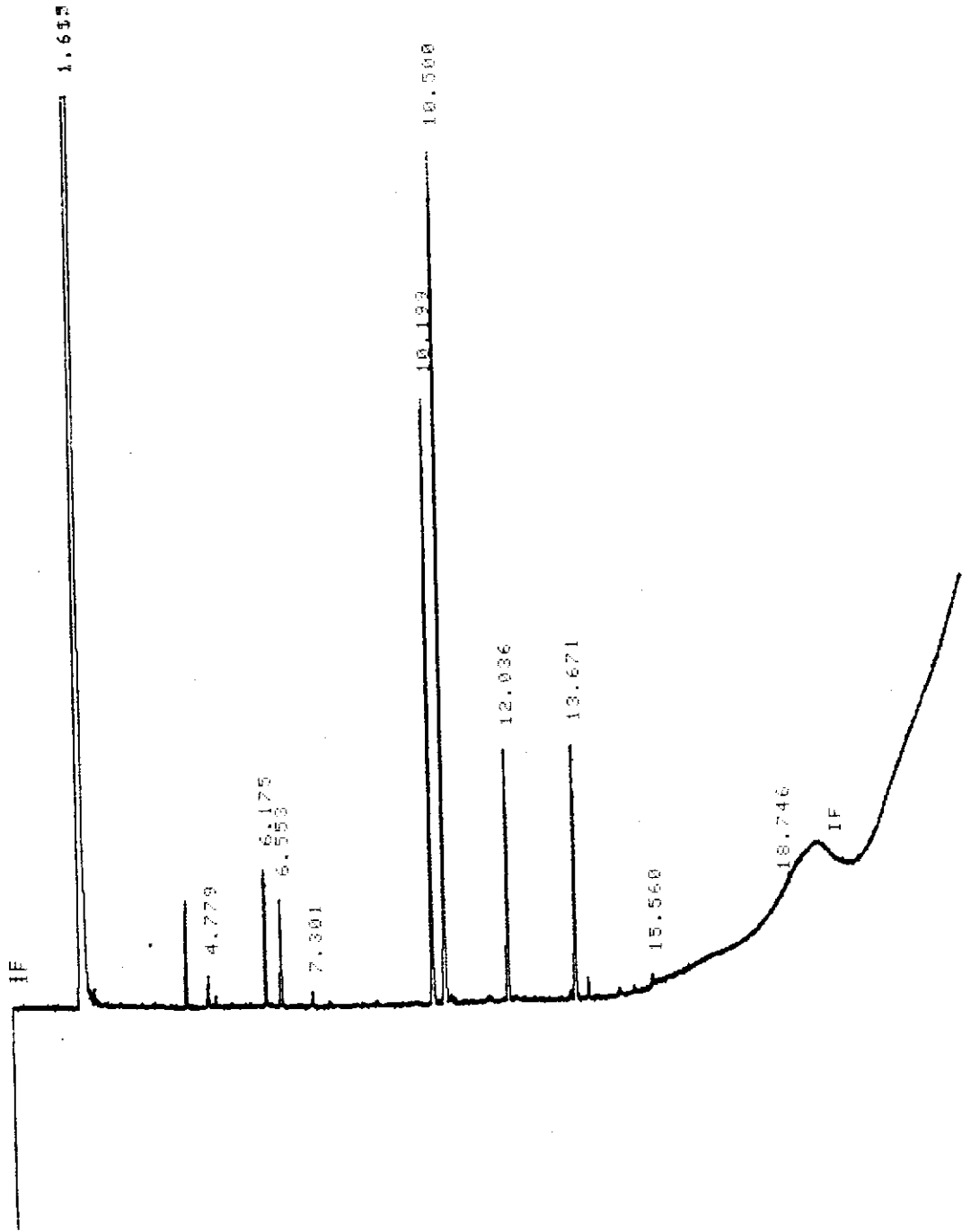
END OF SIGNAL

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BOTTLE: 5 ID#: 46AT 17-JUL-93 02:07:17 FILE DATA:F93716849

2312-3

RUN # 6 JUL 17, 1993 04:09:57
START



STOP

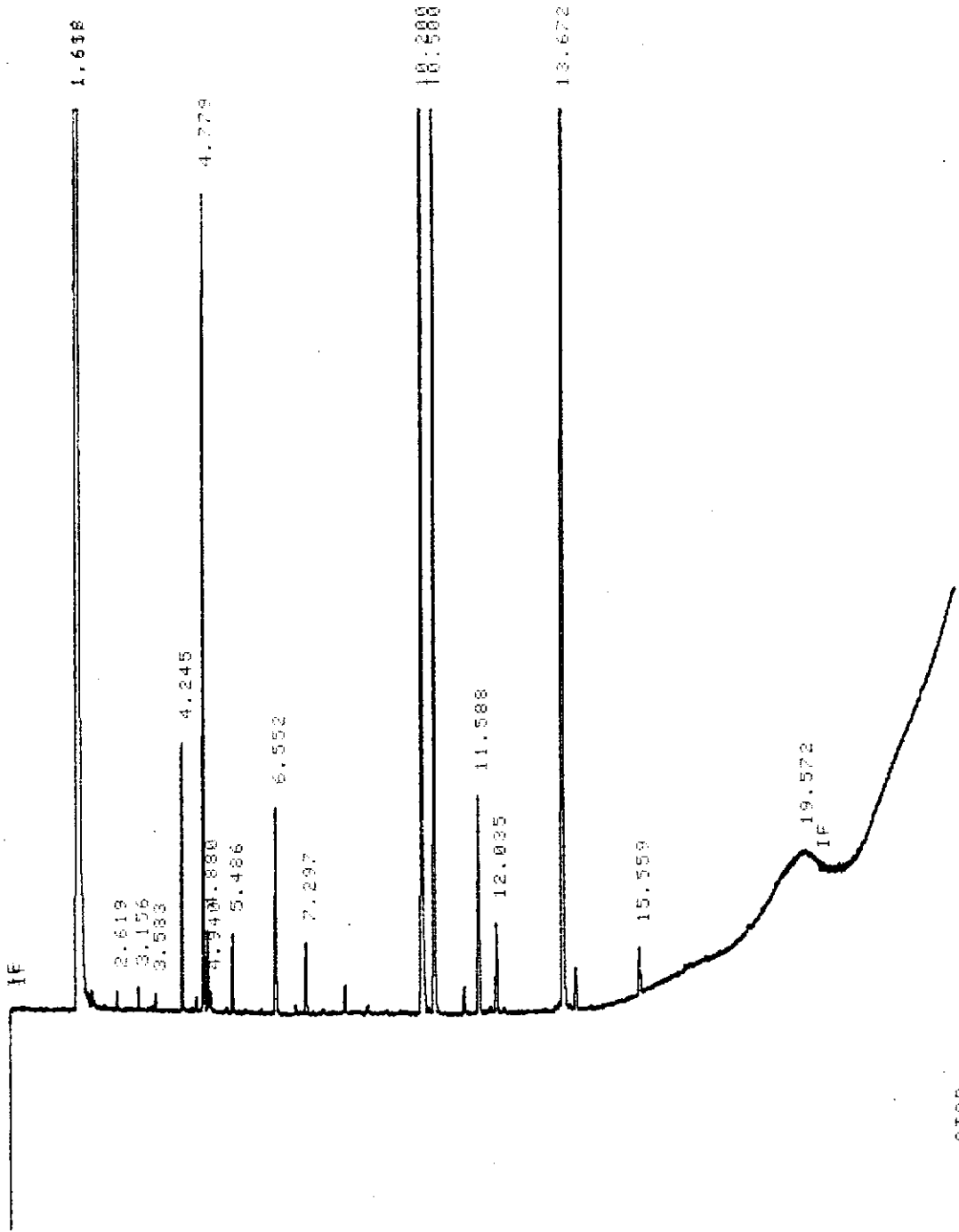
RUN # 6 JUL 17, 1993 04:09:57
START-NO PLOT
END OF SIGNAL

BOTTLE: 6 ID#: 55AT 17-JUL-93 02:37:18

FILE DATA:F93716849

2312-4

RUN # 7 JUL 17, 1993 04:40:13
START



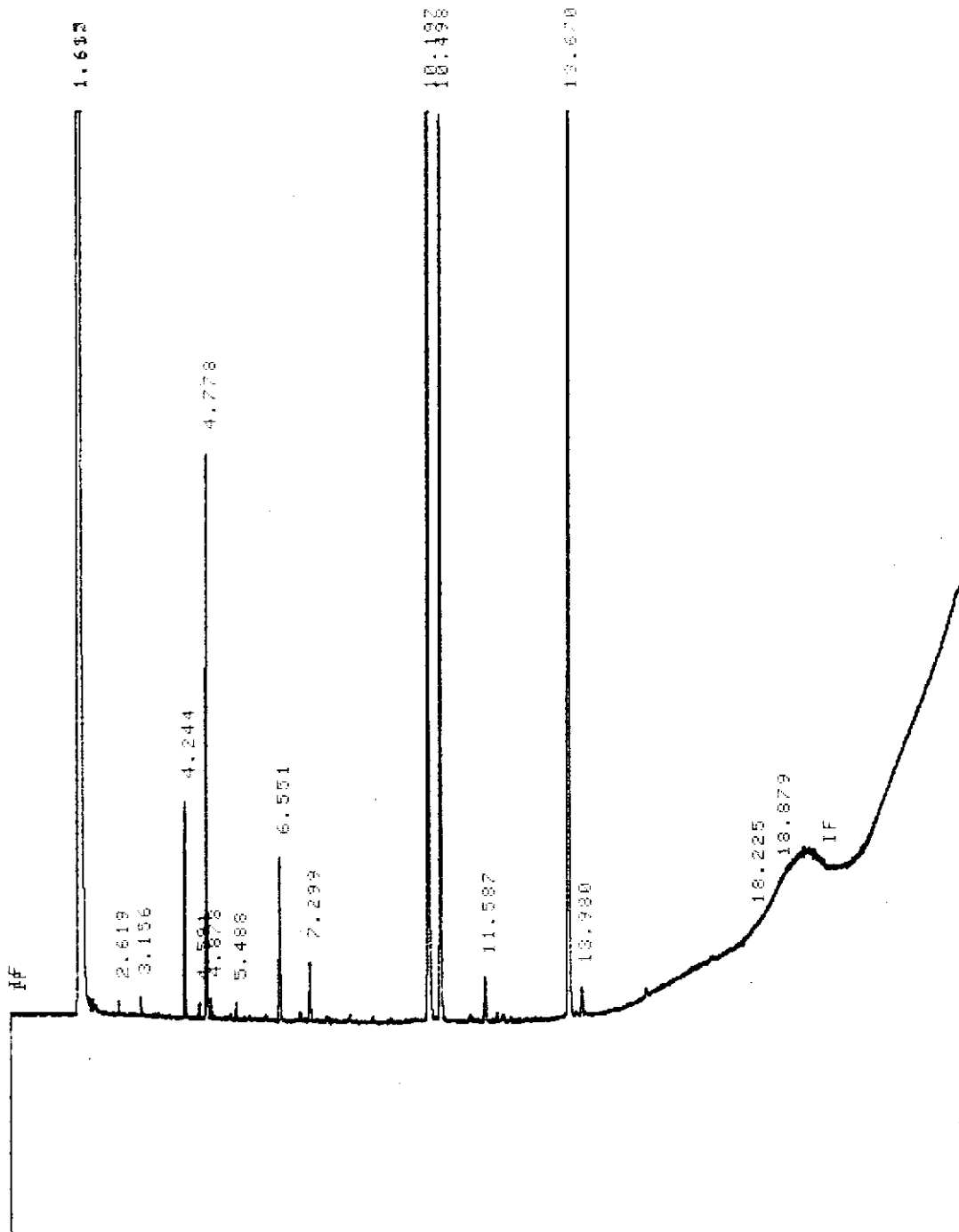
STOP

RUN # 7 JUL 17, 1993 04:40:13
START-No Plot
END OF SIGNAL

BOTTLE: 7 ID#: 6SAT 17-JUL-93 03:08:13 FILE DATA:F93716849

2312-5

RUN # 8 JUL 17, 1993 05:10:33
START



STOP

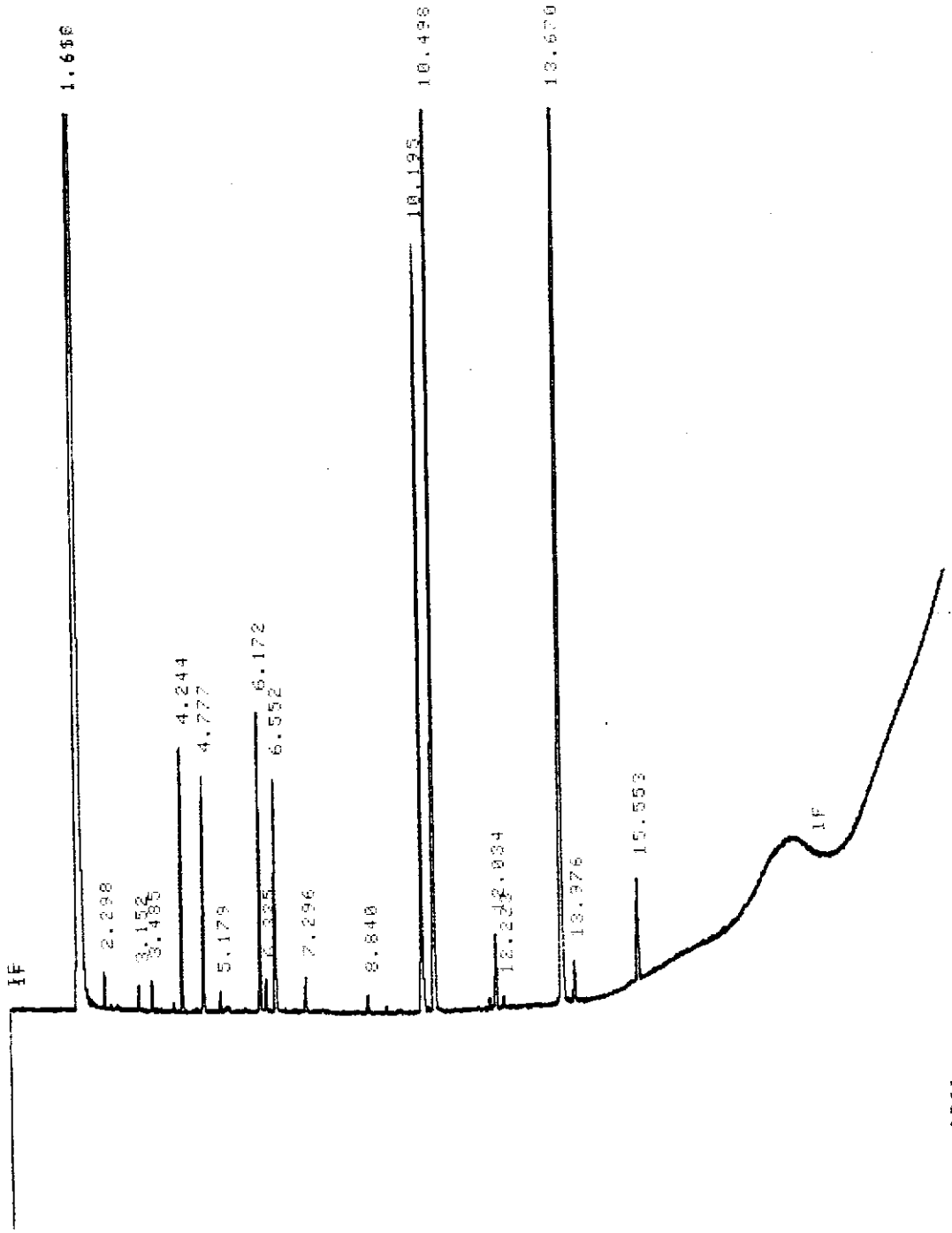
RUN # 8 JUL 17, 1993 05:10:33
START-No plot
END OF SIGNAL

=====

BOTTLE: 8 ID#: 7SAT 17-JUL-93 03:38:17 FILE DATA:F93716849

2312-6

RUN # 9 JUL 17, 1993 05:40:47
START



STOP

Biolog Data
24 Hr Time Point



the **MiL, inc.**

**Interpretation of the Carbon Source
Pattern Recognition Data using a
Multi-well Plate Method (Biolog
Microplate System™) -- Contact Us:
314-878-6626 or Fax 314-878-9376**

The MiL, inc. utilizes the Biolog Microplate System™ for microbial identification and characterization by carbon source pattern recognition. The microplate technique allows for characterization by 95 different tests yielding a potential of 4×10^{28} patterns generated from a single microplate. Each strain of micro-organism yields a distinct pattern and the different species of bacteria will give distinct families of patterns that can be recognized by the Biolog MicroLog™ Software. Microplates are available for Gram Negative (GN), Gram Positive (GP) and E.coli/Salmonella (ES) Analysis. Custom analysis are performed by the MiL, inc. and can be particularly useful in biodegradation or additional selective media development studies. Additional interpretative instructions are provided with such custom services.

To characterize a given microbial isolate the organism is streaked onto a nutrient medium that will support vigorous growth (for example, Nutrient agar, tryptic soy agar or tryptic soy agar supplemented with 5% sheep red blood cells). The more fastidious organisms may require chocolate or BHI agar for growth, whereas many environmental organisms grow better in more minimal media. The culture plates are incubated at 28 to 35° C for 4-18 hours (environmental isolates are typically grown at 28° C with thermophilic strains often incubated at 50° C). After incubation colonies are removed from the culture plate using a saline moistened cotton swab. A suspension of uniform turbidity is prepared in 0.85% saline by comparison with a

known turbidimetric standard. The bacterial suspension is inoculated into the microplate wells (150 µl per well) and the plate covered with the microplate lid. The covered plates are incubated at 28-35° C for 4 hours or overnight (16-24 hours). Should other diluents be requested or used, such changes will be noted.

Microplates may be read at 4 or 24 hours because some organisms give results at 4 hours and may become unreadable at 24 hours. The plates are read in our microplate reader at 590 nm. The absorbance or transmittance (i.e. color) in each well is referenced against the negative control well (A-1) so that any purple color recorded above this control level is read as a positive utilization of the given carbon source. The data are reported as the percent color change as compared to well A-1 utilizing the following formula.

$$\text{Percent color change} = \frac{\text{OD}_{590}(\text{well}) - \text{OD}_{590}(\text{well A-1})}{\text{OD}_{590}(\text{well A-1})}$$

Positive results will be reported in brackets ([]), generally if the Percent Color Change is equal to or greater than 40, the reaction in the given well is considered to be "positive" however the parameters for each substrate may be different and a positive test below a value of 40 is possible. The reported results will be otherwise considered negative. The computer algorithms employed provide standardization of settings ensuring repeatability and avoidance of operator bias. Names of all carbon source substrates employed are provided in the results regardless of response.

We, the MiL's microbiologists, find these methods to be excellent for strain characterization or differentiation between isolates. However, we urge caution in acceptance of the putative identifications to the commercial database and suggest these tests be conducted in conjunction with other methods (we recommend our GC-FAME analyses) when strain identifications are sought.

MICROLOG (TM) 2, RELEASE 3.00

Date : 18/07/83
 Hour : 24
 Plate Type : GN
 Plate # : 1
 Strain Name : 2312-1
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	1	<244>	< 76>	<152>	< 73>	-30	-23	-19	<436-	< 69>	1
B	-25	10	-15	-8	-9	< 47>	-30	-22	-29	<217>	< 71>	{ 21}
C	-22	-16	-21	-25	-32	-20	-30	-25	-23	<355-	<584>	<193>
D	< 95>	<451>	<452>	-26	-20	<112>	<101>	-22	<319>	<174>	<385>	-19
E	-4	<176>	< 65>	<378>	0	<547>	<111>	<141>	< 73-	-41	-84+	-84+
F	<280>	{ 36}	10	< 48>	{ 36}	{ 48+}	1	<278>	<224>	<481>	-17	-23
G	23	<420-	<220>	19	-9	<233>	-17	-20	< 55>	<503>	<147-	< 82>
H	< 39-	19+	-22	-11	-34	<163-	-19	-5	-4+	< 67-	-7	2

BIO-NUMBER : 1706-0106-0007-7156-3570-4534-3117-0104

NO IDENTIFICATION

	CLOSEST SPECIES :	SIM.....	DIST....	AUG.....	MA
X	=> 1) PSEUDOMONAS STUTZERI	0.230	13.644	1.875	7.6
37	2) PSEUDOMONAS MENDOCINA	0.000	15.868	1.750	4.9
38	3) PSEUDOMONAS PUTIDA A	0.000	16.578	1.813	8.3
81	4) ALCALIGENES FAECALIS TYPE II	0.000	16.718	1.938	5.0
94	5) PSEUDOMONAS VIRIBILIVIDA A	0.000	16.931	0.875	3.2
00	6) PSEUDOMONAS FLUORESCENS E	0.000	18.625	1.438	4.7
22	7) ACINETOBACTER CALCOACETICUS/GENOSPECIES1	0.000	18.732	2.438	4.4
69	8) PSEUDOMONAS SOLANACEARUM A	0.000	18.737	1.354	6.8
81	9) PSEUDOMONAS AZELAICA	0.000	19.298	0.781	3.0
84	10) ACINETOBACTER BAUMANNII/GENOSPECIES 2	0.000	19.399	2.000	5.4
75	other :	-----	-----	-----	---
--					

ABBREVIATED NAME : PSD.STU
 FULL NAME : PSEUDOMONAS STUTZERI
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

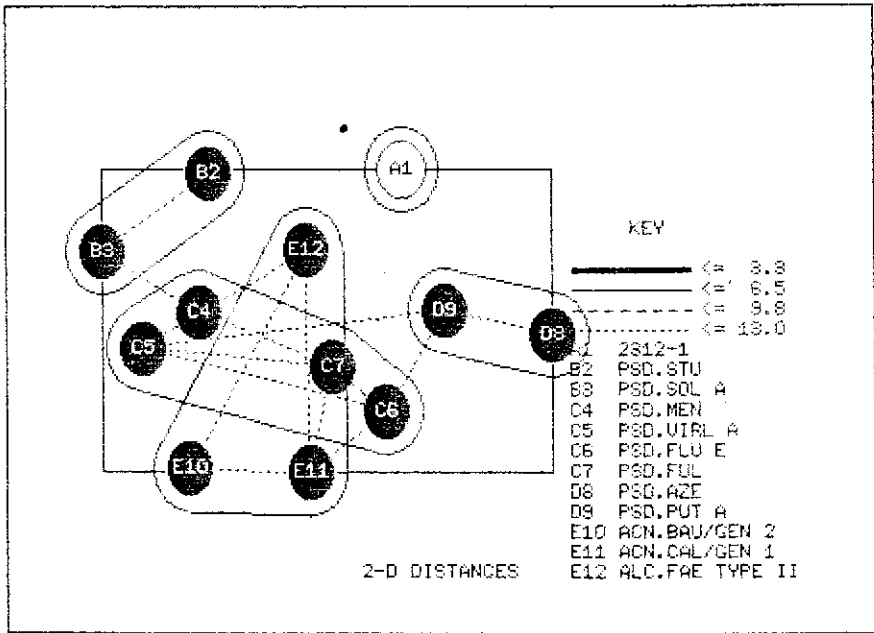
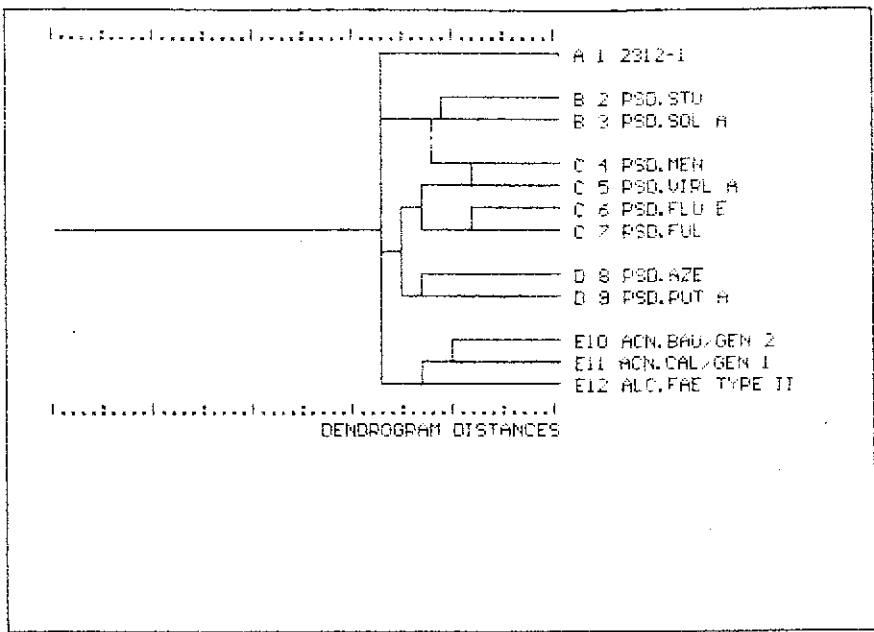
	1	2	3	4	5	6	7	8	9	10	11	12
A	0	24	34	41	36	36	0	0	0	0	9	9
B	16	72	0	0	14	86	0	9	0	36	36	57
C	0	9	59	7	9	41	14	0	19	0	60	24
D	74	53	43	16	0	9	29	0	0	45	91	36
E	0	45	79	57	16	79	57	36	0	17	0	79
F	71	43	9	43	45	41	7	86	48	79	0	31
G	0	0	93	14	24	53	45	0	16	38	0	21
H	21	55	9	0	10	0	19	21	48	0	0	0

24 HOUR DATA :

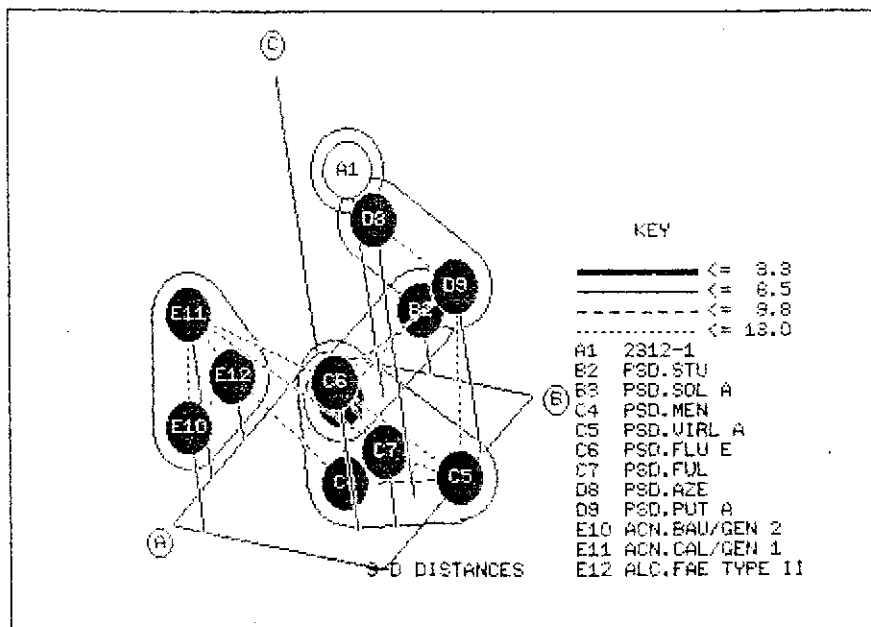
	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	100	88	100	100	0	0	0	0	36	0
B	0	81	0	0	7	100	0	0	0	93	76	66
C	0	0	69	0	0	16	0	12	0	0	100	100
D	83	100	90	57	0	50	83	0	59	81	100	24
E	0	100	81	100	62	100	100	84	0	57	91	100
F	100	83	47	83	86	100	55	100	79	100	0	0
G	0	0	100	0	12	100	81	0	17	40	0	29
H	7	100	0	0	14	0	22	9	100	0	0	0

CLOSEST SPECIES :

- 1) 11.288 : PSEUDOMONAS SOLANACEARUM A
- 2) 13.499 : PSEUDOMONAS ALCALIGENES B
- 3) 13.765 : CDC GROUP IVC-2
- 4) 13.828 : ALCALIGENES FAECALIS TYPE II
- 5) 13.911 : PSEUDOMONAS MENDOCINA



- A1 2312-1
- B2 PSD.STU
- B3 PSD.SOL A
- C4 PSD.MEN
- C5 PSD.UIRL A
- C6 PSD.FLU E
- C7 PSD.FUL
- D8 PSD.AZE
- D9 PSD.PUT A
- E10 ACN.BAU/GEN 2
- E11 ACN.CAL/GEN 1
- E12 ALC.FAE TYPE II



Date : 18/07/93
 Hour : 24
 Plate Type : GN
 Plate # : 2
 Strain Name : 2312-2
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	-10	-10	-4	<342>	<196>	-15	-15	-11	-4	-14	-9
B	-8	-8	-3	-11	-14	-5	-5	-2	-8	-11	-13	-15
C	-5	0	-9	-9	-11	-9	-10	-19	-13	-2	<469>	<240>
D	<220>	<207>	{ 82+}	-3	-11	-4	-14	-10	-15	<242>	<570>	-9
E	{ 7-	0	<189>	-19	<269>	<696>	-14	<379>	<580-	-3	-6	<672>
F	<307>	{ 39}	-9	{124+}	{114+}	<221>	{ 31+}	<371>	<368>	<704>	-9	-6
G	<402>	<552>	<224>	<204>	{ 39}	<300>	-6	-8	<258>	{ 42}	-4	<314>
H	-1	-9	-9	-10	-12	<233>	-11	{ 4-	-7	-12	-7	-9

BIO-NUMBER : 0300-0000-0003-7006-1331-4734-7511-0100

SPECIES IDENTIFICATION : PSEUDOMONAS ALCALIGENES A

	CLOSEST SPECIES	SIM.	DIST.	AVG.	MA
X =>	1) PSEUDOMONAS ALCALIGENES A	0.904	1.591	1.125	4.5
50	2) ALCALIGENES FAECALIS TYPE II	0.004	3.423	1.938	5.0
94	3) AQUASPIRILLUM PUTRIDICONCHYLIIUM	0.000	4.481	0.313	1.1
06	4) AQUASPIRILLUM DISPAR	0.000	7.000	0.083	0.4
75	5) PSEUDOMONAS PSEUDOALCALIGENES	0.000	7.373	1.750	7.2
13	6) ACINETOBACTER CAL. BV ALCALIGENES	0.000	7.446	0.250	3.3
13	7) ACINETOBACTER GENOSPECIES 15	0.000	7.635	1.250	10.7
12	8) ACINETOBACTER CALCOACETICUS/GENOSPECIES1	0.000	7.946	2.438	4.4
69	9) ACINETOBACTER JUNII/GENOSPECIES 5	0.000	8.505	1.000	5.2
13	10) ACINETOBACTER SPECIES GROUP B3	0.000	8.616	0.234	4.6
50	other :	-----	-----	-----	---
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ABBREVIATED NAME : PSD.ALC A
 FULL NAME : PSEUDOMONAS ALCALIGENES A
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

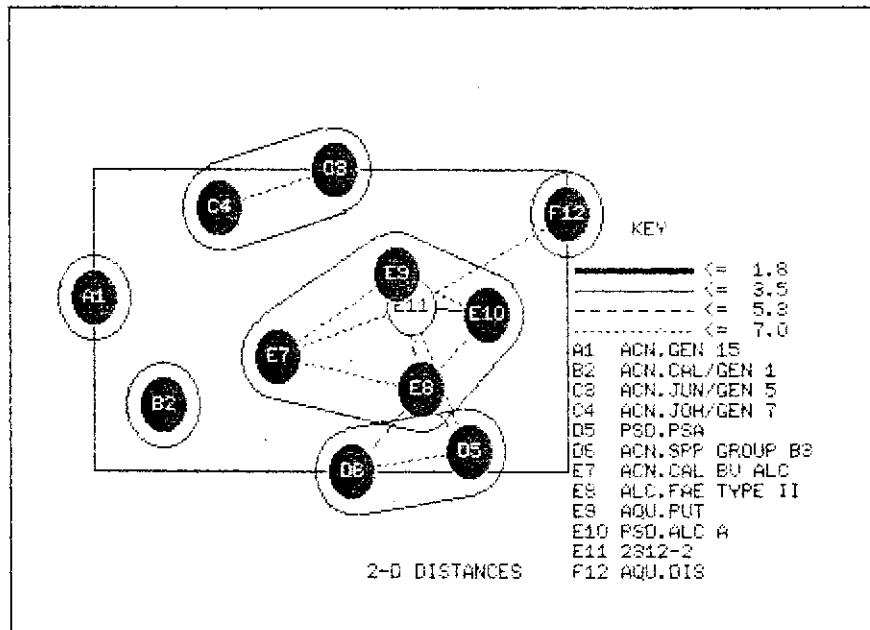
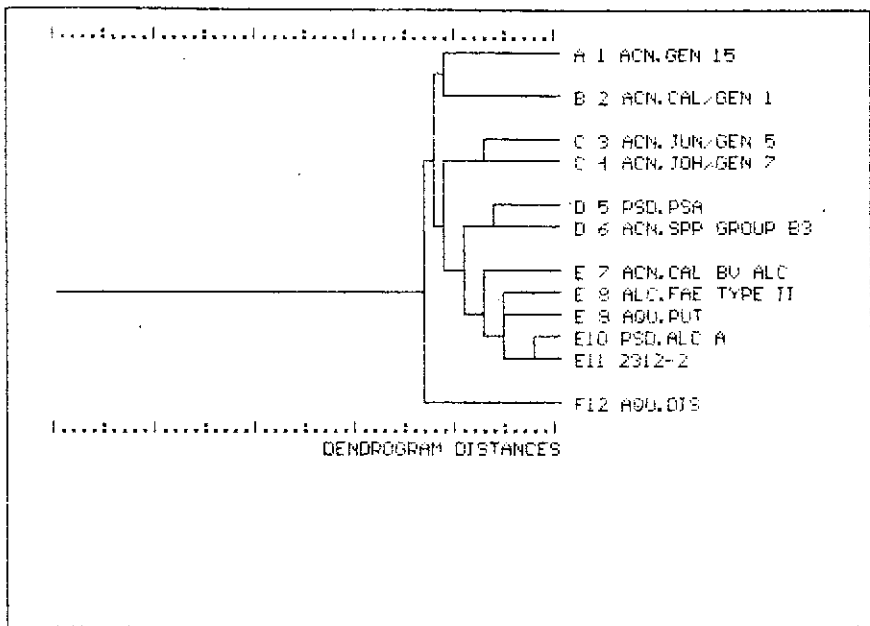
	1	2	3	4	5	6	7	8	9	10	11	12
A	0	17	0	0	100	100	0	0	0	0	0	17
B	0	0	0	0	0	0	0	0	0	0	0	0
C	0	0	0	0	0	0	8	0	0	0	92	0
D	83	67	58	8	0	0	0	0	0	58	58	33
E	0	0	75	8	75	100	0	67	0	0	0	83
F	67	67	0	67	75	83	83	92	92	100	17	83
G	42	67	100	67	42	92	25	50	100	58	0	67
H	50	33	0	8	0	75	8	0	8	0	0	17

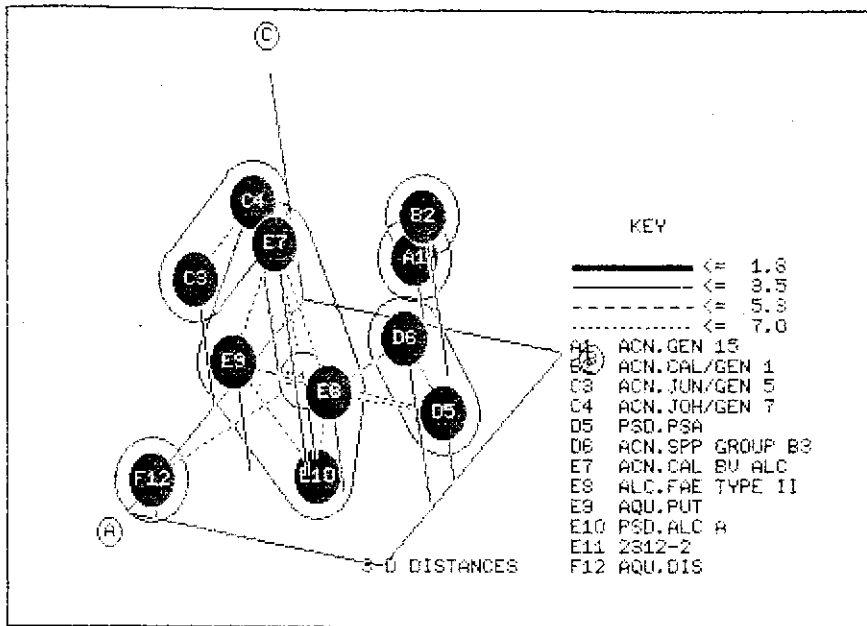
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	0	0	100	100	0	0	0	0	0	0
B	0	0	0	0	0	0	0	0	0	0	8	0
C	0	0	0	0	0	0	0	0	0	0	100	33
D	92	100	92	17	0	0	0	0	0	100	83	17
E	0	0	100	25	100	100	0	100	0	0	8	83
F	100	83	0	92	100	100	100	100	100	100	8	67
G	75	75	100	83	67	100	33	58	100	83	0	83
H	50	33	0	8	0	100	8	0	17	0	0	8

CLOSEST SPECIES :

- 1) 5.776 : AQUASPIRILLUM PUTRIDICONCHYLUM
- 2) 6.123 : ALCALIGENES FAECALIS TYPE II
- 3) 8.317 : AQUASPIRILLUM BENGAL
- 4) 8.476 : BORDETELLA AVIUM
- 5) 8.553 : PSEUDOMONAS PSEUDOALCALIGENES





MICROLOG (TM) 2, RELEASE 3.00

Date : 18/07/93
 Hour : 24
 Plate Type : GN
 Plate # : 3
 Strain Name : 2312-3
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus AI control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	10	{ 49}	27	<170>	< 77>	-9	<184>	-4	<436>	<201>	9
B	7	<256>	-1	<330>	< 94>	<379>	-21	6	-3	< 79>	<258>	<237>
C	< 69>	8	< 92>	3	-3	12	-4	<173>	5	2	<116>	<194>
D	<242>	<511>	<494>	<122>	<207>	< 75>	<438>	<204>	21	<131>	<340>	-10
E	4	11	<111>	<407>	<122>	<421>	<142>	<245>	<304>	<482>	{ 56}	<326>
F	<160>	<158>	<143>	<125>	<110>	<134>	<145>	<263>	<200>	<322>	12	<209>
G	< 83>	<343>	<198>	< 95>	< 91>	<232>	<316>	< 82>	<197>	<116>	<114>	<259>
H	<299>	<331>	<121>	8	1	<135>	<100>	{ 18-	<257>	< 90>	12	0

BIO-NUMBER : 0326-2707-5023-7766-1777-7775-7777-7154

SPECIES IDENTIFICATION : PSEUDOMONAS FLUORESCENS B

	CLOSEST SPECIES	SIM	DIST	AVG	MA
X=>	1) PSEUDOMONAS FLUORESCENS B	0.889	1.307	2.250	5.7
87	2) PSEUDOMONAS FUSCOVAGINAE	0.000	7.703	0.688	3.1
62	3) PSEUDOMONAS CORRUGATA	0.000	8.279	1.703	4.4
56	4) PSEUDOMONAS PUTIDA A	0.000	8.994	1.813	8.3
81	5) PSEUDOMONAS FRAGI	0.000	9.240	2.500	9.0
00	6) PSEUDOMONAS FLUORESCENS C	0.000	9.755	2.000	4.6
50	7) PSEUDOMONAS PUTIDA B	0.000	9.900	2.188	8.2
50	8) PSEUDOMONAS FLUORESCENS A	0.000	9.919	1.625	4.6
13	9) FLAVIMONAS ORYZIHABITANS	0.000	9.935	2.203	4.8
31	10) PSEUDOMONAS GLADIOLI	0.000	10.144	1.375	6.6
62	other :	-----	-----	-----	---
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ABBREVIATED NAME : PSD.FLU B
 FULL NAME : PSEUDOMONAS FLUORESCENS B
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

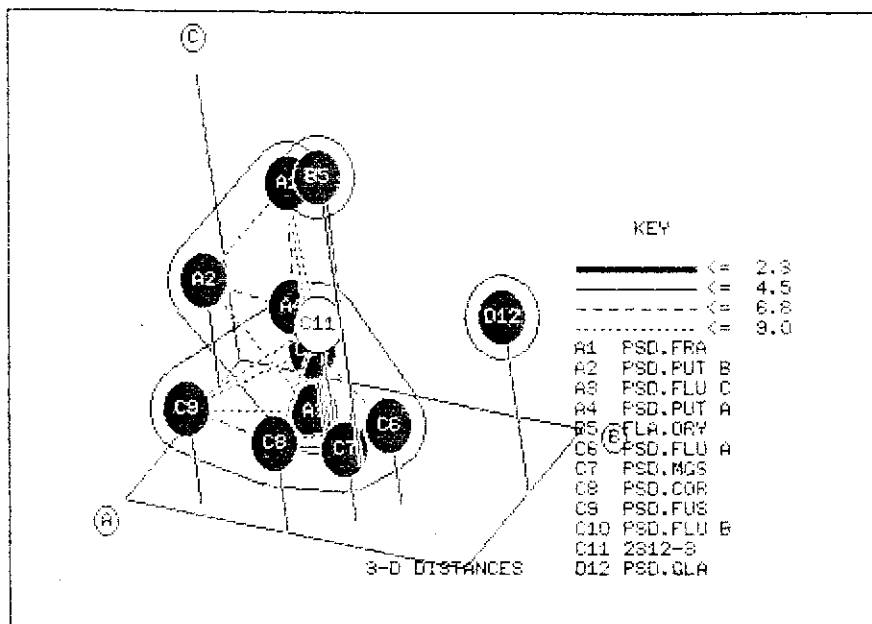
	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	14	7	77	46	0	38	0	85	62	0
B	0	77	0	77	31	92	0	0	7	15	69	77
C	15	0	69	0	0	7	46	54	7	0	15	38
D	79	77	85	54	77	8	100	54	14	23	77	0
E	7	0	8	85	15	85	31	77	54	92	15	100
F	85	8	15	8	23	62	62	100	77	100	23	100
G	8	46	79	38	23	54	54	31	92	62	15	85
H	31	77	38	7	0	15	54	0	62	23	0	7

24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	14	7	100	100	0	86	0	100	93	0
B	0	100	0	100	43	100	0	0	7	86	100	100
C	29	0	100	7	0	7	64	86	7	0	100	79
D	79	100	100	79	100	14	100	93	14	64	100	50
E	7	0	57	100	57	100	100	100	100	100	29	100
F	100	100	64	100	100	100	100	100	100	100	29	100
G	100	50	79	64	43	100	100	64	100	100	93	100
H	100	100	93	7	7	71	93	0	100	86	7	7

CLOSEST SPECIES :

- 1) 6.137 : PSEUDOMONAS CORRUGATA
- 2) 6.730 : PSEUDOMONAS AURANTIACA
- 3) 7.157 : PSEUDOMONAS FUSCOVAGINAE
- 4) 7.739 : PSEUDOMONAS FLUORESCENS A
- 5) 7.791 : PSEUDOMONAS PUTIDA A



MICROLOG (TM) 2, RELEASE 3.00

Date : 18/07/93
 Hour : 24
 Plate Type : GN
 Plate # : 4
 Strain Name : 2312-4
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	< 52-	<104>	< 94>	43+	14+	-47	-40	-35	-49	-58	-40
B	-49	-26	-45	< 29-	-42	<129>	-39	-9	-20	< 49>	-50	-28
C	-32	-32	-30	-28	-36	-36	-50	-6	-26	-50	<139>	< 74>
D	<118>	<108>	< 99>	2	-20	<111>	< 80>	-22	<109>	< 86>	<131>	-11
E	-62	< 41>	<116>	< 83>	< 90>	<151>	< 97>	<122>	-11	<114>	-7+	<123>
F	<100>	-42	10	-18	23	< 40+	1	<120>	< 70>	<103>	-43	-10
G	-55	-49	<100>	-29	-31	< 66>	-41	-46	< 87>	< 46>	-54	17
H	-44	<110>	-27	-54	-62	-58	-19	-4	< 39+	-54	-47	-46

BIO-NUMBER : 3400-0104-0003-7156-3765-4034-1114-2000
 FALSE POSITIVE WARNING : High optical density in A1 control well. True positives could be under-reported. Check inoculation procedures.
 SPECIES IDENTIFICATION : PSEUDOMONAS STUTZERI

	CLOSEST SPECIES :	SIM.....	DIST....	AUG.....	MA
X	=> 1) PSEUDOMONAS STUTZERI	0.510	8.193	1.875	7.6
37	2) ALCALIGENES FAECALIS TYPE II	0.000	13.531	1.938	5.0
94	3) PSEUDOMONAS MENDOCINA	0.000	14.398	1.750	4.9
38	4) NEISSERIA DENITRIFICANS	0.000	14.803	1.531	11.4
06	5) PSEUDOMONAS SOLANACEARUM A	0.000	15.321	1.354	6.8
81	6) PSEUDOMONAS VESICULARIS	0.000	15.334	2.438	4.7
75	7) GILARDI PINK GRAM NEGATIVE ROD	0.000	15.620	2.813	9.5
06	8) OLIGELLA UREOLYTICA/URETHRALIS	0.000	15.828	1.813	5.5
06	9) NEISSERIA ANIMALIS	0.000	15.865	0.563	6.6
44	10) COMAMONAS TERRIGENA	0.000	16.910	1.938	8.3
44	other :	-----	-----	-----	---
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ABBREVIATED NAME : PSD.STU
 FULL NAME : PSEUDOMONAS STUTZERI
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

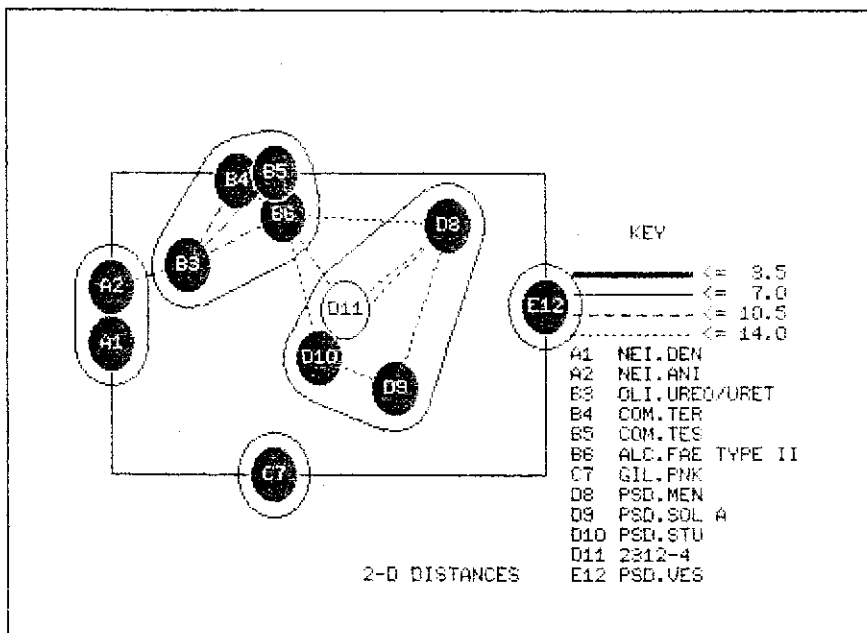
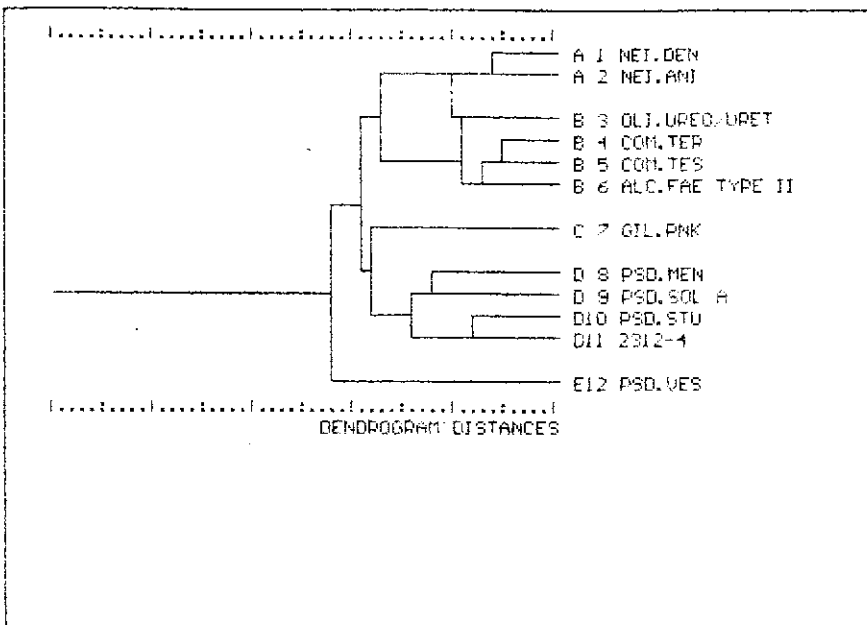
	1	2	3	4	5	6	7	8	9	10	11	12
A	0	24	34	41	36	36	0	0	0	0	9	9
B	16	72	0	0	14	86	0	9	0	36	36	57
C	0	9	59	7	9	41	14	0	19	0	60	24
D	74	53	43	16	0	9	29	0	0	45	91	36
E	0	45	79	57	16	79	57	36	0	17	0	79
F	71	43	9	43	45	41	7	86	48	79	0	31
G	0	0	93	14	24	53	45	0	16	38	0	21
H	21	55	9	0	10	0	19	21	48	0	0	0

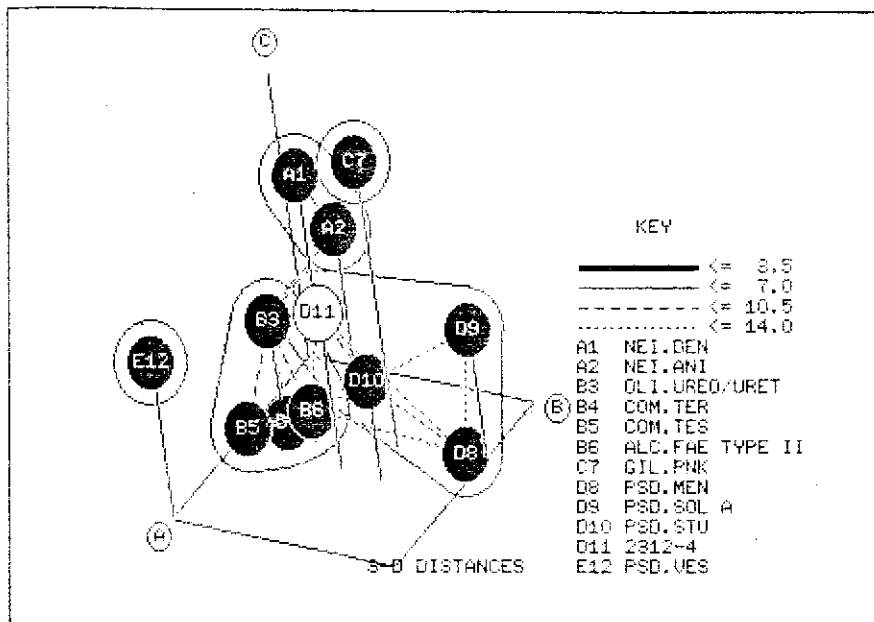
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	100	88	100	100	0	0	0	0	36	0
B	0	81	0	0	7	100	0	0	0	93	76	66
C	0	0	69	0	0	16	0	12	0	0	100	100
D	83	100	90	57	0	50	83	0	59	81	100	24
E	0	100	81	100	62	100	100	84	0	57	91	100
F	100	83	47	83	86	100	55	100	79	100	0	0
G	0	0	100	0	12	100	81	0	17	40	0	29
H	7	100	0	0	14	0	22	9	100	0	0	0

CLOSEST SPECIES :

- 1) 11.288 : PSEUDOMONAS SOLANACEARUM A
- 2) 13.499 : PSEUDOMONAS ALCALIGENES B
- 3) 13.765 : CDC GROUP IVC-2
- 4) 13.828 : ALCALIGENES FAECALIS TYPE II
- 5) 13.911 : PSEUDOMONAS MENDOCINA





MICROLOG (TM) 2, RELEASE 3.00

Date : 18/07/93
 Hour : 24
 Plate Type : GN
 Plate # : 5
 Strain Name : 2312-5
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" ID choice positive > 90% of time
 XXX- = data positive or borderline, "=" ID choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	-48	{ 21+ < 29>	18	-3	< 43>	< 32>	< 49-	-66	-42	-57	
B	-65	-44	-68	-56	-46	< 41>	-63	< 48>	-54	3+	-53+	-50+
C	-64	-66	-61	-65	-23	-59	-58	-37+	< 41>	8	< 45>	7
D	-4	< 28-	< 33-	-19	-65	< 38-	< 62>	-54	< 36>	5	< 40-	-59
E	-69	-7	< 17-	{ 22}	< 22-	< 43>	< 17-	9	-26	-5	-47	< 40>
F	< 23>	-55	-49	-40	-9	-22	-34	< 50>	< 35>	< 31>	-66+	-30
G	-78	-74	< 14-	-67	-48	-19	-63	-33	8	-19	-67	-43
H	< 27-	< 32>	-67	-65	-73	-68	-55	-23	< 34>	< 39>	-51	-61

BIO-NUMBER : 0470-0120-0012-3152-1341-4034-1000-6014
 FALSE POSITIVE WARNING : High optical density in A1 control well. True positives could be under-reported. Check inoculation procedures.
 NO IDENTIFICATION

	CLOSEST SPECIES :	SIM.....	DIST....	AUG.....	MA
X	=> 1) VIBRIO MIMICUS	0.056	17.889	2.313	4.1
09	2) DELEYA AESTA	0.036	18.045	2.000	4.8
13	3) PSYCHROBACTER IMMOBILIS	0.030	18.117	2.250	5.1
62	4) METHYLOBACTERIUM MESOPHILICUM	0.001	19.072	2.125	5.3
87	5) CDC GROUP EF-4	0.000	19.928	1.625	4.9
25	6) PSEUDOMONAS STUTZERI	0.000	19.963	1.875	7.6
37	7) PLESIOMONAS SHIGELLOIDES	0.000	20.071	2.031	6.4
19	8) MORAXELLA BOVIS	0.000	20.111	1.625	5.8
25	9) HYDROGENOPHAGA PALLERONII	0.000	20.868	1.125	6.3
13	10) SHEWANELLA PUTREFACIENS A	0.000	21.169	2.219	5.5
31	other :	-----	-----	-----	---
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ABBREVIATED NAME : VIB.MIM
 FULL NAME : VIBRIO MIMICUS
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

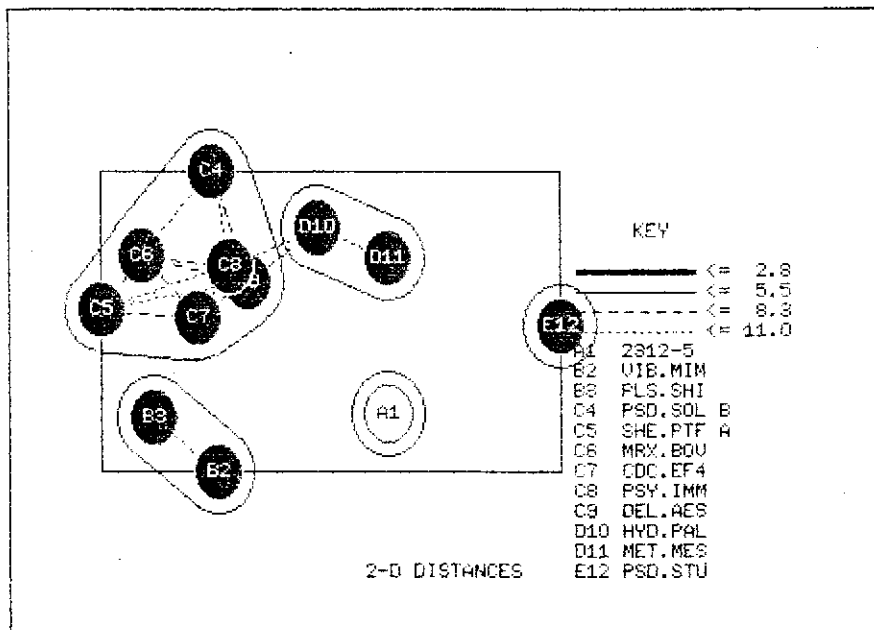
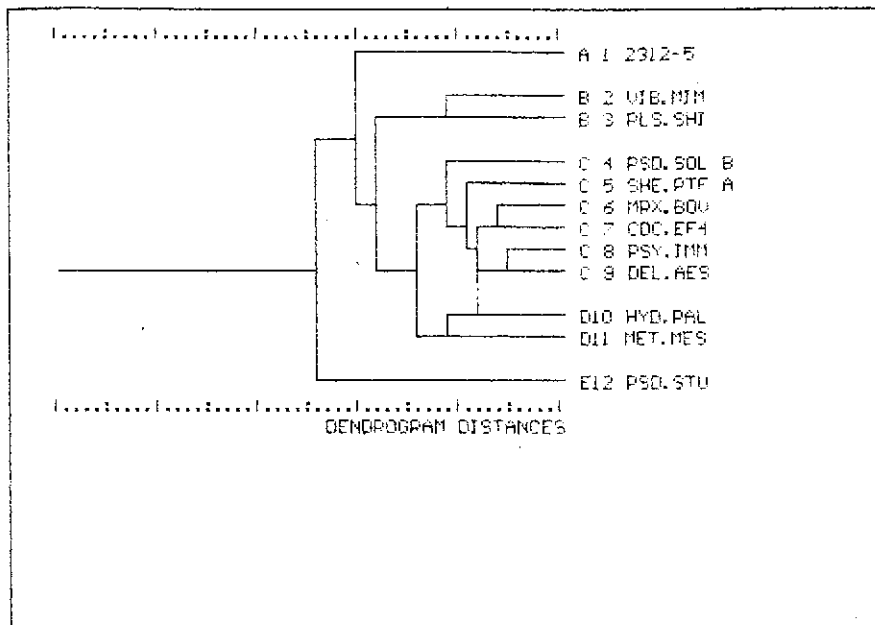
	1	2	3	4	5	6	7	8	9	10	11	12
A :	0	36	84	58	44	28	12	100	0	0	0	0
B :	12	72	0	16	52	100	0	12	16	76	76	60
C :	0	32	72	8	0	64	0	76	40	0	88	44
D :	40	0	0	0	0	8	52	0	0	0	0	0
E :	0	0	0	20	0	92	0	0	0	0	0	80
F :	20	24	0	8	0	8	12	56	20	44	36	60
G :	0	0	0	0	0	36	0	0	16	0	0	0
H :	0	92	52	56	8	0	0	0	48	12	12	0

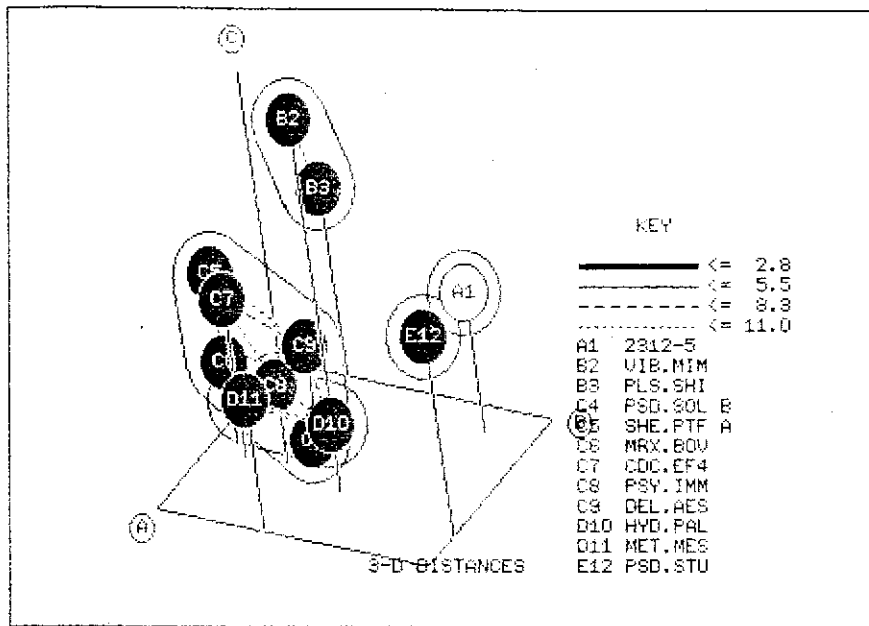
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A :	0	0	100	100	52	52	64	100	0	0	0	0
B :	0	80	0	28	36	100	0	92	12	100	100	100
C :	0	48	88	0	0	68	0	100	20	0	100	56
D :	28	0	8	0	0	0	100	0	72	0	0	0
E :	0	0	0	16	0	100	0	0	0	0	0	88
F :	16	24	24	0	0	0	16	88	76	72	92	72
G :	0	0	0	0	0	40	0	0	12	0	0	0
H :	8	100	64	68	0	0	0	0	48	48	0	0

CLOSEST SPECIES :

- 1) 6.086 : VIBRIO CHOLERAЕ
- 2) 7.865 : VIBRIO SPLENDIDUS 2
- 3) 8.549 : VIBRIO VULNIFICUS
- 4) 9.178 : VIBRIO DAMSELA
- 5) 9.313 : AEROMONAS DNA GROUP 11





MICROLOG (TM) 2, RELEASE 3.00

Date : 18/07/93
 Hour : 24
 Plate Type : 6N
 Plate # : 6
 Strain Name : 2312-6
 Strain # : W.L00
 Other Info : ?
 Input Mode : Reader : BIOLOG MICROSTATION
 Data Base : MicroLog GN

POSITIVE/NEGATIVE DATA

XXX = percent change in optical density versus A1 control well
 <XXX> = positive, {XXX} = borderline, XXX = negative
 -XXX = percent change negative
 XXX+ = data negative or borderline, "=" IO choice positive > 90% of time
 XXX- = data positive or borderline, "=" IO choice positive < 10% of time

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	15	63	42	<187>	<124>	-8	<196>	-15	56	30	-11
B	0	<125>	29	< 86-	48	<287>	0	10	2	< 89>	<162>	36
C	12	46	< 84>	21	19	20	5	<141-	13	2	<164>	<159>
D	<254>	<309>	<305>	<180>	20	17	<309>	2	-5	<133>	<267>	-11
E	<322>	<260>	<159>	<305>	<176>	<216>	<126>	<243>	<159>	-3	< 98>	<251>
F	<246>	<179>	< 77-	<162>	<159>	<172>	<104>	<312>	<228>	<233>	17	13
G	<265>	<170>	<145>	<138>	< 99>	<150>	<298>	< 89>	<167>	<118>	{ 64+	<188>
H	<250>	<309>	48	3	-12	<256>	<282>	<111>	<237>	-3	-13	-25

BIO-NUMBER : 0320-2506-1023-7446-7773-7774-7777-6170

SPECIES IDENTIFICATION : PSEUDOMONAS AERUGINOSA

	CLOSEST SPECIES	SIM.	DIST.	AUG.	MA
X =>	1) PSEUDOMONAS AERUGINOSA	0.621	5.490	1.297	4.1
94	2) PSEUDOMONAS FLUORESCENS C	0.003	7.193	2.000	4.6
50	3) PSEUDOMONAS PUTIDA A	0.001	7.649	1.913	8.3
81	4) PSEUDOMONAS FRAGI	0.000	10.491	2.500	9.0
00	5) PSEUDOMONAS PUTIDA B	0.000	11.176	2.188	8.2
50	6) PSEUDOMONAS FLUORESCENS B	0.000	11.726	2.250	5.7
87	7) PSEUDOMONAS CITRONELLOLIS	0.000	12.058	0.234	2.9
31	8) PSEUDOMONAS SOLANACEARUM A	0.000	12.838	1.354	6.9
81	9) PSEUDOMONAS AURANTIACA	0.000	13.785	0.719	5.0
19	10) ALCALIGENES FAECALIS TYPE II	0.000	14.055	1.938	5.0
94	other :	-----	-----	-----	---
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ABBREVIATED NAME : PSD.AER
 FULL NAME : PSEUDOMONAS AERUGINOSA
 DATA BASE CATEGORY : CLINICAL

4 HOUR DATA :

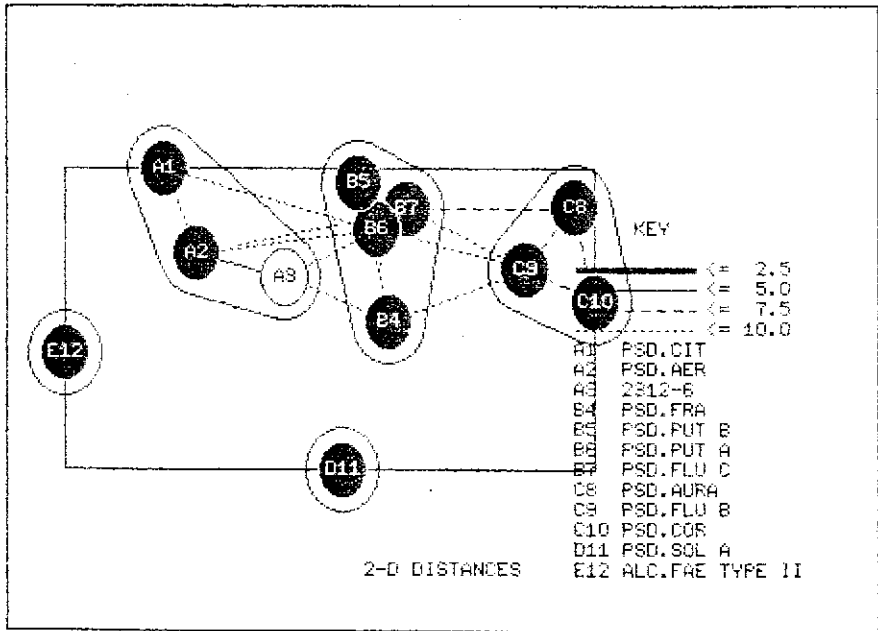
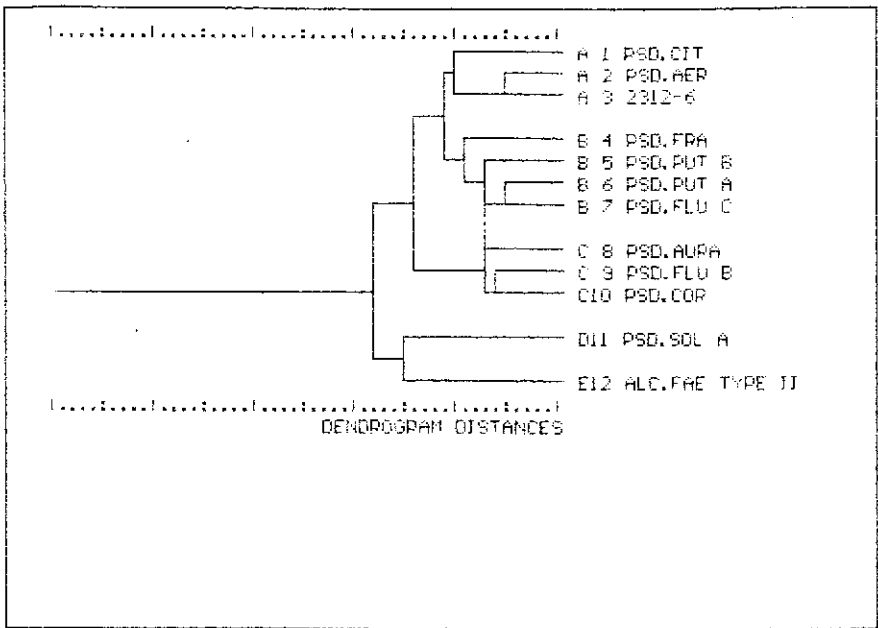
	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	0	0	35	22	0	27	0	0	0	0
B	0	31	0	0	0	65	0	0	0	0	0	10
C	0	0	22	0	0	0	0	0	0	0	14	27
D	57	45	51	24	0	0	45	0	0	20	47	0
E	14	35	35	53	20	51	29	37	0	0	0	76
F	61	33	0	42	57	51	24	57	47	63	0	12
G	43	29	47	37	12	24	43	24	51	39	18	57
H	71	57	8	0	0	27	61	8	43	0	0	0

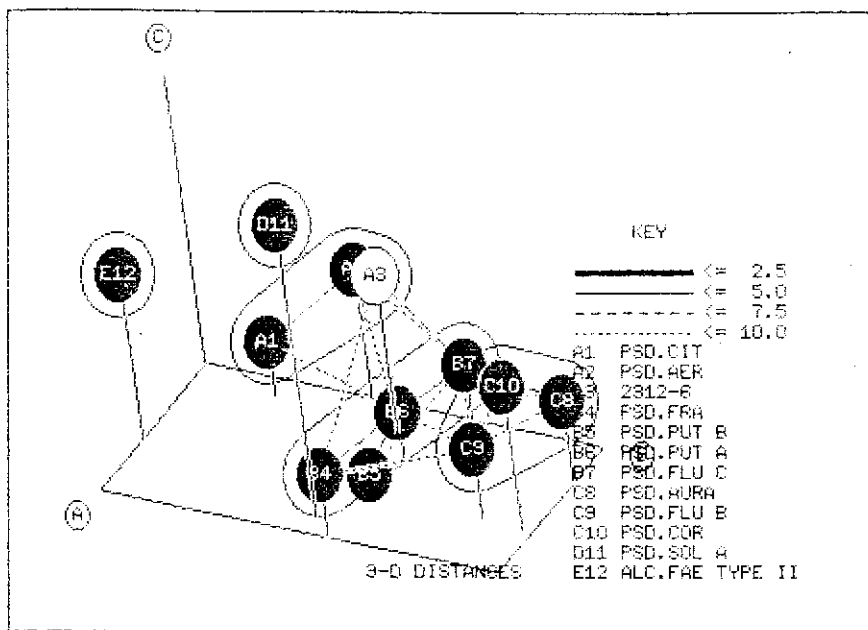
24 HOUR DATA :

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	0	0	0	88	91	0	94	0	0	10	0
B	0	100	0	0	0	100	0	0	0	15	100	10
C	0	0	28	0	0	0	0	0	0	0	100	94
D	57	100	100	97	0	0	100	0	0	49	100	0
E	93	100	75	100	72	100	100	85	100	0	85	100
F	100	100	0	42	93	100	51	100	100	100	0	12
G	100	100	85	100	12	100	100	24	100	45	100	100
H	100	100	27	0	0	100	100	28	100	0	0	0

CLOSEST SPECIES :

- 1) 7.599 : PSEUDOMONAS CITRONELLIS
- 2) 9.463 : PSEUDOMONAS FLUORESCENS C
- 3) 9.782 : PSEUDOMONAS PUTIDA A
- 4) 12.051 : PSEUDOMONAS FLUORESCENS E
- 5) 12.085 : PSEUDOMONAS PUTIDA B





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